

PLC MARK TEN

EFFECTIVE ENVIRONMENTAL SANITATION -

A disinfectant/deodorizer/sanitizer when used on previously cleaned environmental hard surfaces.* Sanitizes at 1/4 ounce per gallon of water and disinfects at 3 1/2 ounces per 5 gallons of water. Formulated for use in Food Processing Plants, Restaurants, Bars, Hospitals, Nursing Homes, and Institutions.

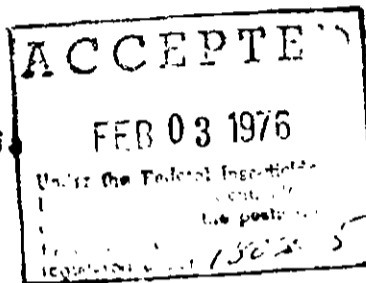
- DISINFECTANT
 - STAPHYLOCIDAL - *Staphylococcus aureus*
 - PSEUDOMONACIDAL - *Pseudomonas aeruginosa*
 - BACTERICIDAL - *Salmonella choleraesuis*
- DEODORIZER - 2 stage
 - Bactericidal - kills odor causing bacteria
 - Neutralizes odor particles
- SANITIZER
 - Staphylococcus aureus*
 - Escherichia coli*
- FUNGICIDAL - *Trichophyton interdigitale* - athlete's foot fungus
- VIRUCIDAL
 - Influenza A₂* - Hong Kong, causative virus of flu
 - Herpes Simplex* - causative virus of cold sores, fever blisters
 - Adenovirus type 5* - causative virus of acute respiratory disease
 - Vaccinia Virus* - causative virus of small pox
- PHENOL COEFFICIENTS - Guaranteed minimum
 - Staphylococcus aureus* 92
 - Salmonella typhosa* 60
- NO RINSE - Authorized by FDA for a No Rinse Claim when used for sanitization of food processing and food contact items at 200 ppm. of active quaternaries.
- Authorized by USDA for use in federally inspected meat and poultry plants as a sanitizing solution.

*Supportive data on file with the Environmental Protection Agency, Washington, D. C.

EPA Reg. No. 18035-5

EPA Est. 18035-MN-1

PRIVATE LABEL CHEMICALS, INC.
2280 TERMINAL ROAD
ST. PAUL, MINN. 55113



SUPPORTING DATA

PROCEDURE

Quaternary ammonium chloride was prepared as follows:

1. 40% (12% 30% (16% 5% (12% 4% (16%) quaternary ammonium chlorides	25.0
2. 40% (12% 32% (14%) dimethyl ethyl benzyl ammonium chlorides	25.0
Total ingredients	50.0
	100.0

This quaternary ammonium chloride product was made up as separate concrete preparations in the following instances:

- #69-315
- #69-B-13
- #69-B-24

These formulations were tested for bactericidal and fungicidal activity by the appropriate AOAC test procedures.

The test data obtained are summarized as follows.

Bactericidal

- (1) At 400 ppm, active against *Staphylococcus aureus* and *Escherichia coli*; at 450 ppm, active against *Salmonella typhimurium*; at 550 ppm, active against *Escherichia coli* by the Use Dilution method.
- (2) A Guaranteed Minimum phenol coefficient of 526 and 604 against *Staphylococcus aureus* and *Escherichia coli*, respectively, on a 100% activity basis.
- (3) Sanitizing activity at 200 ppm, active, with a hard water tolerance of 750 ppm, synthetic hard water (as CaCO₃) for *Staphylococcus aureus* and *Escherichia coli*.

Fungicidal

Fungicidal activity at 1/800 aqueous dilution (1250 ppm) against *Aspergillus niger* and *Penicillium notatum* on a 100% activity basis.

FUNGICIDAL EFFICACY

Fungicidal by the AOAC Fungicidal Test

highest aqueous dilution of germicide (on a 100% activity basis) which kills *Aspergillus niger* and *Penicillium notatum* in 10 minutes, but not in 5 minutes.

formulation	dilution	kill (-)/no kill (+) after (min.)		
		5	10	15
#69-315 (a)	1/250	-	-	-
	1/400	-	-	-
	1/500	+	-	-
	1/600	+	-	-
	1/700	+	-	-
	1/800	+	-	-
	1/900	+	-	-
phenol resistance	1/60	+	-	-
	1/70	+	+	+
#69-315 (b)	1/600	+	-	-
	1/700	+	-	-
	1/750	+	-	-
	1/800	+	-	-
	1/850	+	-	-
	1/900	+	-	-
	1/950	+	-	-
phenol resistance	1/60	+	-	-
	1/70	+	+	+

Azolectin/Tween 80 dextrose broth used as subculture medium.

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BACTERICIDAL EFFICACY

Bactericidal by the AOAC Use Dilution Method

* negative subcultures/10 replicate stainless steel carriers inoculated with *Staphylococcus aureus* or *Escherichia coli* and treated with germicide at several aqueous dilutions for 10 minutes.

rate	formulation	phenol coefficient	# negative subcultures/10 replicates		
			SA	SC	PA
1/8	#69-315	427		10/10, 10/10, 10/10	
1/9		427	10/10, 10/10, 10/10		
1/10		427	10/10, 10/10, 10/10		
1/12	#69-315	427		30/30	10/10, 10/10, 10/10
1/14		427	10/10, 10/10, 10/10		
1/16		427	10/10, 10/10, 10/10		
1/18	#69-315	427		30/30	10/10, 10/10, 10/10
1/20		427	10/10, 10/10, 10/10		
1/22		427	10/10, 10/10, 10/10		
1/24	#69-315	427		30/30	10/10, 10/10, 10/10
1/26		427	10/10, 10/10, 10/10		
1/28		427	10/10, 10/10, 10/10		
1/30	#69-315	427		30/30	10/10, 10/10, 10/10
1/32		427	10/10, 10/10, 10/10		
1/34		427	10/10, 10/10, 10/10		

ADENOVIRUS TYPE 5

Procedure. The virus was grown in HeLa cell cultures and had a titer of 10^6 infectious doses per 0.1 ml after 8 days incubation at 37°C. For the test, 1 ml of virus was added to 9 ml of the germicide. After 10 minutes contact at room temperature the virus-germicide mixture was diluted in broth to the titer of the virus and inactivation was determined by comparing the titer of the treated virus with that of the untreated virus. Viral control. Saline was substituted for the germicide. The presence of active virus was determined by the inoculation of 0.1 ml of the test preparations into each of four cell culture tubes. Cells were incubated for 8 days and observed for cytopathogenic effect.

Dilution of virus-germicide or virus-saline mixture.	Results	
	Control Virus-Saline	Test Virus-Germicide
10 ⁻¹	++++	TTTT
10 ⁻²	++++	++++
10 ⁻³	++++	0000
10 ⁻⁴	++++	0000
10 ⁻⁵	++++	0000
10 ⁻⁶	++++	0000
10 ⁻⁷	0000	0000
10 ⁻⁸	0000	0000

+ = Virus present
 0 = Virus absent
 T = Toxicity due to germicide. Presence or absence of virus could not be determined.

Summary. Under the above test conditions the compound inactivated four logs of virus.

INFLUENZA A₂

Procedure. The virus was grown in the allantoic sac of 10 day old chick embryos. The virus was harvested after 40 hours incubation at 37°C and had an infectivity titer of 10^6 egg infectious doses per 0.1 ml of allantoic fluid, as determined by hemagglutination of a 0.5% suspension of chicken red blood cells. The virus diluted 1:5 in saline was mixed with 9 parts of the test dilution of the germicide. After 10 minutes contact at room temperature, ten-fold serial dilutions were prepared in broth and 1:1 ml of the virus-germicide (or virus-saline as the control) was inoculated into each of six ten day old embryos via the allantoic cavity. Active virus was indicated by the hemagglutination of a 0.5% suspension of chicken red blood cells by the infected allantoic fluid.

Dilution of virus-germicide or virus-saline mixture.	Results	
	Control Virus-Saline	Test Virus-Germicide
10 ⁻¹	+++++	TTTT00
10 ⁻²	+++++	TTTT00
10 ⁻³	+++++	TTTT00
10 ⁻⁴	+++++	TTTT00
10 ⁻⁵	+++++	TTTT00
10 ⁻⁶	+++++	TTTT00
10 ⁻⁷	+++++	TTTT00
10 ⁻⁸	+++++	TTTT00

+ = Virus present
 0 = Virus absent
 T = Toxicity due to germicide. Presence or absence of virus could not be determined.

Summary. Under the above test conditions the compound inactivated four logs of virus.

HERPES SIMPLEX

Procedure. The virus was grown and assayed in rabbit kidney culture. The titer in cell culture was 10^7 tissue culture doses per 0.1 ml as indicated by the cytopathogenic effect of the virus after 5 days incubation at 37°C. For the test, 1 ml of virus was added to 9 ml of the germicide. After 10 minutes contact at room temperature the virus-germicide mixture was diluted in broth to the titer of the virus and inactivation was determined by comparing the titer of the treated virus with that of the untreated virus. Viral control. Saline was substituted for the germicide. The presence of active virus was determined by the inoculation of 0.1 ml of the test preparations into each of four cell culture tubes. Cells were incubated for 5 days and observed for cytopathogenic effect.

Dilution of virus-germicide or virus-saline mixture.	Results	
	Control Virus-Saline	Test Virus-Germicide
10 ⁻¹	++++	TTTT
10 ⁻²	++++	0000
10 ⁻³	++++	0000
10 ⁻⁴	++++	0000
10 ⁻⁵	++++	0000
10 ⁻⁶	++++	0000
10 ⁻⁷	0000	0000
10 ⁻⁸	0000	0000

+ = Virus present
 0 = Virus absent
 T = Toxicity due to germicide. Presence or absence of virus could not be determined.

Summary. The compound completely inactivated the virus of herpes simplex under the above test conditions.

VACCINIA VIRUS

Procedure. The virus was grown and assayed in rabbit kidney culture. The titer in cell culture was 10^7 tissue culture doses per 0.1 ml as indicated by the cytopathogenic effect of the virus after 5 days incubation at 37°C. For the test, 1 ml of virus was added to 9 ml of the germicide. After 10 minutes contact at room temperature the virus-germicide mixture was diluted in broth to the titer of the virus and inactivation was determined by comparing the titer of the treated virus with that of the untreated virus. Viral control. Saline was substituted for the germicide. The presence of active virus was determined by the inoculation of 0.1 ml of the test preparations into each of four cell culture tubes. Cells were incubated for 5 days and observed for cytopathogenic effect.

Dilution of virus-germicide or virus-saline mixture.	Results	
	Control Virus-Saline	Test Virus-Germicide
10 ⁻¹	++++	TTTT
10 ⁻²	++++	TTTT
10 ⁻³	++++	TTTT
10 ⁻⁴	++++	TTTT
10 ⁻⁵	++++	TTTT
10 ⁻⁶	++++	TTTT
10 ⁻⁷	++++	TTTT
10 ⁻⁸	++++	TTTT

+ = Virus present
 0 = Virus absent
 T = Toxicity due to germicide. Presence or absence of virus could not be determined.

Summary. Under the above test conditions the compound inactivated four logs of virus.

CHEMICAL AND PHYSICAL PROPERTIES

pH undiluted - 7.3	Phosphates - Contains no phosphorous compounds.
pH diluted 3:640 - 7.8	Film residue - Leaves no scum or residue at recommended use dilution.
Odor - Pleasant and mild.	Corrosion factor - Non-corrosive to metals at the proper use dilution.
Appearance - Translucent colorless liquid.	Toxicity - The use of protective gloves is recommended for manual application. Concentrate produces eye damage and severe skin irritation.
Viscosity - Free flowing liquid	Stability - Stable at normal temperatures. Does not discolor on exposure to light or heat.
Foam level - Moderate.	Active quaternary - 10%
Specific gravity - 1.0014	
Solubility - Readily soluble at recommended dilution in hot, cold, soft, or hard water.	
Discoloration - Non-staining.	

USE DIRECTIONS

FOOD PROCESSING EQUIPMENT: Use this product at 1/4 ounce per gallon of water. Sanitizes previously cleaned Food Processing Equipment and Food Utensils at 200 ppm. of active quaternaries. At this level, POTABLE WATER RINSE IS NOT REQUIRED.

RESTAURANT AND BAR RINSE: Use this product at 1/4 ounce per gallon of water. Sanitizes previously washed and rinsed dishes, glassware, silverware, cooking utensils.

DAIRIES: To sanitize dairy equipment such as tanks, lines, pails, and milk cans, first clean and rinse the equipment thoroughly. Then apply sanitizing solution containing 1/4 ounce of this product to 1 gallon of water (200 ppm.). Follow recommendations of local Health Board.

INSTITUTIONS: Sanitize with 1/4 ounce of this product to 1 gallon of water. Disinfect with 3 1/2 ounces of this product to 5 gallons of water. For heavily soiled or contaminated areas, a precleaning step is recommended. For sanitizing and disinfecting Floors, Walls, and Inanimate Hard Surfaces: Locker Rooms, Garbage Pails, Sink Tops, and Shower Stalls.

HOSPITALS: Use this product at 3 1/2 ounces per 5 gallons of water. Apply with cloth, sponge mop, or spray. Disinfects Floors and Walls as well as Sink Tops, Garbage Pails, Telephones, Shower Stalls, Locker Rooms, and Restrooms.

COLD DISINFECTION: Submerge instruments into solution containing 1 1/2 ounces of this product per gallon of water for 10 minutes for disinfection of previously cleaned surgical instruments, barber and dental equipment.

ROOM FOGGING: Apply solution of this product as a fog, to minimize the danger of cross-infection from environmental surfaces, before applying standard routine terminal cleaning and disinfecting practices.

Remove all human, animal, and plant life from room. Before fogging, food products and packaged materials must be removed from the room or carefully protected. Open closet doors and drawers. Set up 34" revolving platform in center of room. Mount FOGMASTER TRI-JET model 6208 (manufactured by AFA CORP. subsidiary of Thiokol Chemical Corp.) or other fogging device, delivering equivalent spray at 3 RPM. Fill Sprayer reservoir with 1 1/2 ounces of this product. Set sprayer mechanism to deliver 1 gallon of solution. Fog for 15 minutes for an average 2100 cu. ft. room. For different room sizes, vary spray time proportionately to ensure complete wetting of exposed surfaces. Wait 2 hours before entering room after treatment. A potable water rinse must be made on all food contact surfaces before their use. Rinse fogging equipment thoroughly with clear water following use.

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