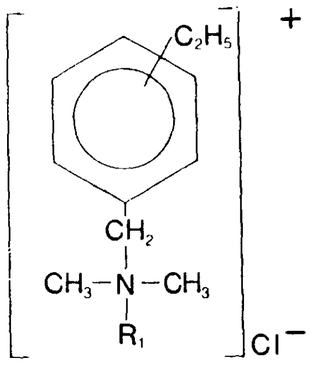
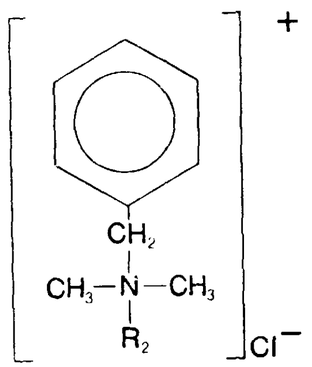




SaniMaster II

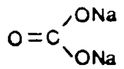
CLEANER – DISINFECTANT – DEODORIZER
E. P. A. Reg. No. 6109–13

Manufactured by
ServiceMaster Industries, Inc.
2300 Warrenville Road
Downers Grove, Illinois 60515, U. S. A.

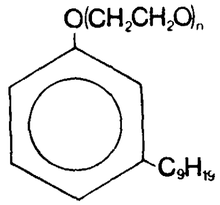
SaniMaster II is a germicidal detergent designed for use in hospitals, food handling areas, washrooms, and any other areas where disinfection is required. It is a quaternary based product utilizing two quaternary ammonium chloride compounds of the following composition:

Alkyl (60% C_{14}, 30% C_{16}, 5% C_{12}, 5% C_{18}) Dimethyl Benzyl Ammonium Chlorides	4.5%
Alkyl (68% C_{12}, 32% C_{14}) Dimethyl Ethylbenzyl Ammonium Chlorides	4.5%

For hard water [illegible] mine tetraacetic acid [illegible] metal ions such as calcium and magnesium which [illegible]. Laboratory tests have directed SaniMaster II is effective against *Staphylococcus aureus* and *Escherichia coli* in the presence of 350 and 300 ppm. hard water respectively as determined by the A.O.A.C. Germicidal and Detergent Sanitizer test method.



To enhance the detergency of SaniMaster II, sodium carbonate at a level of 47 is used as a builder to provide alkalinity.



For cleaning, SaniMaster II utilizes a non-ionic surfactant specifically selected for removal of soil on hard surfaces.

For disinfection of hard surfaces, use SaniMaster II at a concentration of 1 oz./gal. (1:128). When disinfecting surfaces contaminated with heavy soil or organic material, a precleaning step is required.

For sanitizing use SaniMaster II at a concentration of 1.4 oz./5 gal. (1:458). This corresponds to an active quaternary level of 200 ppm. When used on food equipment or food contact surfaces, use at this level. To properly sanitize such items thoroughly, clean them with a detergent. Rinse with potable water. Follow with SaniMaster II at 1.4 oz./5 gal.

Rinse with potable water.

Rinse empty container thoroughly with water and discard.

Danger: Keep out of reach of children.

Causes skin irritation, may cause eye damage.
Do not get in eyes, on skin, or on clothing.
Harmful if swallowed. Avoid contamination of food.

A TEST REPORT FROM

Onyx Microbiology Laboratory

Jersey City, New Jersey

Date: September 14, 1973

BACTERICIDAL, FUNGICIDAL, AND SANITIZING TEST RESULTS

PURPOSE

To determine the bactericidal, fungicidal, and sanitizing properties of SaniMaster II.

TEST SAMPLES

The product tested was identified as SaniMaster II, manufactured by ServiceMaster Industries, Inc., Downers Grove, Illinois. SaniMaster II is registered with the Environmental Protection Agency under E.P.A. Reg. No. 6109-13.

Three samples were received on 5—10—73 and identified as follows:

Sample No. 1	SaniMaster II — prepared 2—1—73
Sample No. 2	SaniMaster II — prepared 4—19—73
Sample No. 3	SaniMaster II — prepared 5—25—73

SUMMARY OF TEST RESULTS

I. Use—Dilution Method

A. Three samples representing three discrete preparations of SaniMaster II satisfactorily demonstrated disinfection against *Staphylococcus aureus*, ATCC No. 6538, *Salmonella choleraesuis*,ATCC No. 10708, and *Pseudomonas aeruginosa*, PRD 10, ATCC No. 15442 at a 1 oz/gal. (1:128) dilution.

B. In addition, one sample of SaniMaster II satisfactorily demonstrated disinfection against the following organisms at 1:128 (1 oz/gal.) dilution.

Staphylococcus epidermidis, ATCC No. 14990
Klebsiella pneumoniae, ATCC No. 4352
Serratia marcescens, ATCC No. 13880
Escherichia coli, ATCC No. 11229
Proteus vulgaris, ATCC No. 8427
Salmonella typhi, ATCC No. 6539

II. Fungicidal Test

One sample of SaniMaster II satisfactorily demonstrated fungicidal activity at a dilution of 1:128 (1 oz/gal.) against *Trichophyton mentagrophytes*, (NIH) Strain No. 640, ATCC No. 9533.

III. Sanitizing Tests

Three samples representing three discrete preparations of SaniMaster II at a use dilution of 1:458 (1 oz/28 pts.) demonstrated sanitizing properties when tested against *Staphylococcus aureus*, ATCC No. 6538, and *Escherichia coli*, ATCC No. 11229, in the presence of 350 and 300 ppm synthetic hard water (as $CaCO_3$) respectively.

IV. Virucidal Tests

SaniMaster II is a cleaner, disinfectant based on a prototype formulation containing 9.0% active quaternary. The virucidal data presented was used to support labeling claims of SaniMaster II for E.P.A. registration. The prototype formulation demonstrated virucidal activity against the following viruses at 1oz. gal.(1:128) dilution.

Type 2 Adenovirus	Vaccinia Virus
Influenza A_2 (Hong Kong)	Herpes Simplex

Date: 9/14/73

PROCEDURE

The test samples were evaluated by the bactericidal, fungicidal, and sanitizing tests at concentrations equivalent to the dilutions presented in this report. The samples were tested in accordance with the *Official Methods of Analysis of the Association of Official Analytical Chemists*, Eleventh Edition, 1970, Chapter 4 — Disinfectants.

1. Bactericidal Tests

A. Use–Dilution Method

Samples of SaniMaster II were evaluated by the use-dilution procedure given in the above-cited reference, paragraphs 4.007 — 4.011.

Three samples representing three discrete preparations were tested with 60 replicate carriers against *Staphylococcus aureus*, ATCC No. 6538, 30 replicate carriers against *Salmonella choleraesuis*, ATCC No. 10708, and 30 replicate carriers against *Pseudomonas aeruginosa*, PRD–10, ATCC No. 15442. One of the three preparations represented a 60 day old sample used to check product stability.

B. One preparation of SaniMaster II was evaluated with 10 replicate carriers against each of the following organisms.

Staphylococcus epidermidis, ATCC No. 14990
Klebsiella pneumoniae, ATCC No. 4352
Serratia marcescens, ATCC No. 13880
Proteus vulgaris, ATCC No. 8427
Salmonella typhi, ATCC No. 6539

II. Fungicidal Tests

The fungicidal activity of the test material was determined by the procedure given in the above-cited reference, paragraphs 4.018 — 4.022. The tests were conducted against *Trichophyton mentagrophytes*, (NIH) Strain No. 640, ATCC No. 9533.

III. Sanitizing Tests

The test material was evaluated by the Germicidal and Detergent Sanitizer method in the above-cited reference, paragraph 4.023. The tests were conducted against *Staphylococcus aureus*, ATCC No. 6538, and *Escherichia Coli*, ATCC No. 11229.

SUPPORTING DATA

I. Use-Dilution Tests

Table 1. SaniMaster II vs. *Staphylococcus aureus*, ATCC No. 6538
Exposure: 10 minutes at 20°C
Subculture medium: AOAC Letheen Broth
Use-Dilution: 1:128 (1 oz./gal.)

SAMPLE	POSITIVE SUBCULTURES	NEGATIVE SUBCULTURES
1 60 days old	0/60	60/60
2	0/60	60/60
3	0/60	60/60

Table 2. SaniMaster II vs. *Salmonella choleraesuis*, ATCC No. 10708.
Exposure: 10 minutes at 20°C
Subculture medium: AOAC Letheen Broth
Use-Dilution: 1:128 (1 oz./gal.)

SAMPLE	POSITIVE SUBCULTURES	NEGATIVE SUBCULTURES
1 60 days old	0/30	30/30
2	0/30	30/30
3	0/30	30/30

Table 3 SaniMaster II vs. *Pseudomonas aeruginosa*, PRD 10, ATCC No. 15442
Exposure: 10 minutes at 20°C
Subculture medium: AOAC Letheen Broth
Use Dilution: 1:128 (1 oz./gal.)

SAMPLE	POSITIVE SUBCULTURES	NEGATIVE SUBCULTURES
1 60 days old	0/30	30/30
2	0/30	30/30
3	0/30	30/30

Table 4 SaniMaster II vs. additional organisms using one sample preparation.
Exposure: 10 minutes at 20°C
Subculture medium: AOAC Letheen Broth
Use Dilution: 1:128 (1 oz./gal.)

ORGANISM	POSITIVE SUBCULTURES	NEGATIVE SUBCULTURES
Staphylococcus epidermidis ATCC No. 14990	0/10	10/10
Klebsiella pneumoniae ATCC No. 4352	0/10	10/10
Serratia marcescens ATCC No. 13880	0/10	10/10
Escherichia coli ATCC No. 11229	0/10	10/10
Proteus vulgaris ATCC No. 8427	0/10	10/10
Salmonella typhi ATCC No. 6539	0/10	10/10

II. Fungicidal Tests

Table 1. SaniMaster II vs. ***Trichophyton mentagrophytes***, ATCC No. 9533 using one sample preparation.
Exposure: 20°C
Incubation Period: 10 days at 25 to 30°C
Subculture medium: Azoleetin/Tween 80 dextrose broth

Exposure Time In Minutes	SAMPLE DILUTION 1:256 1/2 oz/gal.	 1:128 1 oz./gal.	 1:128 1 oz./gal.	 1:64 2 oz./gal.	 1:64 2 oz./gal.
5	+	+			
10	+				
15			-		

\+ growth of test organism in subculture tubes.
\- no growth in subculture tubes.

III. Sanitizing Tests

Table 1. SaniMaster II vs. *Escherichia coli*, ATCC No. 11229.
Use Dilution: 1:458 (1 oz./28 pts.)
ppm active quaternary: 200
Subculture medium: Tryptone glucose ext. agar.
Exposure: 25°C
Culture density: 1.14×10^8

Sample	Hard water ppm. as $CaCO_3$	30 sec. exposure No. Survivors	 % Kill	60 sec. exposure No. Survivors	 % Kill
1 60 days old	250	1100	99.999	30	>99.999
	300	1590	99.999	320	>99.999
	350	1905	99.998	330	>99.999
	400	2250	99.998	390	>99.999
	450	3300	99.997	595	>99.999
2	250	880	>99.999	45	>99.999
	300	1410	99.999	360	>99.999
	350	1710	99.999	395	>99.999
	400	2185	99.998	415	>99.999
	450	3500	99.997	680	>99.999
3	250	930	>99.999	60	>99.999
	300	1575	99.999	245	>99.999
	350	1860	99.998	390	>99.999
	400	2230	99.998	425	>99.999
	450	3650	99.997	680	>99.999

Hard water tolerance as determined against *E. coli* 300 ppm.

Table 2. SaniMaster II vs. *Staphylococcus aureus*, ATCC No. 6538
Use-Dilution: 1:458 (1 oz./28pts.)
ppm active quaternary: 200
Subculture medium: Tryptone glucose ext. agar
Exposure: 25°C
Culture density: 1.1×10^8

SAMPLE	Hard Water ppm as $CaCO_3$	30 sec. exposure No. Survivors	30 sec. exposure % Kill	60 sec. exposure No. Survivors	60 sec. exposure % Kill
1	250	500	> 99.999	< 10	> 99.999
60 days	300	1325	99.999	210	> 99.999
old	350	1645	99.999	300	> 99.999
	400	2085	99.998	395	> 99.999
	450	3100	99.997	345	> 99.999
2	250	680	> 99.999	15	> 99.999
	300	1285	99.999	180	> 99.999
	350	1590	99.999	315	> 99.999
	400	1920	99.998	425	> 99.999
	450	3100	99.997	590	> 99.999
3	250	615	> 99.999	10	> 99.999
	300	1305	99.999	195	> 99.999
	350	1610	99.999	325	> 99.999
	400	1965	99.998	370	> 99.999
	450	3000	99.997	615	> 99.999

Hard water tolerance as determined against *S. aureus* 350 ppm.

Vaccinia Virus

Germicide dilution 1:128 (1 ounce per 1 gallon of water)

Procedure. The virus was grown and assayed in rabbit kidney cell culture. The titer in cell culture was 10^7 tissue culture doses per 0.1 ml as indicated by the cytopathogenic effect of the virus after 5 days incubation at 37° C.

For the test, 0.1 ml of undiluted virus was added to 0.9 ml of the germicide diluted in water. After 10 minutes contact at room temperature the virus-germicide mixture was diluted in broth to the titer of the virus and inactivation was determined by comparing the titer of the treated virus with that of the untreated virus. **Viral control.** Saline was substituted for the germicide. The presence of active virus was determined by the inoculation of 0.1 ml of the test preparations into each of three cell culture tubes. Cells were incubated for 5 days and observed for cytopathogenic effect.

RESULTS

Dilution of virus-germicide or virus-saline mixture.	Controls Virus-saline	Test Virus-Germicide
10^{-1}	+ + +	T T T
10^{-2}	+ + +	T T T
10^{-3}	+ + +	0 0 0
10^{-4}	+ + +	0 0 0
10^{-5}	+ + +	0 0 0
10^{-6}	+ + +	0 0 0
10^{-7}	+ + 0	0 0 0
10^{-8}	0 0 0	0 0 0

\+ Virus present
0 Virus absent
T Toxicity due to germicide. Presence or absence of virus could not be determined.

Summary. Under the above experimental conditions the test virus was inactived by the germicide.

10/11/71
Morton Klein

Influenza A_2 (Hong Kong)

Germicide dilution 1:128 (1 ounce per 1 gallon of water)

Procedure. The virus was grown in the allantoic sac of 10 day old chick embryos. The virus was harvested after 40 hours incubation at 37° C and had an infectivity titer of 10^7 egg infectious doses per 0.1 ml of allantoic fluid, as determined by hemagglutination of a 0.5% suspension of chicken red blood cells.

The virus diluted 1:5 in saline was mixed with 9 part of a 1:64 dilution of the germicide. After 10 minutes contact at room temperature, ten-fold serial dilutions were prepared in broth and 0.1 ml.of the virus-germicide (or virus-saline, control) inoculated into each of six 10 day old embryos via the allantoic cavity. Active virus was indicated by the hemagglutination of a 0.5% suspension of chicken red blood cells by the infected allantoic fluid.

RESULTS

Dilution of virus-germicide or virus-saline mixture	Controls Virus-saline	Test Virus-Germicide
10^{-1}	+ + + + + +	0 0 0 0 0 D
10^{-2}	+ + + + + +	0 0 0 0 0 0
10^{-3}	+ + + + + +	0 0 0 0 0 0
10^{-4}	+ + + + + +	0 0 0 0 0 0
10^{-5}	+ + + + + D	0 0 0 0 0 0
10^{-6}	+ + + + + D	0 0 0 0 0 0
10^{-7}	0 0 0 0 0 0	0 0 0 0 0 0

\+ Virus present
0 Virus absent
D Eggs dead of non-specific factors.

Summary. Under the above experimental conditions the test compound completely inactivated Influenza A_2.

10/11 71
Morton Klein

Virus of Herpes simplex

Germicide dilution 1:128 (1 ounce per 1 gallon of water)

Procedure. The virus was grown in rabbit kidney cell culture. The titer of the virus was 10^7 tissue culture doses per 0.1 ml. The presence of the virus was indicated by the cytopathogenic effect in cell culture. For the test, 0.1 ml of undiluted virus was added to 0.9 ml of germicide diluted in water. After 10 minutes contact at room temperature the virus-germicide mixture was diluted in broth to the titer of the virus and inactivation was determined by comparing the titer of the treated virus with that of the untreated virus. **Viral control.** Saline was substituted for the germicide. The presence of active virus was determined by the inoculation of 0.1 ml of the test preparations into each of three cell culture tubes. Cells were incubated at 37° C or 4 days and observed for cytopathogenic effect.

RESULTS

Dilution of virus-germicide or virus-saline mixture.	Controls Virus-Saline	Test Virus-Germicide
10^{-1}	+ + +	T T T
10^{-2}	+ + +	T T T
10^{-3}	+ + +	0 0 0
10^{-4}	+ + +	0 0 0
10^{-5}	+ + +	0 0 0
10^{-6}	+ + +	0 0 0
10^{-7}	+ + 0	0 0 0
10^{-8}	0 0 0	0 0 0

\+ Virus present
0 Virus absent
T Toxicity due to germicide. Presence or absence of virus could not be determined.

Summary. Under the above experimental conditions the germicide inactivated the virus of Herpes Simplex.

10 11 71
Morton Klein

Type 2 Adenovirus

Germicide dilution 1:128 (1 ounce per 1 gallon of water)

Procedure. The virus was grown and assayed in Hela cells. The titer of the virus was 10^5 tissue culture doses per 0.1 ml. Tubes were read after 7 days incubation at 37°C. For the test, 0.1 ml of undiluted virus was added to 0.9 ml of germicide diluted in water. After 10 minutes contact at room temperature the virus-germicide mixture was diluted in broth to the titer of the virus and inactivation was determined by comparing the titer of the treated virus with that of the untreated virus. **Virus control.** Saline was substituted for the germicide. The presence of active virus was determined by the inoculation of 0.1 ml of the test preparations into each of three cell culture tubes. Cells were incubated at 37°C for 5 days and observed for cytopathogenic effect.

RESULTS

Dilution of virus-germicide or virus-saline mixture	Controls Virus-saline	Test Virus-Germicide
10^{-1}	+ + +	T T T
10^{-2}	+ + +	T T T
10^{-3}	+ + +	0 0 0
10^{-4}	+ + +	0 0 0
10^{-5}	+ + 0	0 0 0
10^{-6}	0 0 0	0 0 0

\+ Virus present
0 Virus absent
T Toxicity due to germicide. Presence or absence of virus could not be determined.

Summary. Under the above experimental conditions the test virus was inactived by the germicide.

10/11/71
Morton Klein

WELLS LABORATORIES, INC.

25-27 LEWIS AVENUE, JERSEY CITY, N. J. 07306
PHONES • Jersey City: (201) 653-6036 • New York City: Digby 9-0127
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SAUL FRANCES, Ph.D., *Executive Director*
ARTHUR F. PETERSON, Ph.D.
FRANK PORTER, Ph.D.
KWANG CHUNG, Ph.D.

Laboratory No. D-7814

June 11, 1971

REPORT ON PRIMARY SKIN IRRITATION
STUDIES IN RABBITS
USING SANIMASTER II

CLIENT: Onyx Chemical Company
190 Warren Street
Jersey City, N. J. 07302

Sample: On June 4, 1971, a screw capped bottle containing a clear liquid was received. It was identified as Sanimaster II.

Procedure: The method employed is described in the U.S. Department of Agriculture, Federal Insecticide, Fungicide and Rodenticide Act, section 362.3 of the regulations (7CFR part 362), paragraph (c).

Results: Please see attached table.

Primary Skin Irritation Index = 4.0

Conclusions: Sanimaster II, tested as indicated, produced mild to moderate erythema and edema in both abraded and intact rabbit skins.

Respectfully submitted,

Joseph P. de Arevedo
Laboratory Animal Technologist

Arthur F. Peterson, Ph.D.
Director of Laboratories
WELLS LABORATORIES, INC.

JPA:AFP/av

Laboratory No. D-7814

June 11, 1971

PRIMARY SKIN IRRITATION INDEX – RABBITS

	Exposure time Hours	Exposure Unit Value Rabbit 1	Exposure Unit Value Rabbit 2	Exposure Unit Value Rabbit 3	Exposure Unit Value Rabbit 4	Exposure Unit Value Rabbit 5	Exposure Unit Value Rabbit 6
Erythema and eschar formation:							
Intact skin	24	2	2	2	2	2	2
do	72	2	2	2	2	2	2
Abraded skin	24	3	3	3	3	3	3
do	72	3	3	3	3	3	3
Subtotal		10	10	10	10	10	10
Edema formation:							
Intact Skin	24	1	1	1	1	1	1
do	72	1	1	1	1	1	1
Abraded skin	24	2	2	2	2	2	2
do	72	2	2	2	2	2	2
Subtotal		6	6	6	6	6	6
Total		16	16	16	16	16	16
AVERAGE		4	4	4	4	4	4

Primary Skin Irritation Index 4.0

WELLS LABORATORIES, INC.

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SAUL FRANCES, Ph.D., *Executive Director*
ARTHUR F. PETERSON, Ph.D.
FRANK PORTER, Ph.D.
KWANG CHUNG, Ph.D.

Laboratory No. D-7815

June 28, 1971

REPORT ON EYE IRRITATION TESTING
IN RABBITS USING SANIMASTER II

Client: Onyx Chemical Co.
190 Warren Street
Jersey City, N.J. 07302

Sample: On June 4, 1971, a screw-capped bottle containing a clear liquid was received. It was identified as: Sanimaster II.

Procedure: The method employed is described in the U.S. Department of Agriculture, Federal Insecticide, Fungicide and Rodenticide Act, Section 362.116 of the regulations (7CFR part 362) paragraph (d).

Dose: 0.1 ml. in one eye of each of six rabbits.

Results: Please see attached table.

Conclusions: Sanimaster II, tested as indicated produced corneal, iritic and conjunctival irritation. None of the 6 test rabbits showed complete correction at the end of 14 days of observation.

Respectfully submitted,

Joseph P. de Azevedo
Laboratory Animal Technologist

Arthur F. Peterson, Ph.D.
Director of Laboratories
WELLS LABORATORIES, INC.

JPA:AFP/av

Lab No D 7815 June 28, 1971

Findings:

Rabbit No. 1	DAYS 1	2	3	4	5	6	7	8	9	10	11	12	13	14
I. Cornea														
A. Opacity	0	1	1	1	1	1	1	1	1	1	1	1	1	1
B. Area	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X B X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
II. Iris														
A. Values	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
III. Conjunctivae														
A. Redness	3	3	3	3	3	3	3	3	2	2	2	1	0	0
B. Chemosis	3	3	3	3	3	3	3	3	3	2	2	1	0	0
C. Discharge	3	3	3	3	3	3	3	3	2	2	2	1	0	0
(A+B+C) X 2	18	18	18	18	18	18	18	18	14	12	12	6		
Total	18	28	28	28	28	28	28	28	24	22	22	16	10	10

Rabbit No. 2

I. Cornea														
A. Opacity	0	1	1	1	1	1	1	1	1	1	1	1	1	1
B. Area	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X B X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
II. Iris														
A. Values	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
III. Conjunctivae														
A. Redness	3	3	3	3	3	3	3	3	2	2	2	1	0	0
B. Chemosis	3	3	3	3	3	3	3	3	2	2	2	1	0	0
C. Discharge	3	3	3	3	3	3	3	3	2	2	2	1	0	0
(A+B+C) X 2	18	18	18	18	18	18	18	18	12	12	12	6		
Total	18	28	28	28	28	28	28	28	22	22	22	16	10	10

Rabbit No. 3

I. Cornea														
A. Opacity	0	1	1	1	1	1	1	1	1	1	1	1	1	1
B. Area	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X B X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
II. Iris														
A. Values	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
III. Conjunctivae														
A. Redness	3	3	3	3	3	3	3	3	2	2	2	1	0	0
B. Chemosis	3	3	3	3	3	3	3	3	2	2	2	1	0	0
C. Discharge	3	3	3	3	3	3	3	3	2	2	2	1	0	0
(A+B+C) X 2	18	18	18	18	18	18	18	18	12	12	12	6		
Total	18	28	28	28	28	28	28	28	22	22	22	16	10	10

Lab. No. D-7815 June 28, 1971

Findings:

Rabbit No. 4	DAYS 1	2	3	4	5	6	7	8	9	10	11	12	13	14
I. Cornea														
A. Opacity	0	1	1	1	1	1	1	1	1	1	1	1	1	1
B. Area	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X B X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
II. Iris														
A. Values	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
III. Conjunctivae														
A. Redness	3	3	3	3	3	3	3	3	3	3	2	1	0	0
B. Chemosis	3	3	3	3	3	3	3	3	3	3	2	1	0	0
C. Discharge	3	3	3	3	3	3	3	3	3	3	2	1	0	0
(A+B+C) X 2	18	18	18	18	18	18	18	18	18	18	12	6		
Total	18	28	28	28	28	28	28	28	28	28	22	16	10	10

Rabbit No. 5

I. Cornea														
A. Opacity	0	1	1	1	1	1	1	1	1	1	1	1	1	1
B. Area	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X B X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
II. Iris														
A. Values	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
III. Conjunctivae														
A. Redness	3	3	3	3	3	3	3	3	3	3	2	1	0	0
B. Chemosis	3	3	3	3	3	3	3	3	3	3	2	1	0	0
C. Discharge	3	3	3	3	3	3	3	3	3	3	2	1	0	0
(A+B+C) X 2	18	18	18	18	18	18	18	18	18	18	12	6		
Total	18	28	28	28	28	28	28	28	28	28	22	16	10	10

Rabbit No. 6

I. Cornea														
A. Opacity	0	1	1	1	1	1	1	1	1	1	1	1	1	1
B. Area	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X B X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
II. Iris														
A. Values	0	1	1	1	1	1	1	1	1	1	1	1	1	1
A X 5		5	5	5	5	5	5	5	5	5	5	5	5	5
III. Conjunctivae														
A. Redness	3	3	3	2	1	1	1	2	3	3	2	1	0	0
B. Chemosis	3	3	3	2	1	1			3	3	2	1	0	0
C. Discharge	3	3	3	2	1	1			3	3	2	1	0	0
(A+B+C) X 2	18	18	18	12	6	6	6	8	18	18	12	6		
Total	18	28	28	22	16	16	16	23	33	33	22	16	10	10