



UNITED STATES ENVIRONMENTAL PROTECTION AGENCY  
WASHINGTON, D.C. 20460

15  
(9-22-98)

OFFICE OF  
PREVENTION, PESTICIDES AND  
TOXIC SUBSTANCES

MEMORANDUM

**SUBJECT:** EFED RED Chapter for Malathion  
PC Code 057701; Case No. 818961  
DP Barcodes: D238903 and D238906 + D238904

**TO:** Dana Lateulere, Chemical Review Manager  
Insecticide Risk Branch  
Special Review and Reregistration Division(7508W)

**FROM:** Brian Montague, Fisheries Biologist *Brian Montague 9/21/98*  
Team Leader, Ecological Effects Assessment  
Norman Birchfield, Chemist *Norman Birchfield 9/22/98*  
Environmental Exposure Assessment  
Richard Mahler, Hydrologist *Richard J. Mahler 9/22/98*  
Environmental Fate Assessment

**THROUGH:** Arnet Jones, Chief *Arnet Jones 09/21/98*  
Environmental Risk Branch I/EFED(7507C)

This memo provides a summarization of EFED's Environmental Risk Assessment for the Malathion Reregistration Eligibility Document. It includes brief summaries of suggested mitigation, weaknesses and uncertainties of malathion data sets, additional required data needs, and the major areas of environmental risk concern. The Malathion OPP RED team may convene in future months to review all Malathion Chapters as they are completed and discussions regarding mitigation of malathion hazard potential may be modified or enhanced by these discussions. Any OPP Malathion team recommended modifications to the Ecological Risk Chapter will be returned to EFED for further comment.

EFED has summarized the major ecological issues of concern for malathion usage as follows:

1. Malathion is highly toxic to aquatic organisms at concentrations which have been monitored or are predicted to occur in association with many presently registered use patterns. Malathion has been implicated in numerous fish kill incidents over its five decades of use. Malathion has been shown to be toxic to economically valuable invertebrate species under label permitted application in field conditions.
2. Malathion is potentially hazardous to reproductive success of certain species of birds and mammals at presently registered use rates. In addition, malathion may pose sublethal effects to terrestrial and aquatic organisms through neurological inhibition which may adversely impact their survival in the natural environment. These concerns are based on laboratory and field study observations, measurement of reduced acetylcholinesterase levels in exposed wildlife, and measurement of malathion residues in tissues of organisms which were adversely affected from malathion exposure.
3. Malathion is highly toxic to beneficial insects. Direct contact during application, post application contact with foliar residues, and contact with pollen transported residues have all proven toxic to honeybees. Malathion is highly toxic to aquatic larval stages of terrestrial insects when transported to water. Use of malathion in indiscriminate types of applications (fogging or broad-target aerial) to urban and rural sites is expected to impact numerous species other than the intended target insect pests.
4. Malathion has high potential to drift off target from aerial ULV applications and has been monitored at levels high enough to pose concern for non-target invertebrates as far as 200 meters from the application site (Penn. State Boll Weevil Eradication study-see RED text for details). This concern is heightened by the fact that there are presently no aerial buffers on any malathion labels.

The overall impact of malathion on the environment has likely been mitigated by several factors over its long history of usage in the United States. Some of these factors are:

1. Malathion appears to degrade rapidly in the presence of active soil microbial activity. This activity varies in certain types of soil with degradation most rapid in moist soils.
2. Malathion appears to degrade quickly in water with high pH (> 7.0).
3. In general, malathion has not been observed to be as persistent on soil or in water as some of the other organophosphate insecticides with which it competes and it displays lower acute toxicity to birds and mammals.
4. In general, large tributaries with rapid flow rates have been adequately flushed to prevent buildup of malathion to acutely toxic levels for fish. Thus, most fish kills have been in small streams or ponds where slow flow rates permitted concentrations to exceed toxic levels for fish or where heavy rainfall events to large watershed areas allowed high

concentration pulse loads to impact small aquatic habitat areas.

Areas where malathion labels might be modified to mitigate potential hazard to wildlife and aquatic organisms are suggested below. The present agreements by registrants to place restrictions on levels of usage for food crops may greatly reduce potential exposure. However, there are many other non-food uses of malathion where further label modifications, restrictions or advisories may aid in reduction of malathion concentrations in the environment.

### **Spray Drift and Runoff Reduction**

#### **Reduce ULV Usage**

Substitute use of aerial application formulations other than ULV. Aerial uses of ultra-low volume formulations are highly prone to drifting long distances, posing risks to human and wildlife through inhalation exposure and through deposition on food and water sources. The use of alternate formulations for aerial application, which are less prone to drift because of larger particle size, would dramatically reduce offsite movement of malathion.

#### **Require Protective Buffer Zones**

As mentioned previously, monitoring studies have shown that buffer zones will reduce off-target spray drift of malathion to aquatic habitats. The buffer zones required to prevent drift of ULV sprays may prove impractical where vegetation is sparse (>200 meters), however smaller buffer zones for conventional aerial application methods may prove practical for normal use. In addition, the types (heights) of vegetation common to the buffer zones may dictate the relative distance required to achieve adequate protection for aquatic habitats. At the present time no buffer zones are included on any malathion labels for aerial or mistblower application methods.

#### **Reduce Usage in Areas Prone to Runoff and Slower Degradation**

In many agricultural use sites malathion soil microbe degradation is rapid enough such that runoff does not pose as serious threat to many aquatic habitats and drinking water sources. However, application of malathion in residential areas, where it is prone to contact anthropogenic surfaces which are dry and microbially inactive, increase malathion persistence and also increase formation of its toxic degradate malaaxon. In addition, little malathion and malaaxon is expected to adsorb to most anthropogenic surfaces relative to soil, thus greatly increasing runoff potential. By limiting residential malathion application to ground sprays (as opposed to widespread aerial applications) and uses directed toward soil (such as garden use) malathion runoff levels in urban environments would be expected to decrease.

### **Reduction of Malathion Applied**

#### **Application rate reduction**

ULV formulations have low application efficiency due to their high drift potential. Thus, it may be possible to decrease application rates and still maintain comparable efficacy by using formulations with higher efficiencies. In some cases use rates may be higher than required for efficacy. For example, application rates in the cotton boll weevil eradication program vary from state to state and are controlled by individual states. The application rate varies from 0.625 lbs /

A in Mississippi to 1 lb / A in Louisiana. Applications rates are largely dictated by previously used rates and may not be the lowest effective rate. Members of USDA APHIS have reported even lower effective rates with improved organic co-solvent formulations. The predicted effect to aquatic exposures from reduction of rates is further detailed under mitigation discussions in the Malathion RED document.

#### Reduced Use in High Persistence Soils

Malathion persistence varies greatly in soil, ranging from less than one day to greater than 5 days. Soils with longer malathion persistence (e.g. dry with low organic matter content) would be expected to have higher runoff potential as residues fail to degrade before a rainfall event occurs. Reducing malathion usage or application rates in areas with soils that lead to longer malathion persistence would reduce runoff potential.

#### Reduction of seasonal use

Lower numbers of permitted seasonal applications at use rates in excess of 1.25 lbs ai/A will reduce levels and length of exposure of sensitive bird species and possibly other equally sensitive terrestrial wildlife species to multiple peaks of malathion levels which are in exceedance of chronic concern levels. In addition, the amount of residues potentially available for runoff to aquatic habitats would be reduced.

#### Increasing Multiple Application Intervals to 7 or More Days

Terrestrial first order residue estimates indicate that malathion degrades rapidly enough to avoid terrestrial residue buildup on vegetation in typical scenarios if intervals are 7 days or more. Slight increases in residues occur with 3 or 5 day intervals. Seven day or greater intervals appear to provide little residue increase over levels predicted for a single application when presently known fate parameters are used in estimating terrestrial EECs..

#### Storage conditions

Malathion degradation to products of higher toxicity under improper storage conditions is well documented, however effects due to impurities and degradates during normal use are not (with the exception of the mass poisoning of 2,800 spray men in Pakistan in 1976 resulting in five deaths, Aldridge *et al*, 1979). Malathion stored for long periods of time clearly increases ecological and human health risks, thus it would be appropriate to limit the age of malathion formulations being used and strengthen label language to reflect the increased toxicity associated with improper storage conditions.

#### Use Characterization

The environmental risk is based on the use information for malathion provided by USDA, the Biological and Economic Analysis Division of OPP, and use information provided by Cheminova on behalf of all malathion formulators. Cheminova and IR4 have indicated that number of uses which appear on present labels will no longer be supported including forest uses, tobacco, hops, and certain minor crops. Where no support was indicated for a particular use, it was excluded from the EFED assessment of potential ecological risk. Cheminova and IR4 have also provided the Agency with maximum application rates, minimum intervals, and maximum seasonal application numbers for all supported food crops. Risk assessment procedures were based on the

understanding that any labels exceeding these residue tolerance testing scenario limits for food crops would be revised during the reregistration of all malathion products. Including cotton boll weevil control malathion use totals over 3,370,000 lbs ai per year on food crops alone. Non-crop use is also extensive with over 4,170,000 lbs ai per year used for mosquito control, household uses, industrial sites, ornamental plants, forage and fodder, stored grains, roadways, forest lands, parks, ornamental turf, farmsteads and lots, cemeteries, golf courses, and fallow land. The extent of non-crop use is much less clear than that for food crop use. Maximum use rates were used to assess potential risk from non-crop uses of malathion.

### **Water Resources Assessment**

The water resource assessment, based on the known fate properties of malathion along with extensive monitoring data, indicates the following points.

1. Malathion is routinely detected in tributaries and retention ponds near urban areas when active monitoring programs to detect malathion are undertaken.
2. Malathion appears to undergo conversion to malaoxon during the processing of surface water to drinking water and also in the presence of certain purification chemicals used in swimming pools.
3. Despite the use of large buffer zones during ULV aerial applications to protect non-target water resources such as reservoirs, ponds and streams, malathion continues to be detected in these resources at varying levels.
4. Malathion detection is higher in urban areas than in areas where agricultural use predominates. EFED believes this is due to anthropogenic surface materials common to urban areas which are devoid of soil bacteria needed to degrade malathion and to multiple uses within a single watershed which drains via interconnected storm water systems to a single pond or storm water discharge point.

### **Ecological Risk Characterization**

#### **Risk to Terrestrial Vertebrates**

Malathion has been shown to offer potential chronic hazard to bobwhite quail at consistent dietary concentrations equal to or above 350 ppm. Based on this data malathion may offer potential hazard to birds at application rates above 2.5 lbs ai/acre if based on typical expected residues and above 1.25 lbs ai/acre if based on maximum expected concentrations. A mitigating factor may be that malathion residues may degrade quickly between applications if microbial activity is high, such as might be expected on soils. On the other hand, applications are permitted with as little as 3 day intervals and average half-life estimates on foliage ranged from 1 to 10 days, dependent on crop use and weather conditions. Effects noted in the avian reproduction study were reduced egg hatch and regressed ovaries which could indicate that early exposure to malathion may have led to the observed effects. EFED does not have data regarding long term exposure to repeated pulses of high residues and their potential relationship to chronic effects on reproduction. Direct exposure of nesting birds could lead to exposure of eggs or juvenile birds. Laboratory data has

been reviewed which indicates that developmental effects will occur when embryos of birds are exposed to malathion. The likelihood of this type of exposure is dependent on the method of application, the treated habitat type, and the time of year that applications are made. Sublethal effects to birds from acetylcholinesterase inhibition may include reduced nesting behavior, disorientation, and lost motor coordination leading to reduced ability to cope with the daily stresses of survival under natural conditions. This is also true, to some extent for mammals, reptiles or other terrestrial organisms which, though not acutely effected, may be sublethally susceptible to inhibition of acetylcholinesterase in neural processes.

#### Risk to Aquatic Organisms

Exposure to aquatic habitats has been documented in numerous monitoring studies at residue levels which exceed acute levels of concern for many species of invertebrates. Based on this information it is presumed that adverse effects may have occurred, but due to lack of population effect monitoring, gone undocumented. These monitored exposure levels have also reached levels of concern for adverse effects to fish, but less often. Malathion does have a number of reported fish kill incidents associated with its use in urban scenarios. In many of these incidents certain species of fish have been observed to be more severely impacted (e.g., sticklebacks) than other species in the same location. Fewer incident reports were received for malathion use in agricultural settings, though they do occur. In most cases, agricultural incidents occurred where farm ponds receive drift or runoff from several neighboring fields. Often feeder ditches funneled runoff to the ponds during rainfall events. Based on fate data, it would appear that the length of exposure to malathion residues may be dependent on the pH conditions of the receiving waters, with acidic conditions contributing to increased persistence. Malathion does not appear to accumulate in sediments. Long term exposure to aquatic organisms would be from repeated pulse loads related to rainfall conditions (primarily in urban scenarios) or repeated drift of residues to surface waters from multiple aerial applications.

#### Risk to Non-Target Beneficial Insects

Malathion is a potent insecticide which is expected to impact non-target insects along with the intended target insect pests. This is especially true when use sites are not a specifically targeted agricultural field or residential/commercial site. Examples of this might be application in ground fogs designed to drift and impact flying mosquitoes (adulticide use) or application to large tracts of rangeland to combat grasshoppers in locations away from agricultural sites which could potentially be impacted by them. Honeybee studies have also indicated that foliar residues of malathion are highly toxic 48 hours after application. Numerous data regarding the high toxicity of malathion to species of insects considered beneficial has been reviewed for or referenced in the RED document.

#### Data Gaps

##### Environmental Fate

Data for the aerobic aquatic metabolism studies are considered supplemental at this time. The deficiency of this study is the alkaline pH of the soil and water used. Degradation of malathion is highly pH dependent, with faster rates at higher pH, thus a quantitative assessment of malathion fate and persistence under acidic conditions when hydrolysis would be slower cannot be

conducted. As a result EFED cannot complete a quantitative assessment of the environmental fate of malathion and its degradates in acidic environments it is likely to contact. An aerobic aquatic metabolism study (162-4) performed under pH conditions that do not favor hydrolysis is requested. EFED believes this additional information will enable a more quantitative assessment of the fate and persistence of this major use chemical in acidic aquatic environments. This request is especially relevant given the sensitivity of numerous aquatic organisms to malathion.

The anaerobic aquatic metabolism study was also conducted under alkaline conditions favoring hydrolysis, EFED believes that hydrolysis data along with open literature data on the persistence of malathion in sediments is sufficient to conclude that malathion will not persist under anaerobic conditions and therefore a repeat of this study is not required at this time.

Fate data is required for malaoxon, the oxidation product of malathion. Malaoxon is commonly believed to be the neurotoxic agent of malathion after oxidation *in vivo* and toxicity data show it to have higher acute toxicity than malathion. EFED acknowledges that maximal conversion to malaoxon under registrant submitted study conditions was low (1.8%), however, under other conditions encountered during malathion use conversion levels as high as 10.7% of applied parent have been reported (CalEPA 1993). HED has indicated that malaoxon is to be included in the tolerance expression for malathion. First tier surface water assessments were performed by making several assumptions about the properties of malaoxon relative to malathion. It is not possible to perform second tier assessments without further data specific to malaoxon. Thus, EFED is requiring data needed to predict malaoxon levels in drinking water and aquatic habitats.

In addition to data on the basic physical properties of malaoxon (solubility, partition coefficient, vapor pressure) EFED requests that the following laboratory studies be submitted for malaoxon. Data from these studies are expected to be sufficient to perform basic fate and exposure modeling of malaoxon.

#### **Malaoxon Study Requirements and Justification:**

##### Degradation

###### 161-1 (hydrolysis)

Malathion hydrolysis is an important route of dissipation under alkaline conditions. The phosphorothiolate ester bond of malaoxon may be more susceptible to cleavage via hydrolysis than the analogous phosphorodithioate ester in malathion.

##### Metabolism

162-1 (aerobic soil) The primary route of malathion degradation on soil is through aerobic metabolism. An open literature study (Paschal and Neville 1976) suggests malaoxon persistence may be greater on soils. Additionally, CalEPA studies have shown levels of malaoxon production exceeding 10% in certain dry, low organic content soils.

###### 162-4 (aerobic aquatic)

Although little or no malaoxon production is observed in registrant submitted aquatic studies malaoxon has been detected in surface waters and the potential for malaoxon runoff may be heightened relative to malathion because it is expected to have higher solubility. Aerobic aquatic metabolism contributes greatly to malathion degradation.

### Mobility

163-1(leaching/adsorption)

EFED is not aware of reports of malaoxon groundwater adsorption/desorption) contamination, however, malathion has contaminated groundwater in several states and has the potential to contaminate surface water through runoff. The increased polarity of malaoxon due to the substitution of oxygen for sulfur increases the expected potential of this chemical to be mobile in soil.

EFED also requests additional information on environmental malaoxon production. Because malathion is used in a large number of settings including more than 60 terrestrial field uses as well as outdoor residential uses including mosquito, Mediterranean fruitfly, and urban pest control uses, it is exposed to a large variety of environmental conditions. This extensive use is likely to result in significant exposure of nontarget organisms to malathion breakdown products. Exposures to humans and wildlife may be through contamination of food, water, and air (by suspended particles) which can result from off-target drift, runoff, and direct application.

### **Ecological Effect Data Requirements and Limitations of Present Data**

The toxicological data, though extensive for malathion, is not complete in several key areas. In addition, much of the data is over twenty years old, and, to some extent, was not conducted in accordance with stricter standards which are required of studies presently submitted to support registration of pesticides. For example, most of the acute toxicity endpoints for aquatic organisms are based on nominal concentrations which, due to malathion's short aquatic persistence, may not be appropriate since this could lead to calculated LC50 values which are higher than would have been estimated if based on mean measured concentrations.

There are also some other areas where the data set is weak. In formulation testing only one presently supported product formulation (57% EC) was tested on 4 species (daphnid, oyster, honeybee, and sheepshead minnow). There are no submitted toxicity data on the mixture of malathion and methoxychlor, a possibly highly lethal combination for aquatic life. Acute toxicity testing (72-1 and 72-2) with this formulation is required for bluegill and *Daphnia magna*. There are no studies regarding the chronic effect levels of malathion to estuarine fish or invertebrates which could conceivably be exposed to repeated pulse load exposures for such uses as citrus and cotton. Chronic testing of technical malathion (72-4) with sheepshead minnow and mysid are required. Full life-cycle testing with fish (72-5) is held in reserve pending decision on an appropriate protocol modification. Further data to elucidate potential effects to non-target insect populations is needed though all normally required data has been satisfied at this time. Acute studies with honeybees indicate that acute contact with direct or latent residues may prove lethal for several days after application. Other beneficial insect populations may also suffer acute losses. There is some indication that amphibians could be affected by malathion exposure. Though not presently a data requirement requested by the Agency, but given what is known about acute and chronic effect levels observed in frogs, a better understanding of potential developmental effects to this taxa is needed to improve this assessment.



Sublethal effects caused by temporary disruption of nervous system functions are difficult to use in present risk assessment procedures, because so little is known about their ultimate effect on non-target species populations. However, malathion has been shown to disrupt nesting success in sharp tailed grouse, loss of ability of laboratory mice to navigate a maze, and loss of swimming ability for fish swimming against a current. All of these effects theoretically could lead to reduced survival of certain species groups, when combined with the normal stress factors associated with survival (e.g., successful rearing of young, escape from predators, and navigation to spawning grounds).

#### **Suggested Label Language Revisions**

1. Many malathion labels presently list application rates in exceedance of 6.25 lbs ai acre. These should be removed from all labels.
2. All malathion labels should specify a maximum number of applications. No labels should contain the statement "Use as Needed" when referring to maximum seasonal applications. Food crop limitations should be based on maximum tested tolerance scenarios.
3. All malathion labels with aerial and possibly mist-blower applications should specify protective buffer zones for aquatic habitats in accordance with spray drift data presently known for malathion or similar types of chemicals. Removal of ULV aerial application is suggested, unless protective buffer zones adequate to prevent drift of residues harmful to aquatic invertebrates is achievable.
4. All malathion labels with multiple applications should specify minimum intervals for each use. These intervals should be adequate to prevent buildup of foliar, soil, and water residues over time by allowing degradation of residues via natural processes.
5. All labels with label rates exceeding 1.25 lbs ai/A should contain precautions in the environmental hazard statement regarding the exposure of nesting birds to direct application. Malathion may lead to chronic reproductive and sublethally adverse effects to sensitive species of birds.
6. Uses which are no longer supported should be removed from all formulation labels. These modified labels should be reviewed for clarification and acceptance by the Agency, since certain uses (forest use on forest insects) are still listed on newest labels submitted by Cheminova, despite the fact that this use is stated as no longer supported. These uses include hops, forest use, forest trees, tobacco, peanuts, use on animals, and any uses directly over water.
7. Strengthen precautionary language on labels pertaining to storage time and stability.

#### **Peer Reviewers**

Arnet Jones (Use and fate), Kevin Costello (Groundwater), Ed Fite (Ecological Risk), James Hetrick (Fate) and within the team membership (B. Montague, Richard Mahler, and Norman Birchfield).

# Malathion Reregistration Eligibility Document

## Environmental Fate and Effects Chapter

### Table of Contents

<b>Introduction</b>	<b>Page</b>
Limitations and Agreements With Registrants Regarding the Uses and Application Scenarios Reviewed and Supported	5
<b>Use Characterization</b>	
Historic Perspective	
Summary of Supported Product Types, Formulations, and Use Scenarios	6
Summary of Major Uses Nationwide	6
Summary of USDA Special Program Uses	7
Bollweevil Eradication Program	
Mediterranean Fruit Fly Control Program	
Grasshopper Control on Federal Rangelands	
Summary of Public Health Uses	7
Mosquito Control-Populated and Rural Areas	
Summary of Supported Agricultural Crop Uses	
Food Crops	7-9
Non Food Crops	9
Summary of Other Non-Agricultural Uses	10-11
Non-Crop Uses-Agriculture Related	
Storage of Grains	
Livestock-Feed Lots and Holding Pens	
Beet Leafhopper Control-Non-Ag Lands-Plant Hosts	
Grasshopper Control Non-Federal Lands	
Fly Control-Cull Fruit and Vegetable Dumps	
Commercial Use Urban Scenarios	
Homeowner Use	
Forestry Uses	
Unsupported Crop Uses	
<b>Exposure Characterization</b>	
<b>Chemical Profile</b>	12
<b>Environmental Fate Assessment</b>	12
Environmental Fate Summary	
Summary of Studies : Fate and Transport	17-25
<b>Terrestrial Exposure Assessment</b>	
Terrestrial Exposure Estimates on Vegetation	25
Single/Multiple Application Tables	
Monitored Residues on Vegetation	29

<b>Water Resource Assessment-Malathion and Malaoxon</b>	
<b>Tier I Water Assessment</b>	29
Summary of Drinking Water Exposure	30
Tier I Model Estimated Residues for Surface Water	33
<b>Tier II Water Assessment</b>	35
Tier II Surface and Ground Water Estimates for Selected Crops	
PRZM-EXAMS Model Estimated Residues	36
Limitations of Modeling Analysis	37
<b>Monitoring Data</b>	
Agricultural Uses	39
Urban/Residential Uses	42
Drinking Water	47
Limitations of Monitoring Analysis	
<b>Groundwater Assessment</b>	49
<b>Ecological Effects Hazard Assessment</b>	
<b>Mode of Action for Malathion</b>	50
<b>Toxicity To Terrestrial Animals</b>	
Birds: Acute Oral and Subacute Dietary Toxicity	51
Avian Reproduction Effects	52
Non-guideline Laboratory Studies with Birds	53
Field Study Observations-Adverse Effects to Birds	53
Mammals: Acute, Subchronic, and Chronic Toxicity	57
Non-Guideline Laboratory Studies with Mammals	58
Toxicity to Reptiles	59
Beneficial Insect Toxicity:	59
Terrestrial and Adult Stages of Semi-Aquatic Insects	
Field Study Observations of Adverse Effects to Insects	60
Terrestrial Field Incidents	62
<b>Toxicity to Aquatic Animals</b>	
<b>Vertebrates</b>	
Freshwater and Estuarine/Marine Fish:Acute Toxicity	63
Freshwater and Estuarine/Marine Fish:Chronic Toxicity	65
Amphibians: Acute and Chronic Toxicity	66
<b>Invertebrates</b>	
Freshwater and Estuarine/Marine Invertebrates:Acute	67
Aquatic	
Semi Aquatic:Larval Life Stages	
Freshwater and Estuarine/Marine Invertebrates:Chronic Toxicity	68
<b>Non-Guideline Laboratory Studies-Aquatic Organisms</b>	69
<b>Field Observations of Adverse Effects to Aquatic Organisms</b>	

Field Studies	73
Aquatic Incident Reports	77
<b>Toxicity To Plants</b>	80
<b>Toxicity of Degradates and Impurities</b>	80
<b>Toxicity of Dual Active Formulations</b>	81
<b>Ecological Risk Assessment</b>	
Risk to Nontarget Terrestrial Animals	83
Birds, Reptiles and Mammals - Calculated Risk Quotients	
Insects: Terrestrial and Semi-Aquatic Adults	94
Risk to Nontarget Aquatic Organisms	
Fish and Invertebrates: Adult Stages-Calculated Risk Quotients	95
Fish and Invertebrates: Early and Full Lifestage -Risk Quotients	96
<b>Endangered Species</b>	97
<b>Risk and Exposure Characterization Summary and Conclusions</b>	99
Expected Paths of Potential Exposure for Wildlife	99
Potentially Exposed Terrestrial and Aquatic Habitats from Malathion Uses	
104	
Inadequacies or Weaknesses of Data	107
Characterization of Predicted Effects to Non-Target Organisms	108
<b>References</b>	117
<b>Appendices</b>	
<b>Appendix 1</b> Data Requirements for Malathion	123
<b>Appendix 2</b> Tier I Aquatic Modeling Inputs	124
<b>Appendix 3</b> Tier II Aquatic Modeling Inputs	132
<b>Appendix 4</b> Additional Aquatic Monitoring Results	136
<b>Appendix 5</b> Terrestrial Crop Scenario Codes and EEC Estimates	143

**List of Tables:**

Table 1.	Malathion Use-Rates - Crop Scenario Numbers
Table 2.	Malathion Non-agricultural Use Summary
Table 3.	Literature: Estimated Soil half-lives
Table 4.	EECs on Mammalian and Avian Food Items
Table 5a.	Foliar residue half-lives Willis and McDowell
Table 5b.	Terrestrial Day 0 EECs
Table 6.	Residues on Alfalfa (Johansen, 1965)
Table 7.	Tier I Surface and Groundwater Concentrations for Drinking water
Table 8.	Malathion-Malaoxon Conversion in Water Treatment
Table 9.	GENEEC Predicted Aquatic EECs
Table 10.	PRZM-EXAMS and GENEEC comparison
Table 11.	Field Runoff-Cotton Bollweevil Program
Table 12 a,b,c	Spray drift to Surface water -Cotton Bollweevil program
Table 13 a	California Medfly Program-Spray drift to water
Table 13 b.	Malathion in Florida Ponds from direct or indirect treatment
Table 14.	Water Treatment and Malathion-Malaoxon conversion
Table 15.	Avian Acute Toxicity
Table 16.	Avian Subacute Dietary Toxicity
Table 17.	Avian Reproductive Sensitivity
Table 18.	Mammalian Acute Oral and Chronic Dietary Toxicity
Table 19.	Nontarget Pollinator Insect Acute and Foliar Contact Toxicity
Table 20.	Aquatic Insect Larvae Acute Toxicity
Table 21.	Terrestrial Field Incidents
Table 22.	Freshwater and Marine / Estuarine Fish Acute Toxicity
Table 23.	Freshwater and Marine Fish Chronic Toxicity
Table 24.	Freshwater and Marine Invertebrate and Estuarine Invertebrate Acute Toxicity
Table 25.	Freshwater and Marine/Estuarine Aquatic Invertebrate Life-Cycle Toxicity
Table 26.	Confirmed and Proposed Malathion Degradates.
Table 27.	Freshwater, Estuarine and Marine Aquatic Incidents.
Table 28.	Toxicity of Mixtures to Birds.
Table 29.	Malathion / Methoxychlor Comparative Toxicities.
Table 30.	Avian Acute RQ Range Table for Shortgrass to Seeds(0.175 to 1.25 lb ai/A)
Table 31.	Avian Chronic RQ Table for Shortgrass to Seeds(0.175 to 1.25 lb ai/A)
Table 32.	Worst Case Scenarios for Dietary Consumption by Small Mammals
Table 33.	Chronic RQ Ranges for Mammals -Exposure to Continuous Residue Peaks
Table 34.	Aquatic Organism Acute Risk Quotients-Invertebrate RQ/Fish RQ
Table 35.	Aquatic Organism Chronic Risk Quotient Table(Invertebrates RQ/Fish RQ)
Table 36.	Prizm Exams Based Aquatic Risk Quotients
Table 37.	Malathion Non Agricultural Use-Worst Case EECs
Table 38.	Maximum Downwind Drift Aquatic EECs and Risk Quotients
Table 39.	Predicted Aquatic Malathion Concentrations with Varying Degrees of Drift
Table 40.	Terrestrial Wildlife Utilization of Major Malathion Usage Areas
Table 41.	Aquatic Habitats and Species Groups - Use Associations
Table 42.	Predicted Aquatic Malathion Concentrations with Varying Application Rates

## **Introduction**

### **Limitations and Agreements With Registrants Regarding the Uses and Application Scenarios Reviewed and Supported**

Registrants who presently intend to support reregistration of malathion products have been contacted and informed that only those use rates presently supported by food tolerances will be reviewed for this Reregistration Eligibility Document. Those crops where tolerances do not exist or where no support from registrants has been received by the Agency are considered revoked and will not be addressed in EFED's risk assessments for Malathion reregistration. Non-food uses of Malathion will be reviewed as has been done with previous REDs. Worst case risk presumptions will be based on the maximum labeled rates, maximum permitted seasonal applications and minimum recommended intervals for these use patterns. The Cheminova and IR-4 Malathion teams have presented proposed uses and application scenarios they intend to support. Any labels which exceed the rates or permitted maximum seasonal applications, or that specify minimum application intervals which are less than the rates presented by these groups will require additional environmental assessment and review.

### **Temporal Uncertainties of Tolerance Test Scenario Use In Risk Assessments**

Assumptions of lower use rates, longer intervals, and limited numbers of seasonal applications based on maximum tolerance tested rates may lead to prediction of lower risk potential than what will occur while mandatory label revisions are in progress. Some malathion labels presently permit application rates well in exceedance of maximum tolerance rates of 6.25 lb ai/A (8.0 to 25.0 lbs ai/acre presently listed on several product labels). The time limits for revision of existing labels will impact the currency of the present assessment for malathion impacts on wildlife and aquatic organisms. This may be tempered by the fact that higher rates, shorter intervals, and unlimited seasonal applications are not as economically practical for the user community and for that reason not common practice. However, given the number of uses and products containing malathion and the number of registrants producing these products, it is expected that application restrictions and mitigation measures may not be implemented for several years pending final agreements with registrants.

# Use Characterization

## Historical Perspective

Malathion has been used for 45 years and is manufactured in over 50 locations throughout the world. Though malathion has been manufactured by a variety of companies, the technical formulation portion of the U.S. business was purchased by Cheminova Agro from American Cyanamid in 1991. As of 1994, all technical malathion produced in the U.S. is manufactured, marketed, and distributed by Cheminova.

## Summary of Supported Product Types, Formulations, and Use Scenarios

### Product Numbers, Types and Percent Active Ingredient

Technical malathion is produced by Cheminova Agro for the U.S. market. At this time 63 formulators of pesticide products use malathion as an active ingredient. Approximately 235 malathion products are produced by these 63 formulators. Some of these products also contain active ingredients other than malathion (e.g., methoxychlor).

Product types include technical ingredient for formulation use, emulsifiable concentrates, wettable powders, dusts, and ready to use (RTU) formulations. There are no granular malathion products presently registered, although malathion is used in a variety of bait type formulations or mixed and applied to food baits for attraction of target insects.

The percent of malathion employed in these 235 products varies considerably and ranges from less than 3% for many of the homeowner use RTU's to as high as 96% malathion for ULV formulations.

### Use Scenarios

The majority of agricultural and public health uses involve the mixture or dilution of EC formulations with water for aerial or ground spray application to target areas either as ULV applications, standard ground application sprays, or ground fogs. Wettable powders are generally diluted in 1 to 100 gallons of water and applied to plant surfaces by ground spray methods. Dust formulations are often used for treatment of grain storage areas or the actual grain itself. Ready to use formulations are often used in the home and applied directly to the insect pest or to areas where the pest is observed. Uses of malathion for direct application to livestock is not being supported. Forest uses on public lands are not being supported, though use on tree farms is still permitted.

### Summary of Major Uses Nationwide

Cheminova has divided the malathion market into four major portions; USDA special program use, commercial agriculture, public health use, and home and garden use. Actual tonnage use estimates are considered confidential business information, the percentages of use can be broadly summarized as USDA, 59-61%; General Agriculture, 16-20%; Public Health, 8-15%; and home and garden use, 10%. These percentages of use may vary with fluctuations in pest pressure or concerns for public health which might occur following natural disasters, e.g. a hurricane.

#### Summary of USDA Special Program Uses

The Boll weevil Eradication Program for cotton crops in the lower states account for the greatest proportion of use by USDA as well as the nation as a whole. Over 97% of USDA's uses of malathion have been towards efforts to eradicate the Mexican boll weevil from U.S. cotton crops. In 1997 USDA applied over 10 million pounds of malathion active ingredient primarily by aerial ULV methods to areas of infestation in Texas and Alabama. Large portions of Texas, Louisiana, Mississippi, and Alabama are targeted for 1998. States where heavy infestations have been eradicated, but lower preventative uses are sometimes still employed, include California, Arizona, Georgia, South Carolina, North Carolina, and Virginia. In general, 10-12 oz. of product (about 1.0 lb of active ingredient) are aerially applied 6-8 times per acre before, during and after the cropping season.

Other programs comprising about 2% of USDA malathion use include the Mediterranean Fruit Fly Control Programs in Florida and California and Grasshopper Control on Federal Rangelands in the western states. Nationwide USDA use has ranged from 80,000 to 360,000 lbs of ai for these programs depending on pest sightings or pressure. Medfly applications generally involve mixture of malathion into a hydrolysate bait at about a 20% malathion concentration which is then applied two to four times at 0.175 lb ai/acre by aircraft or ground spray equipment.

#### Summary of Public Health Uses

Mosquito control in populated and rural areas comprises the major use of malathion for public health uses. In general, public health use would be necessitated by actual disease outbreak caused by a particular pest. Though mosquito control does potentially prevent such outbreaks (particularly after hurricanes or major storms) much of the application is also performed for economic reasons. This type of use is particularly heavy near coastal resort areas, where high bite rates constitute a potential loss of tourist revenue and therefore an incentive for active spraying programs. High use of adulticides is generally an indication that larvicidal uses of pesticides during the early breeding season have failed to control population outbreaks.

#### Summary of Supported Agricultural Crop Uses

Approximately 100 food crop uses of malathion are being supported by Cheminova and IR-4. In the past many labels have often **not** reflected the application rates, intervals, and maximum numbers of application on which acceptable tolerance levels were based. Many of the over 250 product labels were written with no specified intervals or limitations on the numbers of applications which could be made to crops. In an effort to standardize the acceptable label rates for reregistration purposes, Cheminova and IR-4 have agreed to specify maximum rates, minimum intervals, and maximum seasonal application numbers that they support for food uses (see table which follows). Crop scenario codes are indicated by letters for each separate application rate (lbs ai/A), followed by numbers for maximum proposed multiple applications, and (number of days) for the intervals proposed between applications. Thus A10 (7D) indicates an application of 0.175 lbs ai/A is permitted 10 times during the growing season with a minimum interval between applications of 7 days. Many crops may fall under the same application scenario or in some cases only one crop is proposed for that particular scenario. These scenarios are later used in EEC and Risk Quotient tables and allow the reader to associate each value with a particular crop or use pattern.





Table 1.

**Malathion Use Rate Table - Crop Scenarios**  
**Cheminova and IR4 Supported Maximum Tolerance Rates**

Number of Applications													
	Rate lb ai/A	Int. Day	1	2	3	4	5	6	7	8	9	10	12- 25
A	0.175	7D	A1									A10(7D)	
B	0.50	NA	B1										
B	0.61	5D/	C1				C5(5D)						
C	0.61	7D		C2(7D)	C3(7D)								
C	0.61	14D		C2(14D)									
D	0.76	10D	D1				D5(10D)						
E	0.94	3D	E1			E4(3D)							
E	0.94	6D						E6(6D)					
E	0.94	7D			E3(7D)								
F	1.0	7D	F1					F6(7D)					
G	1.25	3D	G1	G2(3D)				G6(3D)					G25 (3D)
G	1.25	5D					G5(5D)						
G	1.25	7D			G3(7D)	G4(7D)	G5(7D)	G6(7D)	G7(7D)	G8(7D)	G9(7D)	G10(7D)	
G	1.25	14D		G2(14D)									
H	1.50	7D			H3(7D)		H5(7D)						
I	1.56	7D		I2(7D)				I6(7D)					
J	1.88	5D					J5(5D)						
J	1.88	7D			J3(7D)	J4(7D)		J6(7D)					
J	1.88	14D		J2(14D)									
K	2.03	6D						K6(6D)					
K	2.03	7D			K3(7D)	K4(7D)							
L	2.5	3D											L25 (3D)
L	2.5	5D			L3(5D)								
L	2.5	7D			L3(7D)		L5(7D)						
M	3.43	5D					M5(5D)						

N	3.75	7D				N4(7D)		N6(7D)					
N	3.75	14D				N4(14D)							
	Rate lb ai/A	Int. Day	1	2	3	4	5	6	7	8	9	10	12- 25
O	4.7	30D		O2(30D)									
P	5.0	7D			P3(7D)	P4(7D)							
Q	6.25	30D			Q3(30 D)								

- 0.175 lb ai/A A10=Orange, Grapefruit, Lemon, Lime, Tangerine, Tangelo, and Kumquat  
0.50 lb ai/A B1=Flax  
0.61 lb ai/A C5(5D)=Sweet Corn , C2(7D)=Hops, C3(7D)=Beans, Corn, Rice, Sorghum, Wheat, and Rye  
C2(14D)=Alfalfa, Clover, Lespedeza, Lupine and Vetch  
0.76 lb ai/A D5=Blueberry  
0.94 lb ai/A E1(3D)=Grass for hay, E4(3D)=Mushroom, E6(6D)=Strawberry, E3(7D)=Pepper and  
spearmint, E7(7D)=Macadamia  
1.0 lb ai/A F6(7D)=Melons, Watermelon, Pumpkin and Squash  
1.25 lb ai/A G1(3D)=Grass for hay, G2(3D)=Field corn , G6(3D)=Mustards, G25(3D)=Cotton,  
G5(5D)=Watercress, G3(7D)=Rice, Sorghum, Wheat, Rye, Barley, Oats and Corn,  
G4(7D)=Blueberry( ULV), G5(7D)=Turnip, Broccoli, Apple, Sweet Corn, Beet, Chayote,  
Horseradish, Parsnip, Radish, Rutabaga, Salsify, Sweet potato, G6(7D)= Cabbage and  
Cherry(ULV), G7(7D)=Carrot , G8(7D)=Mango and Passion fruit , G9(7D)=Asparagus  
G10(7D)=Pears and Quince , G12(7D)=Guava and Papaya, G2(14D)=Alfalfa, Clover, Lupine,  
Vetch nd Lespedeza  
1.5 lbs ai/A H2(7D)=Celery, H6(7D)=Okra  
1.56lbs ai/A I2(7D)=Potato, I5(7D)=Onion, Garlic, Shallot, Leeks  
1.88 lb ai/A J6(5D)=Lettuce, J4(7D)=Blackberry, Raspberry, Loganberry, Boysenberry, Dewberry, Currant,  
Gooseberry, J3(7D)=Cucumber, J6(7D)= Strawberry, J2(14D)=Grapes  
2.03 lbs ai/A K6(6D)=Strawberry(50%WP), K3(7D)= Spinach, Dandelion, Endive, Parsley and Swiss Chard,  
K4(7D)=Blackberry, Raspberry, Gooseberry, Loganberry, Dewberry, Currant and Boysenberry  
2.50 lb ai/A L25(3D)=Cotton, L3(5D)=Figs, L3(7D)=Mustards, Walnuts, and Pecans, L5(7D)=Peas  
3.43 lb ai/A M5(5D)=Tomato, Pepper, Eggplant  
3.75 lb ai/A N4(7D)=Apricots, N6(7D)=Cherry, N4(14D)=Peach and Nectarine  
4.7 lb ai/A O2(30D)=Avocado  
5.0 lb ai/A P3(7D)=Pineapple, P4(7D)=Chestnuts  
6.25 lb ai/A Q3(30D)=Oranges, Grapefruit, Lemon, Lime, Tangerine and Tangelo

### Commercial Non Food Crops

Malathion is presently supported for registration on several non-food crops. These crops include ornamental flowering plants, ornamental lawns and turf, nursery stock plants, ornamental woody plants, pine seed orchards, Christmas trees(commercial), and slash pine.

Previous non-food crop uses which are not being supported by Cheminova or IR-4 include forest uses on douglas fir and spruce(for spruce budworm), hemlock(for hemlock looper), pine trees(for european pine sawfly & Saratoga spittlebug) and larch(for larch casebearer). No

registrant has indicated support for reregistration of malathion for tobacco and therefore it is assumed this use will be revoked.

**Non-Crop Uses-Agriculture Related**

Storage of Grains: Malathion 6% dust formulation is being supported for treatment of stored corn, wheat, barley, oats and rye.

Livestock Feed Lots and Holding Pens: Though direct applications to livestock are no longer supported, treatments of holding facilities and feed lots are still being supported. These treatments are primarily for control of flies and mosquitoes and may be used as sprays or in bait formulations.

Beet Leafhopper Control-Non-Agricultural Lands: This program is confined to specific target areas on non-agricultural lands to control the spread of this pest. The use appears on the Fyfanon ULV and 8EC labels (page 2).

Grasshopper Control Non-Federal Lands: These programs are carried out to control aphids, grasshoppers, and leafhoppers in pasture and rangeland. The Fyfanon 8EC label specifies a mixture of malathion with diesel fuel.

Fly Control-Cull Fruit and Vegetable Dumps: Fyfanon labels contain this use to control drosphilia flies and dried fruit beetles around vegetable and fruit dumps. The 8EC label specifies application as a concentrated drench.

**Summary of Other Non-agricultural Uses**

**Commercial Use Urban Scenarios**

Homeowner Use: Malathion is formulated into numerous home and garden sprays for non commercial use on vegetables, fruit trees, ornamental plants and flowering shrubs, turf, and outside structures. Many are ready to use sprays or dusts and many are combined with other active ingredients.

Use Around Commercial Buildings and Warehouses

Malathion is used to treat for a number of nuisance pests around warehouses, storage yards, inside buildings, outside perimeters of commercial buildings, etc

Table 2.

**MALATHION NON AGRICULTURAL USE SUMMARY**

<b>USE LOCATION WHERE RATE APPLIES (MAX RATE in lbs ai/A)</b>	<b>Max. Rate</b>	<b>Max # Applic.</b>	<b>Min. Interv.</b>	<b>Predicted U.S. Acreage (EPA/OPP)</b>
<b>Nonagricultural rights of way/fencerows/hedgerows</b>	0.598	NS	NS	17,000
<b>Mosquito Control</b> Lakes/Ponds/Reservoirs(human use)(0.5985) Nonag. Uncultivated Areas/Soils (0.6) Polluted Water (0.6) Lakes/Ponds/Reservoirs (No Human Use) (0.628) Swamps/Marshes/Wetlands/Stagnant Water (0.628) Intermittently Flooded Areas/Water (0.628)	0.630	NS	NS	8,227,000
<b>Woodland Use</b> Pine Forest/Shelterbelt (0.9375) Eastern White Pine (Forest) (0.9375)	0.94	NS	NS	17,000
<b>Rangelands/Pastures/Set Aside Acreage/Summer Fallow</b> Canarygrass (1.2) Rangeland or Pastures (1.25) Grass Forage/Fodder/Hay (1.25)	1.25	NS	NS	1,625,000
<b>Ornamental Plant Uses-Nurseries-Homeowner</b> Ornamental trees and Herbaceous Plants	1.746	NS		175,000
Ornamental Nonflowering Plants Ornamental Woody Shrubs and Vines (2.5)	2.50	NS	NS	
<b>Commercial Tree Production</b> Christmas Tree Plantations, (3.125) Ornamental and/or Shade Trees (3.125) Slash Pine (forest) (3.125)	3.125	NS	NS	No estimate provided
<b>Public Parks</b>				67,000
<b>Turf Use/ Golfcourses/Commercial Lawncare</b> Ornamental Lawns and Turf	5.1	NS	NS	35,000 (golfcourses + cemeteries) Commercial landscape= no estimate provided
<b>Total Non-Ag Use per Year-4,100,000 lbs ai/year (USEPA OPP/BEAD estimates)</b>				

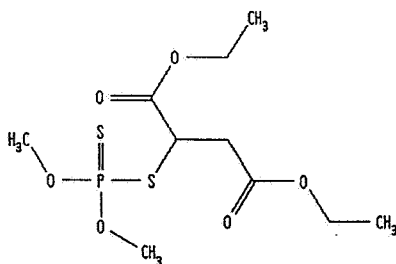
NS=Not Specified

# Exposure Characterization

## Environmental Fate and Chemistry of Malathion

### A. Chemical Profile

#### O,O-Dimethyl phosphorodithioate of diethyl mercaptosuccinate



#### Physical/Chemical properties:

<b>Molecular formula:</b>	<b>C<sub>10</sub>H<sub>19</sub>O<sub>6</sub>PS<sub>2</sub></b>
<b>Molecular weight:</b>	<b>330.3 g/mol.</b>
<b>Physical state:</b>	<b>Clear amber liquid.</b>
<b>Melting point:</b>	<b>2.85° C.</b>
<b>Boiling point (0.7 mm Hg):</b>	<b>156-157° C.</b>
<b>Specific gravity (25° C):</b>	<b>1.23</b>
<b>Vapor pressure (30° C):</b>	<b>4 × 10<sup>-5</sup> mm Hg.</b>
<b>Solubility (25°C):</b>	<b>145 mg/l water.</b>

### B. Environmental Fate Summary

Wildlife and humans may be exposed to malathion and its degradates through contamination of food, water, and air (by suspended particles) which can result from off-target drift, runoff, and direct application. Increased toxicity may be brought about through oxidation (to malaaxon) and isomerization (to isomalathion). Limited data is available on toxic degradates and impurities, but the fate data on malathion is acceptable and shows little persistence. Based on registrant data and open literature reports, EFED concludes that the primary route of dissipation of malathion in surface soils appear to be microbially mediated soil metabolism (half-life <1-2.5 days) and hydrolysis (pH 7 half-life 6.21 days and pH 9 half-life 12 hours) with malathion monoester, ethyl hydrogen fumarate, diethyl thiosuccinate, malathion mono- and dicarboxylic acids, demethyl mono- and dicarboxylic acids, and CO<sub>2</sub> as known degradates.

Other important routes of dissipation from soil suggested by the data include leaching, plant uptake, and surface runoff. Malathion and its degradates in general are soluble and do not adsorb strongly to soils. Data from the field dissipation studies indicate that malathion dissipates rapidly

when applied in the field. Although the anaerobic and aerobic aquatic metabolism and aquatic field dissipation studies indicate very rapid degradation ( $T_{1/2} = <2.5$  days), the metabolism studies were performed under alkaline conditions that favor hydrolysis and the water pH in the dissipation study was not specified. As such it is difficult to separate out the effect of hydrolysis from metabolism.

Malathion is stable to hydrolysis at pH 5 ( $T_{1/2} = 107$  days), to aqueous photolysis ( $T_{1/2} = 94$  and 143 days, corrected for dark control) and soil photolysis ( $T_{1/2} = 173$  days) and does not volatilize appreciably ( $\leq 5.1\%$  of applied volatilized after 16 days). Open literature studies suggest persistence on soil is longer under dry, sandy, low nitrogen, low carbon, and acidic conditions (Walker and Stojanovic 1973).

Acceptable leaching data on parent malathion indicate that it is mobile in all soils tested ( $K_{ds} = 0.82-2.47$ ). Acceptable terrestrial field dissipation data indicate rapid dissipation ( $T_{1/2} = <2$  days). One detection of malathion below 12 inches was found in a terrestrial field dissipation study, indicating leaching as a likely route of dissipation. Similarly, column leaching studies demonstrated that malathion and its degradates, malathion mono- and dicarboxylic acids are very mobile in soil. Data presented to the Agency and in the "Pesticides and Groundwater Database"(U.S. Environmental Protection Agency 1992) demonstrate that malathion has the potential to leach to ground water. Malathion has been detected in ground water in three states (California, Mississippi, and Virginia) at levels ranging from 0.03 to 6.17 ppb. Based on these data and the low  $K_d$  values for malathion it is clear this chemical has the potential to leach to ground water.

Some fate and toxicity data are also available on other malathion degradates and impurities. Technical malathion contains impurities that account for up to 5% of the insecticide. CalEPA has reported 15 impurities present in a representative ultra low volume malathion formulation. These impurities include:

- diethyl fumarate (0.90%)
- diethylhydroxysuccinate (0.05%)
- O,O-dimethylphosphorothioate (0.05%)
- O,O,O-trimethyl phosphorothioate (0.45%)
- O,O,S-trimethyl phosphorodithioate (1.20%)
- Ethyl nitrite (0.03%)
- Diethyl-bis (ethoxycarbonyl) mercaptosuccinate (0.15%)
- S-1,2-ethyl-O,S-dimethyl phosphorodithioate [isomalathion] (0.20%)
- S-(1-methoxycarbonyl-2-ethoxycarbonyl)ethyl-O,O-dimethyl phosphorodithioate (0.60%)
- Bis-(O,O-dimethyl thionophosphoryl) sulfide (0.30%), Diethyl methylthiosuccinate (1.00%)
- S-ethyl-O,O-dimethyl phosphorodithioate (0.10%)
- S-1,2-bis (ethoxycarbonyl) ethyl-O,O-dimethyl phosphorothioate [malaoxon] (0.10%)
- diethyl ethylthiosuccinate (0.10%)
- sulfuric acid (0.05%).

Several of these organophosphate impurities potentiate malathion toxicity and also are toxic alone, but there is almost no data available on their environmental fate. The persistence of one highly toxic phosphorothioate impurity was shown to be 18.7 times longer than malathion in a aerobic soil metabolism study (Miles and Takashima 1991). Phosphorothioates and -dithioates have been intensively studied and induce a delayed toxic effect to mammals at much lower levels than pure malathion (Ali Fouad and Fukuto 1982, Umetsu *et al* 1977, Fukuto 1983, Aldridge *et al* 1979, Toia *et al* 1980). One hydrolysis product, diethyl fumarate (sometimes referred to as DEF), which is also present as an impurity in technical malathion is approximately 3 times more toxic to fathead minnows than malathion (Bender 1969). No registrant submitted fate data and little open literature data exist for impurities of malathion.

The relative concentration of malathion impurities can vary dramatically depending on manufacturing processes and storage conditions. Umetsu *et al* (1977) concluded:

"Storage of technical malathion for 3 to 6 months at 40 degrees C resulted in materials which were noticeably more toxic to mice."

"Needless to say, malathion should not be stored for prolonged periods under conditions where it is subjected to consistently high temperatures."

Regarding the impurities present in formulations from different sources of malathion, Mulla *et al* (1981) state:

"One of these toxic impurities was isomalathion which was 0.3% in brand I (U.S.), 2.1% in brand II (Italian), and 3.1% in brand III (Italian)."

Thus, the composition and toxicological properties of the technical product is affected by initial quality and storage conditions.

One impurity of malathion is the oxon analog, mala~~oxon~~, which is also the active acetylcholinesterase inhibiting component *in vivo*. Under some dry and microbially inactive environmental conditions mala~~oxon~~ is formed from malathion at levels up to 10.7% of the total applied (CalEPA 1993). Monitoring studies conducted during medfly control programs show high levels of mala~~oxon~~ (greater than 328 ppb) in runoff water (CDFG 1988). EFED does not have a complete environmental fate database for mala~~oxon~~ but based on its chemical similarity to malathion (sulfur is replaced by oxygen), the parent and its degradate are expected to have similar chemical properties. The aerobic half-life of mala~~oxon~~ has been reported as 3 and 7 days in basic and acidic soils, respectively (Paschal and Neville 1976). This longer half-life relative to malathion is proposed to be a result of mala~~oxon~~'s biocidal effect on soil microbes which contribute to malathion's degradation.

### Requirements

Data for the aerobic aquatic metabolism studies are considered supplemental at this time. The deficiency of this study is the alkaline pH of the soil and water used. Degradation of malathion is highly pH dependent, with faster rates at higher pH. Thus, a quantitative assessment of malathion fate and persistence under acidic conditions when hydrolysis would be slower cannot



be conducted. As a result, EFED cannot complete a quantitative assessment of the environmental fate of malathion and its degradates in acidic environments it is likely to contact. An aerobic metabolism study (162-4) performed under pH conditions that do not favor hydrolysis is requested as EFED believes this additional information will enable a more quantitative assessment of the fate and persistence of this major use chemical in acidic aquatic environments. This request is especially relevant given the sensitivity of numerous aquatic organisms to malathion.

Although the anaerobic aquatic metabolism study was also conducted under alkaline conditions favoring hydrolysis, EFED believes that hydrolysis data along with open literature data on the persistence of malathion in sediments is sufficient to conclude that malathion will not persist under anaerobic conditions. However, an anaerobic aquatic metabolism study under acidic conditions may be requested if more quantitative data on malathion degradation and degradates is needed.

Fate data is required for malaoxon, the oxidation product of malathion. Malaoxon is commonly believed to be the neuroactive toxic agent of malathion after oxidation *in vivo* and toxicity data show it to have higher acute toxicity than malathion. EFED acknowledges that maximal malaoxon conversion under registrant submitted study conditions was low (1.8%) however under other conditions encountered during malathion use conversion levels as high as 10.7% of applied insecticide have been reported (CalEPA 1993). HED also has indicated that malaoxon is to be included in the tolerance expression for malathion. First tier surface water assessments were performed by making several assumptions about the properties of malaoxon relative to malathion. It is not possible to perform second tier assessments without further information specific to malaoxon; thus, EFED requests data required to predict malaoxon levels in drinking water and aquatic habitats.

In addition to data on the basic physical properties of malaoxon (solubility, partition coefficient, vapor pressure), EFED requests that the following laboratory studies be submitted for malaoxon based on the brief justification provided. Data from these studies are expected to be sufficient to perform basic fate and exposure modeling of malaoxon.

#### Degradation

##### 161-1 (hydrolysis)

Malathion hydrolysis is an important route of dissipation under alkaline conditions. The phosphorothiolate ester bond of malaoxon may be more susceptible to cleavage via hydrolysis than the analogous phosphorodithioate ester in malathion.

#### Metabolism

162-1 (aerobic soil) The primary route of malathion degradation on soil is through aerobic metabolism. An open literature study (Paschal and Neville 1976) suggests malaoxon persistence may be greater on soils. Additionally, CalEPA studies have shown levels of malaoxon production exceeding 10% in certain dry, low organic content soils.

##### 162-4 (aerobic aquatic)

Although little or no malaoxon production is observed in registrant submitted aquatic studies malaoxon has been detected in surface waters and the potential for malaoxon runoff may be heightened relative to malathion because it is expected to have higher solubility. Aerobic aquatic metabolism contributes greatly to malathion degradation.

### Mobility

#### 163-1(leaching/adsorption)

EFED is not aware of reports of malaoxon groundwater contamination. However, malathion has contaminated groundwater in several states and has the potential to contaminate surface water through runoff. The increased polarity of malaoxon due to the substitution of oxygen for sulfur increases the expected potential of this chemical to be mobile in soil.

EFED also requests additional information on environmental malaoxon production. Because malathion is used in a large number of settings including more than 60 terrestrial field uses as well as outdoor residential uses including mosquito, Mediterranean fruitfly, and urban pest control uses, it is exposed to a large variety of environmental conditions. This extensive use is likely to result in significant exposure of nontarget organisms to malathion breakdown products. Exposures to humans and wildlife may be through contamination of food, water, and air (by suspended particles) which can result from off-target drift, runoff, and direct application.

It is clear that under many circumstances malathion degrades rapidly to compounds of lower toxicity, usually through microbial metabolism and hydrolysis. However, in residential uses (*e.g.* aerial and ground application for mosquito control), it is likely that malathion will contact dry, microbially inactive and low organic content surfaces such as concrete, asphalt, dry soil, roofing material, and glass. It is expected that malaoxon production will be increased on these surfaces as malathion is exposed to air for extended periods until it is washed away by rain. This is supported by malaoxon monitoring data in urban streams after residential malathion treatment showing similar or higher levels of malaoxon than malathion in some instances (State of California Department of Food and Agriculture, 1982). Thus EFED proposes that malathion persistence and degradation on anthropogenic surfaces be examined (suggestions from the registrant are invited for particular surfaces to be examined). The State of California EPA has published two studies describing adequate methods for determining malaoxon production on dry soil (CalEPA 1993) and steel sheets (CalEPA 1996) which would be amenable to other abiotic surfaces. Both of these studies showed higher malaoxon production than registrant submitted studies, but maximal levels of malaoxon production were not achieved. On the steel surface a rainfall event removed most of the malathion after only 2 days. On the dry soil malaoxon production did not decrease by the time the study was terminated at 22 days. Runoff of residential malathion and malaoxon greatly increases risk of human and aquatic wildlife exposure through drinking water and habitat contamination and increases the need for this information.

The State of California EPA has published a study describing malaoxon production on low organic content soil (0.6%) with a moisture content less than 1% (CalEPA 1993) showing higher malaoxon production than registrant submitted studies using soils with higher organic (2-2.7%) and moisture (75% of water holding capacity, capacity not stated) content. From the CalEPA data it appears that malaoxon production is favored on dry soils and thus may represent a higher risk scenario for malaoxon production and runoff. EFED believes that data on dry soils may be useful to assess malathion and malaoxon fate and persistence in some use settings which are not ideal for malathion degradation, thus EFED requests the submission of data on malathion degradation and malaoxon production in an aerobic soil metabolism study (162-1) using a soil with a low moisture content (<1%) and low organic content (<1%).

### Conclusions

Malathion is generally nonpersistent, however EFED lacks important information to evaluate the behavior of malathion under acidic aquatic conditions which might likely increase its persistence and alter degradates produced. To adequately determine the environmental fate of malathion and its degradates aerobic aquatic metabolism data under acidic soil and water conditions are needed.

Data are also required on the oxidative degradate malaoxon. Presently EFED has no registrant submitted fate data for malaoxon. Monitoring data suggest that malaoxon production is an important issue in residential areas. Thus, EFED is requesting studies of malathion degradation and malaoxon production on anthropogenic surface which 1.) make up much of the residential environment, 2.) increase malathion persistence, 3.) increase malaoxon production, and 4.) increase runoff potential. Acceptable data on malaoxon solubility, hydrolysis, vapor pressure, soil metabolism, and aquatic metabolism in conjunction with information on malaoxon production under high risk conditions will aid EFED in predicting environmental malaoxon concentrations for ecological and human health concerns.

## **Environmental Fate and Transport Studies**

### **Degradation**

#### **Hydrolysis:**

Several open literature studies (Mulla *et al* 1981, Howard 1991) are consistent with data presented by the registrant showing that malathion is unstable under alkaline conditions and increasingly stable under acidic conditions. Hydrolysis products characterized by the registrant and in the open literature are similar as well. Muhlman and Shrader (1957) report primary products of diethyl fumarate and dimethyl phosphorodithioic acid in base while the registrant's study identifies malathion monocarboxylic acid and ethyl hydrogen fumarate. The reported differences are relatively minor as ethyl hydrogen fumarate is an expected hydrolysis product of diethyl fumarate and the other products are also expected.

In the registrant's study [<sup>14</sup>C]malathion was hydrolytically stable in aqueous buffered pH 5 (half-life = 107 days) solutions. At pH 7 and 9 solutions malathion hydrolyzed relatively rapidly with half-lives of 6.21 days and 12 hours, respectively. Parent malathion accounted for 80.3% and 3.7% of applied after 28 days in the pH 5 and 7 solutions, respectively; while after 53 hours of hydrolysis at pH 9, only 3.6% of applied was parent malathion.

The hydrolysis data requirement is satisfied. (MRID 40941201, 43166301)

#### **Photodegradation in water:**

Open literature reports and registrant submitted data suggest that photodegradation in natural and distilled water is relatively slow. Open literature half-life data ranges from 0.67 (natural river water) to 42 days (distilled water) (Howard 1991).

In the registrant submitted aqueous photolysis study, [2,3-<sup>14</sup>C]malathion degraded with calculated half-lives (corrected for dark control degradation) of 94 and 156 days in sterile pH 4 photosensitized (1% acetone) and nonsensitized aqueous buffer solutions, respectively, that were irradiated continuously with a xenon arc lamp at  $25 \pm 1$  °C for 30 days. Detections of monoethyl maleate, diethyl maleate, malaoxon, mono-acid, diethyl mercaptosuccinate, and diethyl fumarate were noted at concentrations <<10% of applied.

The photodegradation in water data requirement is satisfied. (MRID 41673001, 43166301)

#### **Photodegradation on soil:**

Registrant and open literature data suggest malathion is stable under sunlight. Open literature on photodegradation of malathion in thin films on glass showed production of 6 degradates but at a maximum level of 0.01% after 16.5 hours (Chukwudebe *et al* 1989).

In the registrant submitted soil photolysis study, [2,3-<sup>14</sup>C]malathion degraded with a registrant-calculated half-life of 173 days. The soil used was a pH 6.5 sandy loam. No degradates at concentrations  $\geq 10\%$  were observed. Thus, soil photolysis is not an important route of dissipation for malathion.

The photodegradation on soil data requirement is satisfied at this time (MRID 41695501, 43166301), but EFED requests that the registrant provide an explanation for the long persistence of malathion in irradiated and dark control samples. Soils were stated to possess microbial activity which would lessen persistence making it unclear why malathion was stable under the study conditions.

#### **Photodegradation in air:**

The relatively low vapor pressure of malathion suggests gas phase reactions are only minor routes of degradation. However, malathion in the form of very small droplets (~10 µm), such as might result from ULV formulation application, may reach much higher concentrations in air. No open literature references on photodegradation in air were found.

In a the registrant supported photolysis in air study, >10% [2,3-<sup>14</sup>C]malathion, contained in Tedlar bags, degraded during 4 days of irradiation. The photolysis in air data requirement is not satisfied, however, EFED agrees with the registrant that the study should not be required for several reasons. In the laboratory volatility study, less than 5% of malathion was volatilized from soil surfaces, indicating that obtaining a significant vapor flux that can be experimentally studied for true vapor photolysis is inherently difficult. In addition, the UV absorption spectrum of malathion shows an onset of absorption at about 260 nm, well below the 290 nm cutoff for natural sunlight irradiation and for the xenon light source used in the study. Thus, photodegradation of malathion in air is not expected. This conclusion is also supported by the lack of photodegradation in either the aqueous photolysis or the soil photolysis studies. Finally, as noted in the Rejection Rate Analysis (EPA 1993), this study often generates data that are difficult to interpret, and EPA has dramatically reduced the instances in which this study is required.

## Metabolism

### **Aerobic Soil Metabolism:**

Aerobic soil metabolism is an important route of malathion degradation. Malathion persistence under aerobic soil conditions has been examined in several open literature studies which are reviewed in Table 3. Reported half-life values (from field and laboratory studies) vary from hours to 11 days. Persistence is decreased with microbial activity, moisture, and high pH.

**Table 3.**

Source	Degradation Rate Value	Comments
Miles and Takashima 1991	t <sub>1/2</sub> = 8.2 h (laboratory) t <sub>1/2</sub> = 2 h (field)	malathion was mixed with Lihue soil and incubated at 22°C in lab experiment. Sterilization decreased rate 2-fold.
Walker and Stojanovic 1974	47-95% at 7 days	malathion was incubated with various <i>Arthrobacter</i> species. Degradation in the presence of the 5 most efficient species was reported.

Walker and Stojanovic 1973	$t_{1/2} = \sim 2$ days under non-sterile unfavorable degradation conditions.	Three Mississippi soils were examined at 25-26°C. Soil microflora were important in degradation. Slowest degradation occurred in soils with low nitrogen, moisture, carbon content and increased acidity.
CalEPA 1994	DT50 = 4.2-6.9 days on sand	Measured at five sites under the conditions of the medfly eradication program. Each site consisted of 10 aluminum trays containing 500g of playground sand. Between applications trays were covered.
CalEPA 1993	DT50 < 12 h on sand	Application was under controlled conditions but temperature was not noted.
CalEPA 1993	soil: 38% remaining at 12 hours 15% remaining at 20 days	66% sand, 24% silt, 10% clay, 0.78% water, pH 6.3. Malathion was applied under controlled conditions. Degradation was biphasic.
Kearney <i>et al</i> 1969	75-100% degradation in 1 week	Field persistence
Lichtenstein and Schultz 1964	85% dissipation in 3 days	Conducted under field conditions
Handbook of Environmental Fate and Exposure Data for Organic Chemicals 1991	Reported average literature $t_{1/2} = 6$ d	In this review persistence is stated to vary with moisture content and pH.
USDA	$t_{1/2} = 3$ days used for modeling	This value was chosen for modeling malathion in the boll weevil eradication program based on a personal communication with a previous malathion registrant.

In the registrant submitted study [2,3-<sup>14</sup>C]malathion degraded with a calculated half-life of approximately 0.2 days in two aerobic metabolism experiments using loam soil (pH 6.1) incubated in the dark at 22 ± 2° C and 75% of field capacity. An ancillary experiment was conducted to determine the rate of degradation of malathion in sterile soil. At 4 days posttreatment, malathion comprised close to 100% of the applied radioactivity (97.84% of the extractable radioactivity). This indicates that microorganisms are important in the rapid degradation of malathion in soil under acidic aerobic conditions (MRID 41721701, 43166301).

Numerous degradates were identified in the soil extracts and are identified below as a percent of applied radioactivity: dicarboxylic acid of malathion-18.7-36.7%, the beta monocarboxylic acid

of malathion-2.8-7.3%, the alpha monocarboxylic acid of malathion-1.9-2.5%, and malaaxon-0.6-1.8%.

Soil bound (unextracted) residues averaged 32% of applied at the 6 hour sampling, which increased to 65% at the 24 hour samples and then slowly decreased to 39% after 92 days of incubation. Total  $^{14}\text{CO}_2$  evolution was 45-56% of the applied radioactivity; while volatile organic residues were  $\leq 0.2\%$ .

The aerobic soil metabolism require is considered satisfied (MRID 41721701, 43166301).

#### **Anaerobic Aquatic Metabolism:**

A open literature study (Bourquin 1977) and the registrant's study suggest that malathion persistence in anaerobic environments is short, however, due to the high pH in the registrant's study a quantitative assessment of the degradation and degradation products cannot be performed.

In the registrant submitted anaerobic aquatic metabolism study [2,3- $^{14}\text{C}$ ]- and technical grade-malathion added to a sandy loam soil degraded with a registrant-calculated half-life of approximately 2.5 days in sediment (pH 7.8) and water (pH 8.7). This study provides useful information, but hydrolysis was probably the main route of degradation in the study since the pH of the system was in the basic range which favors hydrolysis. Although most of the residues remained in the water phase (less than 20% of the applied radioactivity was associated with the soil at any sampling interval), the degradation products were similar in both sediment and water phases. The degradation products at maximum concentrations in the water phase were the monocarboxylic acid of malathion (MCA, 28% at Day 4), demethyl monocarboxylic acid (21% at Day 7), dicarboxylic acid (21 % at Day 14) and the demethyl dicarboxylic acid metabolite (39% at Day 45). The degradation products at maximum concentrations in the sediment were the monocarboxylic acid of malathion (4.5% at 6 hours), demethyl monocarboxylic acid (8.1% at Day 45), and dicarboxylic acid (5.2% at Day 4). The EFED calculated half-life for malathion monocarboxylic acid was 11 days.

This study is considered satisfactory with supporting hydrolysis and open literature data suggesting that malathion is unlikely to persist in anaerobic aquatic conditions (MRID 42216301, 43166301). Repeated studies under acidic conditions may be requested if more quantitative data on malathion persistence in these environments are required.

#### **Aerobic Aquatic Metabolism:**

A USGS monitoring study (1998) shows detections of malathion in large rural and urban streams. Many open literature studies have been conducted on the fate and persistence of malathion in the aquatic environment. Reported degradation rates vary and are likely to be significantly increased by biodegradation and pH. Eichelberger and Lichtenberg (1971) found 75% and 90% degradation in river water in one and two weeks, respectively. Guerrant et al

(1970) found malathion half lives in pond, lake, river and other natural waters varied from 0.5 to 10 days and was dependent on pH. Other studies are summarized in Mulla *et al* (1981) and Howard (1991).

Registrant submitted studies were conducted under alkaline conditions which favor hydrolysis. Thus, degradation rate and products may not be representative of acidic aquatic conditions. In the registrant submitted aerobic aquatic metabolism study, a mixture of [2,3-<sup>14</sup>C]- and technical grade-malathion added to a sandy loam soil rapidly degraded in the aerobic aquatic environment with a half-lives of approximately 1.09 days in the water phase (pH 7.8) and 2.55 days in sediment (pH 8.5). As mentioned above, hydrolysis was probably the main route of degradation in the study since the pH of the system was in the basic range and hydrolysis occurs most rapidly at pH 9. Major degradates in water and soil were similar: mono- and dicarboxylic acids of malathion, demethyl monoacid and demethyl diacid, while in sediment no demethyl diacid was detected. The EFED calculated half-life for malathion monocarboxylic acid was 3 days.

This study is considered supplemental at this time (MRID 42271601, 43166301) as quantitative data for malathion degradation in acidic aquatic environments where it is likely to have longer persistence are required. Malathion's high toxicity to aquatic animals and the need to model malathion fate for drinking water assessments in a conservative manner are two important aspects of this assessment which directly rely upon quantitative aquatic fate data.

### **Mobility:**

#### **Leaching/adsorption/desorption:**

The short soil persistence of malathion reduces the risk of leaching to groundwater however it has been detected in the groundwater of at least three states (USEPA 1992). Demethyl and carboxylic acid degradates are expected to be highly mobile particularly in alkaline soils.

Based on batch equilibrium (adsorption/desorption) studies, unaged [<sup>14</sup>C]malathion was determined to be very mobile in sandy loam, sand, loam, and silt loam soils, with respective Freundlich  $K_{ads}$  values of 0.83-2.47; and  $K_{oc}$  values from 151-183. Adsorption was correlated with organic carbon content. Values for  $1/n$  for  $K_{ads}$  were clustered in the range of 0.904-0.978 (MRID 41345201).

Malaaxon was detected in any leachate or soil extracts in concentrations  $\geq 0.12\%$  ( $\geq 6$  ppb) of applied radioactivity. (MRID 43868601, 41345201, 43166301)

#### **Laboratory volatility:**



Three different malathion formulations [Ready To Use (RTU), Ultra Low Volume (ULV), Emulsifiable Concentrate (EC)] added to a silt loam soil did not undergo any appreciable volatilization, when measured under different soil moisture regimes or air flow rates. No more than 5.1% of the applied radioactivity volatilized during the 16 days of the study.

## **Dissipation**

### **Terrestrial field dissipation:**

Open literature studies provide varying rates of terrestrial dissipation. Mulla *et al* (1981) summarizes degradation results from several field studies including: no residues after 6 months (Roberts *et al* 1962), and 85% degradation in 3 days and 97% in 8 days (Lichtenstein and Schulz 1964). The fastest route of terrestrial field dissipation is generally accepted to be via microbial degradation.

In the registrant submitted field dissipation study using a rate of 1.16 lb ai/A, malathion or malaaxon residues were detected at  $\leq 10$  ppb in the 0-6" layer in cotton/bareground sites in GA. Due to the sampling depth it is not possible to determine how much malathion remained at the soil surface relative to that which moved through the first six inches. Residues detected in the plots in the 6-12" layer after the 2nd, 3rd, 4th and 5th treatments averaged 35, 37, 5.6 and 9.4 ppb, respectively. Malathion was detected in the 12-18 inch soil depth at 16 ppb in one replicate soil sample; however, the detection was attributed to contamination. The detection of malathion below six inches along with the low  $K_d$  values reported for malathion make it feasible that leaching below 12 inches may have occurred in the field dissipation studies.

The half-life could not be determined due to the rapid dissipation of malathion, although it is probably  $< 1$  day (MRID 41748901, 43042401, 43166301).

In a field dissipation study located in California, malathion was applied at a maximum rate of 1.16 lb ai/A once a week for 6 weeks. The resulting dissipation half-life was  $< 0.2$  days. In certain instances, malathion was detected below the 12 inch soil depth (MRID 41727701, 43042402, 43166301).

### **Aquatic field dissipation:**

Open literature references detailing persistence in aquatic environments are briefly mentioned under the aquatic metabolism section.

In the first registrant aquatic field dissipation study located in Missouri, malathion was applied at a maximum rate of 0.58 lb ai/A in three weekly applications to a flooded rice paddy (soil pH 6.1, water pH not stated). Malathion residues detected in water samples collected after the first and second application had dissipated below the detection limit (0.01 ppm) in samples taken prior to the second and third applications. In water samples collected one day after the last application,

malathion concentrations averaged 0.017 ppm and had decreased to <0.01 ppm by the second sampling day. Malaoxon residues were <0.01 ppm at all sampling dates.

The data indicate a very rapid dissipation of malathion in water, probably <1 day; however, an accurate half-life could not be determined because of the rapid dissipation (MRID 42058402, 43166301).

In the second aquatic field dissipation study performed in California (soil pH 7.4, water pH not stated), malathion was applied at a rate of 0.58 lb ai/A in three weekly applications to flooded plots. The resulting dissipation half-life could not be determined in the California plot because it was probable that only 1-2% of the intended amount of malathion was applied (MRID 42058401, 43166301).

## **Accumulation**

### **Accumulation in irrigated crops:**

Crop accumulation and residue studies are conducted under the purview of the Health Effects Division. In an accumulation in irrigated crops study located in Missouri, which used the irrigation water from the above mentioned aquatic field dissipation study on rice, malathion or malaoxon did not accumulate in corn, grain sorghum, soybeans or sweet potatoes. The study was unacceptable mainly because the authors did not attempt to determine residues in plants, other than malathion and malaoxon, which were detected in laboratory studies. (MRID 42058402, 43166301).

### **Accumulation in fish:**

Aquatic bioconcentration values ranging from 7.36 (lake trout), 29.3 (coho salmon), 869 (white shrimp), to 959 (brown shrimp) are summarized in Howard (1991).

The registrant submitted study shows [<sup>14</sup>C]malathion residues did not significantly accumulate in bluegill sunfish exposed to 0.99 ppb [<sup>14</sup>C]malathion in a flow-through system for 28 days. Average concentrations of malathion were 3.9 to 18 ppb in the edible fish parts, 21 to 130 ppb for whole fish and 34 to 200 ppb in the nonedible tissue. [<sup>14</sup>C]malathion residue equivalents in the edible fish tissue during depuration ranged from 18 ppb at the start to 4.8 ppb by day 14. Whole fish concentrations decreased from 110 to 4.5 ppb and non-edible fish concentrations decreased from 150 to 5.8 ppb after day 14. Approximately 73, 96 and 96% of the radioactivity depurated from the edible, whole and nonedible portions of fish, respectively. The non-depurated radioactivity consisted of up to 22 other components present in concentrations <10% of total applied radioactivity and were not further identified.

The only significant residue detected in fish tissue was malathion monocarboxylic acid (MCA) in concentrations of 33.3-35.9% (44.8-61.2 ppb) of total radioactive residues (TRR). Up to 22 other components were present in levels of 0.1 to 5.7% (0.1 to 7.7 ppb) and included malathion dicarboxylic acid (MDCA), malaoxon, desmethyl malathion, monoethylfumarate and oxalacetic acid. Malaoxon was present in concentrations  $\leq 2.7$  ppb; while parent malathion was present in concentrations of 0.2 ppb.

Maximum BCFs, as a function of radioactive residues present, ranged from 4.2 to 18, 23 to 135, and 37 to 204 for edible, whole fish and nonedible, respectively (MRID 43106401, 43106402, 43340301).

### **Spray Drift**

No registrant-submitted spray drift studies were reviewed. A study conducted for the Boll Weevil Eradication Program at Penn State (1993) examined malathion drift under conditions of boll weevil control (1 lb/A = 112 mg/m<sup>2</sup>) with an ultra-low volume (ULV) formulation. Deposition up to 21.0, 11.5, 2.9, and 0.7% of that applied was observed at 100, 200, 500, 1000 meters downwind. Due to the size of the particles generated, the ULV formulation is expected to produce the highest levels of drift.

To satisfy spray drift study requirements the registrant, in conjunction with other registrants, formed the Spray Drift Task Force (SDTF). The SDTF has completed and submitted to the Agency its series of studies which are intended to characterize spray droplet drift potential due to various factors, including application methods, application equipment, meteorological conditions, crop geometry, and droplet characteristics. During 1998 EPA plans to complete the evaluation of these studies. In the interim and for this assessment of malathion, the Agency is relying on previously submitted spray drift data and the open literature for off-target drift rates. The simplified rates used are 1% of the applied spray volume from ground applications and 5% from aerial and orchard airblast applications at 100 feet downwind. It is important to note that drift studies on ULV malathion show significantly higher levels of drift. After its review of the new studies, the Agency will determine whether a reassessment is warranted of the potential risks from the application of malathion to nontarget organisms.

The status of the environmental fate data requirements for malathion for terrestrial food crop, terrestrial feed crop, indoor non-food, and residential outdoor uses is summarized in the appendices.

## **C. Terrestrial Exposure Assessment**

### **Terrestrial Vegetation Exposure**

#### **Exposure Concentrations for Nontarget Terrestrial Wildlife and Insects**

For pesticides applied as a nongranular product (e.g., liquid, dust), the estimated environmental concentrations (EECs) on food items following product application are compared to LC50 values to assess risk. The predicted 0-day maximum and mean residues of a pesticide that may be expected to occur on selected avian or mammalian food items immediately following a direct single application at 1 lb ai/A are tabulated below.

**Table 4**

Estimated Environmental Concentrations on Avian and Mammalian Food Items (ppm) Following a Single Application at 1 lb ai/A

Food Items	EEC (ppm) Predicted Maximum Residue <sup>1</sup>	EEC (ppm) Predicted Mean Residue <sup>1</sup>
Short grass	240	85
Tall grass	110	36
Broadleaf/forage plants, and small insects	135	45
Fruits, pods, seeds, and large insects	15	7

<sup>1</sup> Predicted maximum and mean residues are for a 1 lb ai/a application rate and are based on Hoerger and Kenaga (1972) as modified by Fletcher *et al.* (1994).

Residues on Vegetation from Multiple Applications

Predicted residues (EECs) resulting from multiple applications are calculated in various ways. The Agency has employed simple first order dissipation calculations to predict concentrations on vegetative surfaces following spray application at 0.18 to 6.25 lbs ai/acre. The application scenarios are based on maximum tested rates, minimum intervals and maximum numbers of applications tested for establishment of residue tolerances on food. An assumed 90th percentile foliar dissipation half-life of 5.5 days on plant surfaces was derived from averaging of calculated half lives based on 37 reports of malathion residue samples collected on 12 different crops by various researchers from 1957 to 1981 as reported in Willis and McDowell, 1987 (see table below).

**Table 5 a . Foliar Half-life Estimates-Willis and McDowell**

Crop(# samples) Formulation types	Location	Rainfall in mm	Half-life in days
Alfalfa(3) Dust & EC	NC, MA	6.1 to 17.8	0.7, 2.0, and 4.1
Apple(1)	WA	Not reported	3.2 ± 1.2
Chicory(3) Dust, EC, & WP	MD	0	0.7, 0.8, and 1.4
Collards(4) Dust, WP, & EC	NC, MD	0	1.0, 1.4, 1.5, and 1.7
Cotton(5) EC	KY, MS	Not reported	0.3, 0.4, 1.1, 0.6, 6.1
Citrus(1)	FL	Not reported	2.3

Crop(# samples) Formulation types	Location	Rainfall in mm	Half-life in days
Endive(3) Dust, WP, & EC	MD	48.8	1.2, 1.5, 5.9
Leaf lettuce(3) Dust, WP, & EC	MD	0	2.9, 5.8, 6.8
Lima Beans(3) ULV, oil mix	MD	Not reported	1.3, 2.5, 2.8
Kale(1) EC	DC	Not reported	3.1
Turnip(1) EC	FL	60.7	6.4 ± 2.7
Tobacco(8) Dust, WP, & EC	MD	0.8 to 16.0	0.7, 0.7, 1.7, 2.1, 2.4, 2.8, 3.2, 10.9

Based on this foliar dissipation data and resulting residue calculations a table has been developed with food tolerance scenarios as guidance in predicting maximum expected surface residue ranges for terrestrial vegetation and insects.

**Table 5b Malathion Crop Use Terrestrial EEC Range Estimates in ppm-Grasses and Seeds  
Cheminova and IR4 Supported Maximum Tolerance Rates**

		Foliar Dissipation T1/2=5.5 Days			Number of Applications								
	Rate lb ai/A	Int. Day	1 grass- seed	2	3	4	5	6	7	8	9	10	12-25
A	0.175	7D	42-1.2									72-4.5	
B	0.50	NA	120-3.5										
C	0.61	5D	146-4.3				300-19						
C	0.61	7D	146-4.3	189-9	232-14								
	Rate lb ai/A	Int. Day	1 grass- seed	2	3	4	5	6	7	8	9	10	12-25
C	0.61	14D	146-4.3	171-11									
D	0.76	10D	182-5.3				254-16						
E	0.94	3D	226-7										
E	0.94	6D	226-7					420-26					

E	0.94	7D	226-7		357-22				384-24				
F	1.0	7D	240-7					407-25					
G	1.25	3D	300-8.8	505-32				854-53					
G	1.25	5D	300-8.8				614-38						
G	1.25	7D	300-8.8		475-30	490-31	505-32	509-32	510-32	511-32	511-32	512-32	512-32
G	1.25	14D	300-8.8	351-22									
H	1.50	7D	360-10		571-36			611-38					
I	1.56	7D	374-11	425-16			631-39						
J	1.88	5D	451-13					943-59					
J	1.88	7D	451-13		715-45	747-47		766-48					
J	1.88	14D	451-13	528-33									
K	2.03	6D	487-14					908-57					
K	2.03	7D	487-14		772-48	806-50							
L	2.5	3D	600-18		1292-56								1905-119
L	2.5	5D	600-18		1121-68								
L	2.5	7D	600-18		951-59		1011-63						
M	3.43	5D	823-24										
N	3.75	7D	900-26			1490-61		1527-96					
N	3.75	14D	900-26										
O	4.7	30D	1128-33	1153-40									
P	5.0	7D	1200-35	1696-52	1902-69	1987-87							

Days Post Application	Measured Residues Control Plots in PPM	Measured Residues Treated Plots in PPM	Precipitation in inches
0		28.8	0
1		18.9-21.1	0
2		23.8	0
3		27.6	0
4	<0.1	28.2	0.02
5		8.2	0.26
6		2.3	0.01
7		4.5	0
14	<0.1	0.4	0
21	<0.1	<0.1	0

Awad et al (1967) conducted a study entitled The Effect of Environmental & Biological Factors on Persistence of Malathion Applied as Ultra Low Volume or Emulsifiable Concentrate to Cotton Plants. In the study small amounts of insecticide were applied to plant surfaces and glass plates. Samples were taken at application and 1, 3, 6 & 9 days post application. Malathion EC and ULV formulations were used. Residues were obtained by washing leaves or plates with 100 ml of distilled water. EC formulations appeared to penetrate leaf tissues more rapidly than ULV formulations. Lack of absorption led to higher residue levels on glass plate surfaces. Calculated foliar half lives were ULV  $T_{1/2} = 5.5$  days and EC  $T_{1/2} = 23$  days.

#### D. Water Resource Assessment

The highest level of malathion surface water contamination occurs in urban areas. Malathion deposited on anthropogenic surfaces decays slowly and is more likely to be washed off to adjacent water bodies. Agricultural uses of malathion are most likely to contaminate surface water through off-target drift. This is particularly true with aerially-applied, drift-prone ULV formulations.

Malathion contamination of ground water has been observed at higher levels than predicted by modeling. Malathion ground water contamination is surprising given its rapid degradation under most conditions and emphasizes the need for data requested on malathion fate under conditions which do not favor degradation.

EFED uses a tiered system to generate estimates of pesticides in surface and groundwater. First tier assessments utilize the simple farm pond model GENEEC and reviewed monitoring studies. Second tier assessments are intended to reduce the uncertainties of first tier assessments and produce more realistic estimates of pesticide concentrations. Second tier assessments utilize PRZM-EXAMS modeling and a more comprehensive review of available monitoring data.

Water assessments used in this document examine both malathion and when possible its toxic degradate malaoxon. Technical malathion contains, in addition to malaoxon, other impurities that are demonstrated to be toxic or to synergize malathion toxicity. These chemicals include isomalathion and alkyl phosphorothioates and -dithioates. These impurities are normally present individually at levels less than 1% but can increase under improper storage conditions. As EFED possesses very little environmental fate data on these impurities it is not possible to assess their fate or persistence in the environment through modeling. Therefore an assessment of malathion impurities in water is not included in this document.

### 1. Drinking Water

A first tier drinking water assessment has been performed to provide the Health Effects Division with a conservative estimate of malathion and malaoxon in drinking water. Acute and chronic drinking water concentrations were estimated with proposed pesticide use patterns that produced the highest aqueous pesticide levels. The model results reflect first tier drinking water concentration estimates and environmental concentrations as a result of agricultural use. HED has indicated that malathion’s degradate, malaoxon, is to be included in the tolerance expression for malathion. Thus, water concentrations are provided in this assessment for both malathion and, when possible, malaoxon. The results are summarized in the table below. A more detailed discussion is provided in the discussion on ground water and the memo sent to HED is included in the appendices.

**Table 7. Tier I drinking water concentrations for malathion and malaoxon.**

compound / exposure type	surface water		ground water	
	estimated concentration (ppb)	source of concentration	estimated concentration (ppb)	source of concentration
malathion / acute	226	GENEEC peak	3.1	Monitoring data
malathion / chronic	21.2	GENEEC 56-day ave.		
malaoxon / acute	96.0	GENEEC peak	3.1	Derived from malathion monitoring data
malaoxon / chronic	75.5	GENEEC 56-day ave.		

EFED recommends that 226 and 96.0 ppb (Table 7) be considered as the highly conservative first tier estimates for acute surface drinking water levels for malathion and malaoxon, respectively. For chronic surface drinking water levels, 21.2 and 75.5 ppb are recommended for malathion and malaoxon, respectively. The chronic malaoxon value exceeds the chronic malathion level because of its longer expected environmental persistence.

40



First tier groundwater concentrations were derived from monitoring data because they were higher than results from the SCIGROW model (0.142 ppb for cotton). The highest detected malathion concentration in groundwater accepted by EFED was 3.1 ppb. Malaoxon was not examined in this monitoring study, but the same value is expected to be a conservative estimate of malaoxon concentration as malaoxon production usually accounts for less than 10% of malathion degradates. EFED recommends exposure estimates of 3.1 ppb for malathion and 3.1 ppb for malaoxon in ground water.

Standard modeling techniques were modified to estimate malaoxon concentrations. Malaoxon levels were estimated with the GENEEC model with the assumption that fate variables which were not known are the same as those for malathion. Acceptable environmental fate studies specifically for malaoxon; including degradation, metabolism, mobility, dissipation, and solubility data; are needed for a complete assessment.

EFED notes that there is limited information available on the conversion of malathion to malaoxon during drinking water treatment. Available data suggest that conversion of malathion to malaoxon may be more efficient during water treatment than under natural conditions in the field, thus malaoxon may be present at a much higher concentration relative to malathion after water processing.

In a limited sampling of water entering and leaving a water treatment plant in Florida both malathion and malaoxon levels generally decreased after treatment, however, one sample showed an increase in malaoxon (USDA 1997). Data from a more detailed sampling and analysis with a lower detection limit show a much higher rate of conversion (personal communication, Dr. Marion Fuller, Florida Department of Agriculture and Consumer Services).

Data supplied by the Florida Department of Agriculture and Consumer Services Bureau of Pesticides provides malathion concentrations entering and leaving the Hillsborough Water Treatment Plant during the period of July 16 - August 27 1997. At this time the area was being sprayed with baited malathion for medfly control. The reservoirs and other known sources of drinking water reportedly did not receive direct insecticide treatment. Samples were collected at a boat dock prior to entering the plant and as well as after treatment. Table 8 summarizes the data.

**Table 8.** Drinking water entering and leaving the Hillsborough Water Treatment Plant 1997. Detection limits were 0.1 ppb for both malathion and malaoxon.

Malathion-malaoxon conversion in water treatment						
location	malathion (average of detected concentrations, ppb) <sup>1</sup>	s.d. dev.	n	malaoxon (average of detected concentrations, ppb) <sup>1</sup>	s.d. dev.	n
Boat dock	1.00	0.90	13	nd	-	25
Lab/Finished	nd	-	25	1.09	0.73	14

These results suggest that the water treatment process can result in a very efficient conversion of malathion to its oxon. It is likely that the efficiency would vary depending on the type of water treatment process used in sterilization. The stability of malaoxon in the drinking water supply cannot be assessed as EFED does not presently possess hydrolysis data on malaoxon. Therefore it is assumed to be stable in drinking water.

**2. Estimated Concentrations for Surface Water Resources**

**Tier I Assessment**

Summary

Based on fate characteristics, model predictions and actual monitoring studies, the Agency predicts malathion will reach surface and groundwater water from the proposed reregistration uses. Surface water concentrations resulting from agriculture uses were modeled using the GENEEC screening model. Results are presented in the following table.

**Table 9. GENEEC Predicted Environmental Concentrations For Aquatic Exposure**

Estimated Aquatic EECs in PPBs: Peak (top), 21 day mean (middle), and 56 day mean (bottom).

	Rate lb ai/A	Int. Day	Number of Applications											
			1	2	3	4	5	6	7	8	9	10	12-25	
<b>A</b>	0.175	7D											8.24 2.03 0.78	
<b>B</b>	0.50	NA	11.4 2.8 1.07											
<b>C</b>	0.61	7D		23.2 5.7 2.2	27.7 6.8 2.6									

C	0.61	14D		26.8 6.6 2.53									
	<b>Rate</b> lb ai/A	<b>Int.</b> <b>Day</b>	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>	<b>8</b>	<b>9</b>	<b>10</b>	<b>12-25</b>
D	0.76	10D					40.6 10 3.4						
E	0.94	3D	21.7 10.4 2.05										
E	0.94	6D						45.4 11.2 4.24					
E	0.94	7D			42.2 10.4 3.94				42.5 10.5 3.97				
F	1.0	7D						45.2 11.2 4.22					
G	1.25	3D	28.5 7.0 2.66	54.3 13.4 5.07					90.4 22.3 8.5				
G	1.25	5D					66 16.3 6.16						
G	1.25	7D			56.1 13.8 5.24	56.5 13.9 5.3	56.5 13.9 5.28	57.2 14.1 5.4	56.5 13.9 5.28	56.6 13.9 5.28	56.6 13.9 5.28	56.6 13.9 5.28	56.6 13.9 5.28
G	1.25	14D		47.1 11.6 4.53									
H	1.50	7D			67.3 16.6 6.28				67.3 16.6 6.28				
I	1.56	7D		67.8 16.7 6.33			70.8 17.5 6.61						
J	1.88	5D						99.4 24.6 9.28					
J	1.88	7D			84.4 20.8 7.87	84.9 21 7.92		85 21 7.94					
J	1.88	14D		70.8 17.5 6.61									
K	2.03	6D						98.1 24.2 9.16					

<b>K</b>	2.03	7D			91.1 22.5 8.5	91.7 22.6 8.6							
<b>L</b>	2.5	3D											181
<b>L</b>	2.5	5D			128 31.6 12								
<b>L</b>	2.5	7D			112 27.7 10.5		113 27.9 10.6						
<b>M</b>	3.43	5D					81 20 16.9						
<b>N</b>	3.75	7D				169 41.7 15.8		169 41.7 15.8					
<b>N</b>	3.75	14D				142 35 13.2							
<b>O</b>	4.7	30D		171 42.2 15.9									
<b>P</b>	5.0	7D			224 55.3 20.9	225 55.6 21.1							
<b>Q</b>	6.25	Q1			226 55.8 21.2								

**Table 9 Crop Scenario Relationships**

- 0.175 lb ai/A A10=Orange, Grapefruit, Lemon, Lime, Tangerine, Tangelo, and Kumquat  
0.50 lb ai/A B1=Flax  
0.61 lb ai/A C5(5D)=Sweet Corn , C2(7D)=Hops, C3(7D)=Beans, Corn, Rice, Sorghum, Wheat, and Rye  
C2(14D)=Alfalfa, Clover, Lespedeza, Lupine and Vetch  
0.76 lb ai/A D5=Blueberry  
0.94 lb ai/A E1(3D)=Grass for hay, E4(3D)=Mushroom, E6(6D)=Strawberry, E3(7D)=Pepper and  
spearmint, E7(7D)=Macadamia  
1.0 lb ai/A F6(7D)=Melons, Watermelon, Pumpkin and Squash  
1.25 lb ai/A G1(3D)=Grass for hay, G2(3D)=Field corn , G6(3D)=Mustards, G25(3D)=Cotton,  
G5(5D)=Watercress, G3(7D)=Rice, Sorghum, Wheat, Rye, Barley, Oats and Corn,  
G4(7D)=Blueberry( ULV), G5(7D)=Turnip, Broccoli, Apple, Sweet Corn, Beet, Chayote,  
Horseradish, Parsnip, Radish, Rutabaga, Salsify, Sweet potato, G6(7D)= Cabbage and  
Cherry(ULV), G7(7D)=Carrot , G8(7D)=Mango and Passion fruit , G9(7D)=Asparagus  
G10(7D)=Pears and Quince , G12(7D)=Guava and Papaya, G2(14D)=Alfalfa, Clover, Lupine,  
Vetch nd Lespedeza  
1.5 lbs ai/A H2(7D)=Celery, H6(7D)=Okra  
1.56lbs ai/A I2(7D)=Potato, I5(7D)=Onion, Garlic, Shallot, Leeks  
1.88 lb ai/A J6(5D)=Lettuce, J4(7D)=Blackberry, Raspberry, Loganberry, Boysenberry, Dewberry, Currant,  
Gooseberry, J3(7D)=Cucumber, J6(7D)= Strawberry, J2(14D)=Grapes  
2.03 lbs ai/A K6(6D)=Strawberry(50%WP), K3(7D)= Spinach, Dandelion, Endive, Parsley and Swiss Chard,

2.50 lb ai/A	K4(7D)=Blackberry, Raspberry, Gooseberry, Loganberry, Dewberry, Currant and Boysenberry
3.43 lb ai/A	L25(3D)=Cotton, L3(5D)=Figs, L3(7D)=Mustards, Walnuts, and Pecans, L5(7D)=Peas
3.75 lb ai/A	M5(5D)=Tomato, Pepper, Eggplant
4.7 lb ai/A	N4(7D)=Apricots, N6(7D)=Cherry, N4(14D)=Peach and Nectarine
5.0 lb ai/A	O2(30D)=Avocado
6.25 lb ai/A	P3(7D)=Pineapple, P4(7D)=Chestnuts
	Q3(30D)=Oranges, Grapefruit, Lemon, Lime, Tangerine and Tangelo

## Tier II Assessment

### Summary

Since the EEC's derived from first-tier GENEEC simulations were above levels of concern (LOCs) for aquatic organisms, Tier II EEC's were calculated. The second tier assessment for malathion in surface waters utilized PRZM-EXAMS modeling and the review of available monitoring data. It is not possible to perform second tier modeling of malaoxon due to the lack of fate data on this toxic degradate. Required data for malaoxon include degradation (161-1), metabolism (162-1,4), and mobility (163-1,2) and well as data on solubility and rates of formation under suitable conditions.

PRZM 3.1 was used to simulate the agricultural field, and EXAMS 2.97.5 was used to simulate fate and transport in surface water. Each model scenario simulates a single site which represents the use of malathion on a particular crop. The weather and agricultural practices were simulated over multiple years, in this case 24 to 36, so the probability of an EEC occurring at that site can be estimated.

Five application scenarios were simulated, using crops which together represent more than 50% of malathion use in the United States as well as the highest use-rates proposed by the registrant. The EECs derived from these simulations were generally lower than those generated by Tier I GENEEC runs (see Table 10). Twenty-two additional crops have malathion application rates, numbers of applications and application intervals identical to one of the five crops simulated (see Table 1). The EECs generated from the five scenarios may be used as surrogates for these twenty-two crops, recognizing that these predicted EECs may vary due to geographical and other differences.

Monitoring and field studies were also examined in this second tier assessment. Studies reviewed include those associated with Mediterranean fruit fly and boll weevil eradication programs. Data from these studies (summarized in tables 10-13) shows in agricultural settings that the most important source of aquatic malathion is off-target drift and malaoxon is only detected at low concentrations. In urban areas, runoff is more important and malaoxon levels can be much higher.

### PRZM-EXAMS Modeling

PRZM-EXAMS estimates aquatic concentrations in a one hectare pond that is two meters deep next to a ten hectare plot. The pond receives both simulated drift and runoff from the field. PRZM models terrestrial pesticide fate and transport and EXAMS models the aquatic portion.

The Pesticide Root Zone Model (PRZM) is a one-dimensional, dynamic, compartmental model that can be used to simulate chemical movement in unsaturated soil systems within and immediately below the plant root zone. It has two major components-- hydrology (and hydraulics) and chemical transport. The hydrologic component for calculating runoff and erosion is based on the Soil Conservation Service curve number technique and the Universal Soil Loss Equation. Evapotranspiration is estimated either directly from pan evaporation data, or based on an empirical formula. Evapotranspiration is divided among evaporation from crop interception, evaporation from soil, and transpiration by the crop. Water movement is simulated by the use of generalized soil parameters, including field capacity, wilting point, and saturation water content.

Exposure Analysis Modeling System (EXAMS II) is a model that can receive progressive PRZM runoff concentration output to further predict aquatic pesticide concentrations in a simulated pond. The predicted concentrations over the period of time (usually 24-36 years) can be averaged over time to produce peak and time averaged concentrations which take into account aquatic degradation.

EFED has prepared standard PRZM input files for the five crops simulated. The locations used to build these scenarios were chosen to represent areas of greatest malathion use. EFED has prepared draft summary documents which describe the input parameters used to develop the standard scenarios. Once these documents have been finalized, they can be provided upon request.

The five input files were adapted to simulate the application of malathion for the respective crops and states represented in the standard scenarios. Chemical-specific input for malathion was derived to the greatest extent possible from the environmental fate database submitted to the EPA by registrant Cheminova. Application rates, numbers of applications, and application intervals simulated were consistent with the maximum values requested by the registrants for establishing tolerances. Average application rates and numbers of applications were taken from BEAD Quantitative Use Analysis reports. Planting and harvest dates, and likely dates of malathion application, were chosen based on conversations with academic and extension crop specialists, or USDA references. Further details on crop and chemical specific inputs are presented in the appendices.

### **Results**

The PRZM-EXAMS Tier II EECs for malathion are listed along with GENEEC Tier I EECs in the following table for comparison purposes.

**Table 10.** Summary of model results inputting maximal and typical use rates for crops with high malathion usage and high use rates. Intervals and the total number of applications used for these modeling runs were chosen to be conservative. By using maximum use rates (used in food residue tolerance limits), minimal intervals between applications, and the maximum number of applications the modeled use scenarios are expected to reflect a high exposure scenario.

PRZM-EXAMS results								
Crop	% of total a.i. applied / year <sup>1</sup>	Use rate (lbs a.i./A)	Interval (days)	No. of applications	Model Comparisons			
					GENEEC		PRZM-EXAMS	
					peak	56 d ave.	peak <sup>2</sup>	60 d ave. <sup>3</sup>
Cotton	41.6%	Max: 2.5	3	25	181	16.9	291	47.7
		Typ: 0.3	[3] <sup>4</sup>	4	20.6	1.95	7.9	0.50
Sorghum	7.4%	Max: 1.25	7	3	56.1	5.24	26.7	1.95
		Typ: 0.8	[7]	1	18.5	1.74	2.94	0.18
Apple	2.14%	Max: 1.25	7	5	56.5	5.28	0.80	0.19
		Typ: 0.7	[7]	3	31.4	2.93	0.59	0.09
Citrus	0.49%	Max: 6.25	30	3	226	21.2	162	11.1
		Typ <sup>5</sup> : 2.5	[30]	1	57.1	5.33	47.3	2.59
Lettuce	0.45%	Max: 1.88	5	6	99.4	9.28	15.4	2.98
		Typ: 2.0	[5]	1	45.7	4.26	5.63	0.56

<sup>1</sup>Pesticide data compiled by the National center for Food and Agricultural Policy for 1991-1993 and 1995.

<sup>2</sup>Peak concentration expected once in ten years.

<sup>3</sup>The 60 day average value expected once in ten years.

<sup>4</sup>Since data on typical intervals is not available the minimum interval was used in model runs with average use rates and number of applications.

<sup>5</sup>Typical values for oranges.

### Limitations of Modeling Analysis

There are several factors which limit the accuracy and precision of this analysis including the selection of the high exposure scenarios, the quality of the input data, the ability of the models to represent the real world, and the number of years that were modeled.

Scenarios that are selected for use in Tier 2 EEC calculations are ones that are likely to produce large concentrations in the aquatic environment. It should represent a site that really exists and

would be likely to have the pesticide in question applied to it. It should be extreme enough to provide conservative estimates of the EEC, but not so extreme that the model cannot properly simulate the fate and transport processes at the site. Currently, sites are chosen by best professional judgement to represent sites which generally produce EEC's larger than 90% of all site use for that crop. The EEC's in this analysis are accurate only to the extent that the site represents this hypothetical high exposure site. The most limiting part of the site selection is the use of the standard pond with no outlet. Obviously, a Mississippi pond, even with appropriately modified temperature data is not the most appropriate water body for use in all situations. Some water bodies would likely have higher concentrations. These would be shallow water bodies near agricultural fields that receive most of their water as runoff from agricultural fields.

The quality of the analysis is directly related to the quality of the input parameters. Most of the fate data for malathion is complete, however, there is little fate and transport data on toxic impurities and degradates such as malaoxon. In addition, the aquatic persistence inputs were derived from studies conducted under alkaline conditions which would be likely to increase degradation rates and reduce the conservatism of this analysis.

The models themselves represent a limitation on the analysis quality. While the models are some of the best environmental fate estimation tools available, they have significant limitations in their ability to represent some processes. Spray drift is estimated as a straight 5% of the application rate reaching the pond for each application from ground application. In actuality, this value should vary with each application from near zero to higher than 20%. A second major limitation of the models is the lack of validation at the field level for pesticide runoff. While several of the algorithms (volume of runoff water, eroded sediment mass) are well validated and well understood, no adequate validation has yet been made of PRZM 3.1 for the amount of pesticide transported in runoff events. Other limitations of the models used are the inability to handle within site variation (spatial variability), lack of crop growth algorithms, and an overly simple soil water transport algorithm (the "tipping bucket" method). Another limitation is that only thirty-six years of weather data was available for the analysis at most sites and less at others. Consequently, at best there is approximately 1 chance in 20 that the true 10% exceedence EEC's are larger than the maximum EEC in the calculated in the analysis. If the number of years of weather data could be increased it would increase the confidence that the estimated value for the 10% exceedence EEC was close to the true value.

There are certain limitations imposed when Tier II EEC's are used for drinking water exposure estimates. Obviously, a single 10 hectare field with a 1 hectare pond does not accurately reflect the dynamics in a watershed large enough to support a drinking water facility. A basin of this size would certainly not be planted completely to a single crop nor be completely treated with a particular pesticide. Additionally, treatment with the pesticide would likely occur over several days or weeks, rather than all on a single day. This would reduce the magnitude of the concentration peaks, but also make them broader, reducing the acute exposure but perhaps increasing the chronic exposure. The fact that the simulated pond has no outlet is also a limitation as water bodies in this size range would have at least some flow through (rivers) or turnover (reservoirs). Also, irrigation of crops in the desert scenarios was not considered in the



models. EEC's would likely be higher if EFED had irrigation data available. In spite of these limitations, a Tier II EEC can provide a reasonable upper bound estimate of the concentration found in drinking water. Risk assessment using Tier II values can be used as refined screens to demonstrate that the risk is below the level of concern.

### **Monitoring Data**

Monitoring data was collected through two USDA programs: the Boll Weevil Eradication Program and the Mediterranean fruitfly (medfly) control effort.

#### Agricultural

Agricultural runoff monitoring data and field studies are consistent with aerobic soil metabolism studies showing that malathion is normally rapidly degraded on soil to compounds of lower toxicity.

Malathion is water soluble and thus poses the potential to be dissolved in rain water and transported in runoff water from application sites if it not degraded. Levels of malathion in runoff water have been examined mostly using automatic runoff sampling equipment (ARSEs) which consist of collection bottles with funnels recessed in the ground at sites where runoff is expected. The amount of malathion in runoff is expected to be affected by numerous variables including the soil type, half-life on the particular soil, the amount of time between application and precipitation, the amount of precipitation, and vegetation. Table 11 shows runoff monitoring data from five treated cotton fields in the Boll Weevil program close to bodies of water. Sampling was performed close to the field (10-25 feet) and closer to the water (40-135 feet from the field). In most cases, malathion concentrations were lower when the interval between application and rainfall was longer and/or distance from the field was farther. These observations are expected since increasing the interval allows for more degradation to occur and farther runoff travel distances allow malathion to penetrate soil and adsorb to soil particles before reaching shorelines.

**Table 11. Field monitored runoff Cotton Bollweevil Control Program**

Malathion levels were measured in runoff water from cotton fields after rain events. Two sets of measurements were made, one closer to the field and one farther from the field. Adapted from Environmental Monitoring Report: 1997 Southeast Boll Weevil Eradication Program Sensitive Sites and Environmental Monitoring Report: 1996 Southeast Boll Weevil Eradication Program.

Field Runoff			
field no. / sample distance from field	malathion conc (ppb)		time from application to rain (days)
	closer to field	farther from field	
1806-502 / near to field: 20' farther from field: 110'	9.3	1.9	1
	7.5	3.5	3
	>0.3	>0.3	6
1806-504 / near to field: 20' farther from field: 40'	70	33	1
	0.48	nd	6
2025-187 / near to field: 10' farther from field: 70'	0.42	0.53	2
2027-468 / near to field: 15' farther from field: 135'	63	nd	1
	nd	-	5
2100-200 / near to field: 25' farther from field: 50'	4.2	3.8	18
502 / near to field: 20' farther from field: 110'	1.1	nd	3
	0.5	nd	7
504 / near to field: 20' farther from field: 40'	10.9	nd	1
	41.8	15.6	3
	146	93.5	7
7806 / near to field: 0' farther from field: 45'	0.9	0.5	?
	1.7	1.1	6
	<0.3	0.3	14
325 / near to field: 15' farther from field: 60'	8.54	.82	2
	35.8	16.2	9

nd = none detected.

In monitoring projects the stability of malathion in still water has been examined. A half-acre pond surrounded by cotton fields with a 25 foot buffer was monitored for malathion (USDA

BWEP 1993). Pesticide drift was determined to be the most important mechanism of contamination of the pond. Residues levels in the pond were lower before treatment (<0.1-0.44 ppb) and higher immediately after malathion application (<0.33-91.4 ppb). In most cases malathion in the pond degraded to <0.33 ppb within 7 days. Runoff was only a minor contributor of residue to the pond but only two rainfalls occurred during the sampling period. The malathion in the runoff samples collected were 9.75 and 76.3 ppb one day after the first and last treatments, respectively. Other natural bodies of water within treatment areas, but not intentionally receiving direct spray, showed no detectable levels of malathion 3-27 days after applications ceased (USDA BWEP 1995).

**Tables 12 a,b,c.** Spray drift to adjacent moving water. Malathion levels in moving water adjacent to cotton fields was measured before and after treatment. Measurements were made downstream from the field every 15 min from one hour before until 2-3.25 hours after application. Application was made when wind was not blowing directly over the water.

Spray Drift							
Southeast Boll Weevil Eradication Program							
site/comments	applicatio n (aerial / ground)	treatment #	days since last treatment	time (min) / downstream malathion (ppb)			
				before treatment	time	after treatment	time
McCall's Creek:  The creek was separated from the field (13.3 acre) by a continuous 600-700' buffer of 30-60' trees.	a	1	?	nd		nd	
	a	2	8	nd		nd	
	a	3	6	nd		nd	
	a	4	7	nd		nd	
	a	5	<del>7</del>		16.1	-60	nd
North River:  The field (8.3 acre) is separated from the river by a continuous buffer of mature hardwoods and moderately dense understory approximately 125' deep.	g	1	?	-		nd	
	g	2	5	nd		nd	
	g	3	7	nd		nd	
	g	4	6	<0.33	-45	<0.33	45
	g	5	6	<0.33	0	<0.33	0-120
	a	6	10	1.54	-45	1.44	60
	a	7	6	<0.33	0	<0.33	0-120
	a	8	7	1.77	-60	1.46	0
	a	9	10	0.42	-45	0.55	45
Pursley Creek:  The field (95.3 acre) was separated from the creek by 100' of mature hardwoods with a dense understory.	a	1	?	nd		3.54	135
	a	2	7	nd		0.39	120
	a	3	7	nd		1.03	30
	a	4	7	nd		<0.33	75-120

	a	5	7	6.63	-30	3.80	120
	a	6	6	nd		3.35	150
Stewart Creek:  The field (19.2 acre) was separated from the creek by a 25' buffer of low-lying kudzu vegetation.	g	1	?	nd		nd	
	g	2	8	<0.33	-60	nd	
	a	3	7	nd		7.69	60
	a	4	5	nd		3.16	75'
	g	5	7	0.52		<0.33	0-240
	g	6	4	0.51		10.89	15
	g	7	5	<0.33		<0.33	15, 105, 135-250
	a	8	6	1.01		4.52	60
	a	9	12	<0.33		3.49	105

b.)

<b>Spray Drift</b> Environmental Monitoring Report Boll Weevil Cooperative Eradication Program: 1995 Texas Lower Rio Grande Valley						
site/comments	aerial/ ground	treatment #	downstream malathion (ppb) / time (min)			
			before treatment	time	after treatment	time
#204060311/ Canal 200' from treated field.	?	1	0.324	-15	0.297	15
	?	2	4.89	-15	7.26	30
#2144070704 Canal 40' from treated field	?	1	6.38	-30	11.4	0
	?	2	2.27	-45	1.87	0
#212080704/ Canal 150' from treated field	?	1	4.81	-45	4.15	30,120
	?	2	2.4	-30	4.37	120
	?	3	5.92	-45	4.21	0

c.)

Spray Drift						
Environmental Monitoring Report						
Southern Rolling Plains Boll Weevil Eradication Program - 1994-1995						
			peak downstream malathion (ppb) / time (min)			
site/comments	method of app.	treatment #	before treatment	time	after treatment	time
Concho County stream (10303-1408) Samples collected 0.25 miles downstream	Hi-Boy	1	0.849	-15	6.95	105
	Mist blower	2	0.695	-45	86.9	225
	Mist blower	3	0.273	-45	0.503	210
Concho River (10708-2707) Samples collected 0.25 miles downstream	Mist blower	1	0.676	-15	0.813	0
	Mist blower	2	0.871	-60	0.589	150
	Mist blower	3	2.24	-60	7.45	15

Wide buffer strips (125-700 feet) with high vegetation appeared to reduce malathion drift to sensitive areas to levels below detection while narrower and lower buffer afforded less protection (Table 12). With aerial applications, 8 of 19 applications lead to higher aquatic malathion concentrations, whereas only 1 of 10 ground applications resulted in higher malathion levels. Thus aerial applications appear more prone to drift than ground applications. Although increased malathion levels in the streams, rivers, and canals examined after nearby treatments decreased rapidly, decreases are likely due primarily to the movement of contaminated water downstream. To assess malathion stability in moving water a sampling station further downstream would be required along with measurements of the flow rate of the water.

### Residential

Monitoring data suggests that urban malathion use poses the highest risk of contaminating surface water. However, use data is not available to correlate with monitoring data to determine which particular uses have the greatest impact. Total usage and use rates in specific cities is also unavailable. Targeted urban monitoring and preliminary fate experiments suggest however that malathion contacting anthropogenic surfaces is likely to convert to the oxon and has a high runoff potential.

Malathion concentrations in water in and around urban medfly treatment areas in California and Florida have been measured. Although risk assessment of malathion use for medfly control is not included in this document (these generally fall under section 18 local need uses) the monitoring studies associated with this use provide information on malathion fate and transport in residential settings. In urban areas not involved in medfly control measures malathion can be found in runoff water at higher levels than agricultural areas. A monitoring report by United States Geological Survey showed that higher residues are found in urban areas. In this analysis of 11 urban streams (604 samples) and 37 agricultural streams (1530 samples) malathion concentrations were higher in the urban tributaries.

It is likely that proposed residential uses will result in aquatic contamination. Residential malathion uses include outdoor home and garden, public park, and commercial use as well as residential mosquito control. Home use formulations may be applied as a "... spray to lower foundation of house, patios and garbage cans ... along fences; to firewood piles; and other infested areas." (Ortho Malathion 50 Plus Insect Spray label). Malathion on the surfaces described on the this label is likely to persist longer and be more available for runoff than malathion on soil. Fyfanon ULV formulation is applied at 0.2-0.23 lbs/A aerially at 150 mph over residential areas for mosquito control. In addition to covering anthropogenic surfaces it is likely that moderate sized bodies of water receive direct spray during normal aerial mosquito control use. In medfly treatments, malathion is mixed with a bait mixture and applied aerially at nearly the same rate as in mosquito control but with large buffers (up to 200 feet). Medfly applications in residential areas provide useful information on the fate and transport of malathion in these settings, but it is very likely that the smaller particles produced from the ULV formulation used in mosquito control results in more drift than the baited mixture for medfly. Thus, medfly monitoring data of drift will be expected to underestimate drift from ULV mosquito use.

In medfly control efforts larger bodies of water are "flagged" to avoid direct malathion treatment. Thus, contaminated water bodies presumably received insecticide residues by drift and runoff. On average reservoirs in the treatment area which were flagged to avoid direct spray contained 0.16 ppb before treatments and 2.59 ppb immediately after treatment (Table 12). All waters in and around the treatment area, whether protected or not, showed increased malathion levels immediately after treatment. In general, applications were performed approximately weekly with no noted aggregate accumulation of malathion in water.

Rainwater runoff in California medfly treatment area contributed greatly to malathion levels in a stream passing through the treatment area. After precipitation, inflow into the treatment area contained less than 1 ppb while downstream water contained up to 203 ppb malathion. Maxima in 1990 and 1981 were 44.1 and 583 ppb (Environmental Monitoring Results of the Mediterranean Fruit Fly Eradication Program, Riverside County 1994).

**Table 13 a Medfly spraying and malathion levels in bodies of water.** Malathion was measured immediately before and after spraying a bait formulation at ~0.17 lbs ai/A from an altitude of 300 feet. This data was adapted from A Characterization of Sequential Aerial Malathion Applications in the Santa Clara Valley of California, 1981.

Aquatic malathion concentrations resulting from medfly applications												
site	treatment #	days since last spray	malathion					malaoxon (ppb)				
			no. of sample	before spray (ppb)	S.d. Error (ppb)	after spray (ppb)	S.d. Error (ppb)	no. of samp	before spray	S.d. Error (ppb)	after spray	S.d. Error (ppb)
Unprotected <sup>1</sup> natural waters	1	*	14	*	*	4.94	2.71	*	*	*	*	*
	2	9	6-16	0.20	0.05	18.66	5.81	1	*	*	18.0	*
	3	11	13-15	1.50	1.17	9.78	2.47	*	*	*	*	*
	4	7	14-15	0.48	0.13	95.4	53.2	1-2	0.64	*	1.9	0.20
	5	7	13-14	0.66	0.12	4.97	1.05	4-5	0.19	0.046	0.63	0.17
	6	7	11-12	0.57	0.20	23.4	11.6	1-4	0.90	*	0.35	0.10
Average	-	8.2	-	0.68	0.33	26.19	12.8 1					
Protected <sup>2</sup> natural waters	1	*	20	0.091	0.058	0.33	0.07 8	*	*	*	*	*
	2	9	20	0.12	0.07	0.56	0.10	*	*	*	*	*
	3	11	19-20	0.056	0.028	0.90	0.15	*	*	*	*	*
	4	7	14-15	0.12	0.07	1.25	0.22	*	*	*	*	*
	5	7	20-22	0.040	0.019	2.10	0.41	1	*	*	0.40	*
	6	7	15-19	0.053	0.040	0.39	0.08 9	2	*	*	0.45	0.25
Average	-	8.2	-	0.080	0.048	0.92	0.17					
Flagged reservoirs	2	9	2	0.18	0.03	0.75	0.65	1	*	*	2.7	*
	3	11	2	*	*	0.50	0.10	*	*	*	*	*
	4	7	19-20	0.033	0.024	8.39	3.81	2	*	*	0.92	0.29
	5	7	10-12	0.51	0.30	1.90	0.94	*	*	*	*	*
	6	7	8	0.075	0.062	1.42	0.41	1	0.1	*	0.83	*
Average	-	8.2	-	0.16	0.083	2.59	1.18					
Reservoirs outside treatment area	2	9	2	0.05	0.05	0.34	0.07	*	*	*	*	*
	3	11	2-4	0.10	0.10	1.0	0.55	*	*	*	*	*
	4	7	10	0.03	0.03	0.30	0.16	*	*	*	*	*

	5	7	10	0.036	0.024	0.14	0.058	1	1.3	*	*	*
	6	7	8-10	0.18	0.074	0.21	0.087	*	*	*	*	*
Average	-	8.2	-	0.079	0.056	0.40	0.19					

<sup>1</sup>Unflagged and within the treatment area.

<sup>2</sup>Flagged to avoid treatment or outside the treatment area.

\* No data.

**Table 13 b.** Malathion level in 29 ponds in Florida exposed to a.) direct or b.) indirect malathion spray in medfly control. Samples were collected within eighteen hours of approximately weekly treatments of 0.15 lbs/A. Unprotected bodies of water were ~0.1 miles in length and may have received runoff from surrounding watersheds. Protected waters were rivers or larger lakes. Statistically, values below the detection limit (0.1 ppb) were treated as 0 ppb and values below limit of quantitation (0.3 ppb) were treated as 0.15 ppb. The data was adapted from the Environmental Monitoring Report: Cooperative Medfly Project Florida, 1997.

a.)

Unprotected aquatic sites						
site	before			after		
	no. of samples	ave (ppb)	stdev (ppb)	no. of samples	ave (ppb)	stdev (ppb)
Fairgrounds	8	0.06	0.07	9	1.20	1.54
Palm river	9	0.78	0.72	7	3.97	3.24
Ragen Park	6	14.12	14.17	7	35.75	27.50
University Square Mall	7	0.04	0.07	7	3.77	3.67
Pond Lake	6	4.11	4.35	10	9.25	11.78
Bloomington Area	9	0.81	0.71	9	6.12	7.22
Carrollwood	7	1.05	2.01	6	4.77	3.75
Town and Country	6	1.10	1.15	5	6.88	3.07
McDill Site	5	0.12	0.06	4	5.20	2.33
Brandon Town Center	5	3.50	1.86	8	65.71	149.18
Lowry Zoo	7	0.14	0.22	6	1.55	1.86
Sun 'n Fun	8	0.09	0.07	10	7.28	15.48
Hamilton Creek	6	0.61	0.41	7	10.74	19.51
Eagle Lake	7	1.60	2.29	7	13.99	10.39



b.)

Protected aquatic sites						
site	before			after		
	n	ave (ppb)	s.d. (ppb)	n	ave (ppb)	s.d. (ppb)
Moore's lake	10	0.36	0.78	10	0.76	1.66
Lake Weeks	12	0.69	0.67	11	4.85	4.08
Lake Valrico	12	0.03	0.06	11	2.84	6.71
Lake Kathy	12	0.43	0.91	11	5.91	9.15
Lake Walden	6	0.21	0.14	6	2.21	2.37
Alafia River	6	0.13	0.17	6	1.93	4.06
Hillsborough River	8	0.35	0.39	8	5.02	9.13
Platt Lake	2	0.08	0.08	2	0.85	0.15
Lake Magdalene	2	0.08	0.08	2	0.80	0.20
Lake Carroll	2	0.31	0.16	2	1.65	0.55
Crystal Lake	9	0.02	0.05	9	0.46	0.74
Lake Horney	10	0.03	0.06	9	3.47	3.86
Banana Lake	7	0.21	0.33	7	2.48	3.97
Crews Lake	7	0.23	0.19	7	0.82	0.96

Residential settings are expected to be composed of numerous surfaces which may be physically and biologically impervious to malathion. The relative quantities and effects of adsorption and degradation on concrete, roofing, metal, and plastics is unknown in the residential settings where malathion may be sprayed for medfly and mosquito control. Monitoring results suggest that the residential surfaces increase availability of malathion for runoff probably due to lack of microbial activity which decreases metabolism, less water content which decreases hydrolysis, and little adsorption. Although the application rate for mosquito control is low relative to agricultural use (0.20-0.23 lbs/A for aerial mosquito control versus 0.18-6.25 lbs/A for agricultural pest control), application over wide areas may be concentrated in storm drain systems along with malathion from home garden and commercial site use.

The concentration factor appears to be greater in residential settings when comparing residential and agricultural runoff. This is consistent with the results of several USGS and USDA monitoring studies. Preliminary monitoring results for malathion in surface water (USGS 1997) show malathion was detected above 0.01 ppb with a 2.61% frequency in agricultural streams while in urban streams the frequency was 20.86%. The USDA monitoring studies for boll weevil control show an average runoff concentration of 15.5 ppb (Table 11) while average downstream creek concentrations in the urban Santa Clara Valley of central California were 177 ppb during 1981 malathion spraying for medfly.

The highest levels of aquatic malaoxon found in a search of available data was a result of medfly control efforts in California (CDFG 1982). The following table is derived from the monitoring study during the malathion spraying in the Santa Clara Valley. Samples were taken 2-3.5 hours

after the first rainfall 6 days after the last application. These runoff concentrations are much higher than agricultural runoff levels.

**Table 14.** Malathion and malaoxon concentrations in creeks after malathion applications in the Santa Clara Valley.

Sampling Location		malathion (ppb)		malaoxon (ppb)	
		average	sd	average	sd
Adobe Creek	50' upstream	449	17.7	164	33.2
	drain	583	40.3	328	18.4
	100' downstream	361	20.5	169	-
Stevens Creek	50' upstream	159	-	68.0	-
	drain	434	73.5	147	4.2
	150' downstream	156	23.3	68.0	-
Guadalupe Creek, site 1	50' upstream	1.9	0.2	0.8	0.3
	drain	142	-	147	4.2
	150' downstream	23.5	2.1	22.0	-
Guadalupe Creek, site 2	50' upstream	137	25.4	212	9.2
	drain	188	12.0	250	8.5
	150' downstream	169	6.4	231	8.5

Fate data for malathion clearly show that its major routes of degradation are through aerobic microbial metabolism and hydrolysis. Both of these routes are expected to be lower on inert, dry surfaces; thus malathion persistence would be expected to be increased. Malathion persistence on steel plates is extended relative to soil with only 15% lost in two days (State of California 1996) compared to several soils on which 50% can be degraded in 8 hours. Slowed malathion hydrolysis and metabolism is likely to result in increased malaoxon levels via abiotic oxidation. On the steel plate study mentioned previously, malaoxon accounted for 5% of the degradates, significantly higher than the maximum of 1.8% on soil reported by the registrant.

### Limitations of Monitoring Analysis

Although a relatively large amount of monitoring data is available for malathion, the level of detail among the studies varies. Malathion is used on more than 60 crops and settings but detailed monitoring studies were conducted mostly in conjunction with medfly and boll weevil

eradication measures. Other studies lack important data such as detection limits, surroundings, wind direction, and rainfall data making comparisons between studies difficult. Though boll weevil eradication data may be considered to reflect malathion behavior on crops grown in the same geographical areas, more data is necessary to improve quality and to allow detailed statistical analysis. Malathion applications and monitoring associated with medfly control mostly occur in residential settings and thus are probably also representative of residential mosquito control in urban areas.

### 3. Ground Water Assessment

Malathion has some mobility characteristics similar to other chemicals that have been detected in ground water. In addition, malathion has been occasionally detected in ground water at levels ranging from 0.03 to 6.17 ppb.

USGS data in the National Water Quality Assessment show a malathion detection in ground water at > 0.05 ppb. The EPA Pesticides in Ground Water Data Base 1971-1991, National Summary reports malathion detections in ground water in California (1 detection out of 499 wells sampled at a concentration of 0.32 ppb), Mississippi (2 detection out of 263 wells, concentrations of 0.03-0.053 ppb) and Virginia (22 detections made in 9 of 138 wells samples with a range of concentrations of 0.007-6.17 ppb ). All malathion detections in Virginia ground water were made in Westmoreland county as part of the Watershed/Water Quality Monitoring for Evaluating BMP Effectiveness program. Westmoreland is a coastal county and is approximately equally comprised of agricultural and forested land. Analytical detections in this study were conducted using gas chromatography with an electron capture detector and confirmed with flame photometric detector. Samuel Johnson of Westmoreland County Extension provided information on malathion use in this county. In recent years most of the agricultural malathion usage was for the control of cereal leaf beetle on barley and wheat. However, since 1995 a synthetic pyrethroid has replaced most malathion usage in Westmoreland. Mosquito abatement is not a major use of malathion in Westmoreland.

EFED believes these monitoring data illustrate that malathion has the potential for movement into groundwater, especially on soils with low organic matter and high sand content.

Cheminova disputes the ground-water data reported in the PGWDB. In particular, it calls into question the analytical methods used to generate the data in the Virginia study. In addition, Cheminova indicates that the maximum detection in the study was 3.12 ppb, not 6.17 ppb. Noting Cheminova's doubts for the Virginia data, EFED suggests a ground-water concentration estimate of 3.1 ppb for malathion for human-health risk assessment. This value is more conservative than SCI-GROW modeling results using maximal parameters for use on cotton (0.142 ppb). Since this monitoring result is specific for malathion EFED assumes the concentration of malaoxon will not exceed the concentration of malathion. Thus, EFED suggests conservative ground water concentration estimates of 3.1 ppb for malathion and 3.1 ppb for malaoxon.

The fact that malathion is detected in ground water means that under certain conditions it persists longer than its reported 1 to 3 day half lives emphasizing the need for further studies defining degradation rates under unfavorable conditions.

### 3. Ecological Effects Hazard Assessment

#### Mode of Action Summary

Several reviews of malathion and organophosphate toxicology exist including Matsumura (1985).

Malathion's mode of action is through acetylcholinesterase (AChE) inhibition which disrupts nervous system function. AChE is a enzyme made of protein which cleaves the neurotransmitter acetylcholine in nervous system junctions. Inhibiting this enzyme leads to accumulation of the neurotransmitter thus causing signals in the nervous system to persist longer than normal. Typical symptoms for pesticides which act in this manner are defecation, urination, lacrimation, muscular twitching and weakness, and halted respiration. Malathion, along with other phosphorodithioate insecticides (those containing two sulfur atoms bonded to phosphorus) must be oxidized before they have inhibitory potency and toxicity. Oxidation occurs via cytochrome p450 and results in the conversion of the P=S group in malathion to P=O forming its oxon, malaaxon (Murphy et al., 1968). This alteration of the phosphate group enables the molecule to covalently bind AChE resulting in long lasting inhibition of the enzyme.

Malaaxon binds to AChE by mimicking the structure of enzyme's natural substrate, acetylcholine. The similarity between the size, shape, and properties of malaaxon and the neurotransmitter allow it to "fit" in the acetylcholine binding site on the enzyme. Altering the structure of malaaxon or malathion reduces the ability of the oxon to bind AChE resulting in detoxification of the molecule. Detoxification reactions may be a result of enzyme or chemical action on the molecule. Detoxification occurs very rapidly in mammals giving pure malathion a very low acute toxicity [LD50 in rats is 12,500 mg/kg (Fukuto 1983)]. Common detoxification reactions for malathion (and malaaxon) are ester hydrolysis, demethylation, and phosphorothiolate ester hydrolysis. When one or more of these detoxification steps are blocked by another chemical the toxicity of malathion is increased and the added chemical is considered to synergize malathion toxicity. Chemicals which increase the rate of malathion's conversion to malaaxon may also be synergists.

Important detoxification steps occur through nonspecific esterase enzymes which are capable of cleaving malathion to less toxic degradates. Biological and environmental degradates of malathion with greatly lowered toxicity include malathion  $\alpha$ ,  $\beta$ , and diacids, and O-desmethyl malathion (Matsumura 1985). Since organophosphate insecticides are inhibitors of esterases (most specifically AChE) they possess the ability to block detoxification enzymes. Several organophosphate impurities present in technical malathion are known to synergize malathion toxicity probably through blocking malathion detoxification. The toxicity of several malathion impurities alone is also very high (eg the LD50 of O,O,S-trimethyl phosphorothioate in rats is 15

mg/kg, or 833 times more toxic than pure malathion) and cause delayed toxicity suggesting a mode of action other than AChE inhibition. Impurities can be produced through improper storage of malathion as evidenced by a 35% increase in the acute toxicity of technical malathion stored at 40°C for 6 months (Fukuto 1983).

## Toxicity to Terrestrial Animals

### 1. Birds, Acute and Subacute

An acute avian oral toxicity study using the technical grade of the active ingredient (TGAI) is required to establish the toxicity of pesticides to birds. The preferred test species is either mallard duck (a waterfowl) or bobwhite quail (an upland gamebird). Results of avian oral acute tests with Malathion are tabulated below.

**Table 15. Avian Acute Oral Toxicity**

Species	%ai	LD50(mg/Kg) (CL's)	Toxicity Category	MRID	Author	Classi- fication <sup>1</sup>
Mallard duck	95	14D LD <sub>50</sub> =1485 (1020-2150)	Slightly toxic	00160000	Hudson, R.H. and Tucker, 1984, USFWS	Core
Ring-necked pheasant	95	14D LD <sub>50</sub> =167 (120-231)	Moderate	00160000	Hudson R.H. and Tucker, 1984, USFWS	Supple- mental
Horned lark	95	14D LD <sub>50</sub> =403 (247-658)	Moderate	00160000	Hudson, R.H. and Tucker, 1984, USFWS	Supple- mental
Sharp tailed grouse	tech	LD50 =220 (171-240)	Moderate	Reference	Crabtree, D.G., 1965, Denver Wildlife Res. Center, USFWS	Supple- mental

<sup>1</sup> Core (study satisfies guideline). Supplemental (study is scientifically sound, but does not satisfy guideline)

Based on the data reviewed to date malathion displays low to moderate acute oral toxicity to the 3 species of birds tested by USFWS laboratories. Another referenced study was mentioned in the 1966 McEwen and Brown field study with Sharp tailed grouse (see field study section of this document). The study was conducted at the USFWS Denver Wildlife Research Center and is considered valid supplemental data. McEwen and Brown observed a similar LD50 with wild caught grouse. The most sensitive species tested was the ring-necked pheasant. The acute oral data does fulfill 71-1 testing guidelines.

Two subacute dietary studies using the TGAI are required to establish the toxicity of a pesticide to birds. The preferred test species are mallard duck and bobwhite quail. Results of subacute dietary tests with malathion conducted by USFWS laboratories are tabulated in table 16 below.

**Table 16. Avian Subacute Dietary Toxicity**

Species	%ai	LC50(ppm)	Toxicity Category	MRID	Author	Classification
Ring-necked pheasant	95	8D LC <sub>50</sub> =2639 (2220-3098)	Slightly toxic	00022923	Hill, E.F. et al USFWS, 1975	Core
Bobwhite quail	95	8D LC <sub>50</sub> =3497 (2959-4011)	Slightly toxic	00022923	Hill, E.F. et al USFWS, 1975	Core
Japanese quail	95	8D LC <sub>50</sub> =2962 (2453-3656)	Slightly toxic	00022923	Hill, E.F. et al USFWS, 1975	Supplemental
Mallard duck	95	8D LC <sub>50</sub> >5000	Nearly non-toxic	00022923	Hill, E.F. et al USFWS, 1975	Core

Based on the test results reviewed to date malathion displays low toxicity to 4 avian species on a subacute dietary basis. These studies were not submitted or funded by the registrant, but were conducted at the Patuxent Wildlife Research Center by U.S. Fish and Wildlife Service researchers. These studies are considered acceptable by the Agency and fulfill 71-2 guideline requirements.

**2. Birds, Chronic**

Avian reproduction studies using the TGAI are required for malathion because the following conditions are met: (1) birds may be subject to repeated or continuous exposure to the pesticide, especially preceding or during the breeding season, and (2) information derived from mammalian reproduction studies indicates reproduction in terrestrial vertebrates may be adversely affected by the anticipated use of the product. The preferred test species are mallard duck and bobwhite quail. Results of these tests are tabulated below.

**Table 17. Avian Reproductive Sensitivity**

Species	%ai	LOEL Effected Parameters	NOEL	MRID	Author	Classification
Bobwhite quail	96.4	21WK LOEL=350 ppm -regressed ovaries and reduced egg hatch At 1200 ppm reduced shell thickness, # eggs layed , egg viability	110 ppm	43501501	Beavers, J. Wildlife Intl., 1995	Core
Mallard duck	94.0	20WK LOEL =2400 ppm Growth and viability	1200 ppm	42782101	Biolife Assoc. 1993	Core

62

The guideline (71-4) is fulfilled by the studies above. Chronic exposure to malathion in diets produced moderate toxicity to terrestrial avian species and low toxicity to waterfowl species tested to date. At food exposure concentrations of 350 ppm female bobwhite quail exposed to malathion for 10 weeks displayed regressed ovaries and abnormally enlarged/flaccid gizzards. A reduction in numbers of eggs hatched from eggs set was observed at 350 ppm. Reduced egg production, viability of eggs, and embryo survival as well as an increase in the number of cracked eggs (a possible indication of the weakening of the shell structure) was observed at 1200 ppm.

### **Non Guideline Laboratory Studies with Avian Species**

**Malathion - Induced Teratisms in the Developing Chick. Greenburg, J. and N. Latham, 1968. University of Ottawa.**

Methods: Fertile white leghorn chicken eggs were injected via small holes in the end of the shell with 0.1 ml solution of malathion in corn oil using a 20 gauge 1 inch needle. A single injection was administered to eggs at various stages of development (4-12 days incubation time)

Results: 0.1 ml of solution injected into Leghorn Chicken eggs proved lethal to 50% of the embryos after 7 days (dependent on age). 4-5 days embryos were most susceptible.

Abnormalities included lack of feathers, smaller size, beak, plumage and hind limb defects.

In other studies where malathion was injected into eggs at 50 mg/egg chicks showed shortening of legs and bleaching of feathers (Marliac and Mutchler, 1963). For hen eggs injected with 25, 100, 200, 300, 400, and 500 ppm of malathion dissolved in acetone hatchability was significantly reduced at higher levels with hatches of 85%, 87%, 62%, 71%, 42%, and 6%, respectively (Dunachie and Fletcher, 1969). A number of studies were conducted where malathion or malaoxon were injected into chick embryos (Walker, 1971 and Khera and Lyon, 1968). Malaoxon caused reduced survival of embryos at a concentration of 30 micromoles, and those that did survive had severe abnormalities. Malathion at 15 micromoles produced less severe abnormalities.

### **Field Observations of Effects to Birds from Use of Malathion**

**The Ecology of a Small Forested Watershed Treated with the Insecticide Malathion S<sup>35</sup>. S.Giles, Robert H., Jr., 1970, Published by the Wildlife Society.**

Methods: Aerial Application - 2 adjoining Ohio watersheds were observed - with one treated and the other untreated. Malathion was radio tagged with Sulfur 35 radio nuclide. Two 20 acre watersheds were selected for comparison and were primarily deciduous forests. Application rate was 2 lbs/acre and 4 applications were made. Spray residue cards were placed under application areas for residue analysis. Residue collection discs were also suspended above the canopy using helium filled balloons. Glass discs were placed in the trees as well as the shrubs and in soil/litter surfaces.

Summary of Results: (Note; this was a very extensive study and the summary only briefly touches on actual amount of reported data).

Vegetation: Radioactivity was high in the tissues of plants sampled in the treated areas indicating active systemic uptake of malathion. New shoots and leaves showed especially high levels of radioactivity. Metabolites of malathion showed up in new stem and leaf growth up to one year after application.

Bacteria and soil fungi population: No effects observed. Fungi had higher concentrations of radioactivity than surrounding plant tissues.

Soil micro arthropods: Briefly affected.

Earthworms and Snails: No statistically significant effects observed in natural populations. No radioactivity was measured in snail tissues. However, in experimental cans containing soil and earthworms, more dead earthworms were found in the treated plots and several had radioactivity within their tissues suggesting uptake of the radio labeled malathion.

Fish and Crayfish: Stream samples showed no significant effects to mudminnows or blacknose dace. However, effects to brook sticklebacks, which appeared more sensitive, were extensive with over 95% mortality in the treated area. Caged crayfish appeared unaffected.

Insect numbers (all observed species): Greatly reduced, but populations recovered quickly. In stream nets located at temporary dams dead insect numbers were 1270 1 hour after spraying and decreased to 640 and 598 individuals collected 2 and 3 hours after spraying, respectively.

Reptiles and Amphibians: No effects noted - Toads adsorbed high loads of residues into tissues. However the route of intake of residues was not certain (skin adsorption, ingestion, etc)

Birds: Showed some reaction up to 48 hours post application, but no lasting effects noted. Lack of singing was observed throughout treated areas immediately after application and persisted for 2 days. By day 4 singing intensity was equal in treated and control areas. Possible explanations include sublethal insecticidal response, behavioral response due to loss of food, or possibly temporary emigration from the treated areas. Some radioactivity was detected in collected bird's whole organ tissues. Insectivorous birds had the highest detection of radioactivity on feathers.

Mouse, shrew and chipmunk populations: Up to a 45% reduction in population of white footed mice *Peromyscus leucopus novaboracensis* was estimated for the treated areas, based on pre and post treatment trapping counts. No difference in populations of short-tailed shrews or black-tailed shrews was determined, though residues were detected in costal cartilage, kidney, and heart tissues samples. Chipmunk populations were reduced 55% in treated areas following applications. The study author concludes "As with the mice this is not a lethal effect, but apparently one of productivity and survival."

Large mammals: Appeared unaffected.

### **Use of Enzyme Profiles to Monitor Residues in Wildlife. Dieter, Michael P., 1975 USFWS, Patuxent Wildlife Research Center.**

Methods: Starlings were fed varying concentrations of various pesticides and plasma enzyme levels were measured.

Results: Starlings fed 160 ppm of malathion for 12 weeks showed 30% decrease in AChE and 50% decrease in 1 acetate dehydrogenase activity.



**Effects of Ultra-Low Volume Applications of Malathion in Hale County Texas Hill, Elwood F. et al., 1971. Journal of Med Entomology**

Methods: A number of field observations were recorded following 9 ULV aerial applications of malathion over Hale County by C123 cargo planes for mosquito control. During this operation nontarget insects and wildlife were also monitored for possible adverse effects. Nontarget insects were sampled twice weekly using sweep nets and traps (vehicle mounted). House sparrows and other species were surveyed, particularly insectivorous species. 116 sparrows were banded prior to spraying in Plainview, Texas. Carcass searches were also conducted post application. AChE levels were determined in the sparrows prior to spraying and 30 hrs. following applications 1, 3, 5 and 7. Diptera, Hemiptera, and Coleoptera were main insect groups collected in sweep nets and traps.

Results: Only 9.5% of the banded sparrows (11 of 116) were recaptured. No bird carcasses were recovered. Brain AChE levels in the captured sparrows were not significantly inhibited - but a slight reduction from 0.023 to 0.018 was observed. Unprotected honeybees were killed, but covered hives were not seriously effected.

**Acute Toxicity of Dieldrin and Malathion to Wild Sharp-tailed Grouse. McEwen, Lowell and Robert L. Brown. Journal of Wildlife management. Vol. 30, No. 3, July 1966. MRID 113233**

Methods: 52 live trapped grouse from Montana were given oral doses of dieldrin, malathion, and lactose (controls) and released after tagging. They were subsequently observed by capture or radio tracking.

Malathion Results: The lethal dose of malathion was observed to occur between 200-240 mg/kg. Reaction to malathion occurred within 72 hours - either death or full recovery. Sublethal signs included depression, slow reactions, blinking, head nodding, and eventual heart or respiratory failure. Radio tracked grouse displayed normal to severe reactions once released. 8 of 12 birds were recovered. Predators are suspected in the disappearance of unrecovered birds (in one case a bird moderately dosed with dieldrin was confirmed killed by a coyote). Grouse that were dosed carried transmitters up to 12 days after release. All confirmed predator kills had received what were considered sublethal doses of the test material. Other birds were discovered to have been attacked and injured. The radio transmitters did not hinder all birds as many were recovered in healthy condition. The sublethal effect of the malathion and dieldrin are suspected. All controls survived and successfully bred.

**Physiological Effects of Malathion on the House Sparrow *Passer domesticus*. Mehrotra, K. N. et al, 1966. Indian Agricultural Research Institute Delhi, India.**

Methods: Birds were offered treated grain with 5% ai malathion dust. (Concentration 56.7g/56.8 kg of grain or approximately 100 ppm) to determine deterrent effect. Oral doses were administered at 1,2,5 and 10 mg/bird in acetone or approximately 50, 100, 250, and 500 mg/kg based on mean avg wt.

Results: Sparrows showed 75% reduction in feeding on treated seed vs. untreated seed (4 grams of treated seed consumed vs 21 gms. of untreated seed on average). Orally dosed birds showed increased respiration, head droop, ejection of white fluid from mouth, chronic and tonic convulsions at 5 mg/kg or more. Birds that did recover did so in about 1 hour. AChE inhibition was 83%, 75%, 50% and 25% at 19 mg, 5 mg, 2 mg and 1 mg per kg of body weight, respectively within 5 minutes of ingestion. The 1 and 2 mg/Kg dosed birds recovered in 24 hours. 57% and 18% mortality was observed at 10 and 5 mg/kg feed residue concentrations.

**The Effects on Quail, Migratory Birds and Non-Game Birds from Application of Malathion and Other Insecticides. Parsons, Jack K. and Billy Don Davis, Texas Parks and Wildlife Dept., 1971. Study conducted from 1964 to 1968.**

Methods: During bollweevil control programs on cotton game and nongame bird populations near cotton fields were observed. Applications was aerial at 12 to 16 oz. (approx. 1.2 lb ai) of technical malathion per acre. Up to 7 applications were made at 5-22 day intervals. Sites were <160 acres with one site containing a lake where waterfowl visited. Caged quail were placed in fields in four groups each containing 10-12 birds each. Birds were aged and weighed. Once birds were exposed to spray they were returned to a lab shelter area. Exposed feed and insects were fed to the test birds. Thorough carcass searches were made 24 hours after spraying occurred.

Results: No major differences in weight gain were noted between treated and control birds. No toxicant related mortality was noted after 3 application of malathion. No dead birds were located adjacent to fields. Sublethal indicators other than weight were not measured (e.g. AChE levels).

**Brain Cholinesterase Activity in Birds After a City-Wide Aerial Application of Malathion. Kucera, Emil. Manitoba Dept. of Environment and Workplace Safety and Health. 1987.**

Methods: Aerial application of malathion was made over Winnipeg in July 1983 after health officials determined that there was concern for possible outbreak of equine encephalitis from mosquito population increases. Malathion was applied as ULV solution using 95% malathion. Application rate was 210 ml/ha over the entire city to control mosquitos. House sparrows and pigeons were monitored for AChE levels prior to spraying and post application. Dead birds were also analyzed when reported to the researchers by the public. 20 caged sparrows were placed in a local park where application was made.

Results: Actual deposition rate for sprayed area was 140 ml/ha. and the area was monitored for 3 hours post application. 41 sparrows and 39 pigeons were collected within 2 weeks of spraying. Caged exposed sparrows were sacrificed and examined as well. No significant differences were noted (6-12% variation) in AChE levels of post spray--to prespray birds. Some reservation is expressed that study birds may all have been exposed to ground fogging applications prior to aerial application exposure.

**Fluctuations in Bird Populations on the Island of Bota as Related to an Experimental Program to Control The Melon Fly. John Engbring, 1989. U.S. Fish and Wildlife Service, Honolulu, Hawaii.**

Methods: An experimental program to control melon flies on the Island of Bota in the Northern Marianas Islands, provided the USFWS with an opportunity to monitor avian populations while subjected to exposure to malathion laced bait sprays (Cue-lure) that were aeriially applied. Survey stations were established at 164 locations 150 meters apart in 8 different transects located on back roads adjacent to forested/semi-forested areas. 8 minute counts were conducted at each station within 4 hours of sunrise before, during, and after the application of the pesticide baits. A total of 1,169 eight minute counts were made, 368 in Aug. 1988, 432 in December 1988, and 369 in Aug. 1989. Applications were made at 3 week intervals beginning in Oct. 1988 at up to 5 -30 gms/hectare depending on bait type.

Results: Of the 10 native species counted 5 increased in number and 5 decreased. The author was not certain if this was a normal annual fluctuation or one caused by pesticides. Populations of the white throated ground dove, the Phillipine turtle dove, and possibly the bridled white eye were significantly lower in the following year. No acute mortality was reported. The other 20 native species were observed and populations appeared unaffected. Even insectivorous species did not appear to suffer population decreases.

**Mammals, Acute and Chronic Toxicity**

Wild mammal acute toxicity testing is required on a case-by-case basis, depending on the results of lower tier laboratory mammalian studies, intended use pattern and pertinent environmental fate characteristics. In most cases, rat or mouse acute toxicity values are obtained from the Agency's Health Effects Division (HED) and substitute for wild mammal testing. These toxicity values are reported below.

**Table 18. Mammalian Acute Oral and Chronic Dietary Toxicity**

Specie	%ai	LD50 (mg/Kg)	Study ID	NOEL (parameter)	Study ID and Author	Cate-gory
Mice	Tech 90%	720-886	Doc #000389	500 ppm ( growth -2 yr. chronic study)	Reference Doc#000389 N.C.I.,1979	N.R.
Guinea pig	Tech	570	Doc # 000389	Not determined	Reference Doc#000389 Pham.Exp.Ther. 12:327,1953	N.R.
Sheep	Tech	<150	Doc # 000389	Not determined	Reference Doc# 000389 J.Vet.Am.Medical Ass., 1957	N.R.

67

Cow	95%	560(adult ) 80 (calf)	Doc # 000389	Not determined	Ref Doc # 000389 Golz and Shaffer, 1956	
Cat	Tech	>500	Doc # 000389	Not determined	Reference Doc# 000389	N.R.
Rat (Wistar albino)	57% EL	1763	Doc # 000317	ataxia, tremors, salivation, diarrhea observed	Reference Doc#000317 Doc# 000389	Suppl.
Rat	Tech	390-2100	Doc # 000389 Am. Cyanami d 1956	1000 ppm(growth) 32 day ChE reduction-100 ppm 240 mg/kg/day reduced pup survival and BW	Reference Doc#000389 Karlow and Martin , 1965	N.R.
Rabbit	Tech	>900	Doc# 000389, J. Econ. Entomol . 48:139	50 mg/kg/day resorption of embryos	MRID 260123 Ref.Doc.#000389 Food and Drug Research Lab,1984	N.R.

In one rat study oral LD50 ranged from 1000 to 1845 mg/Kg with females being more sensitive than males.

The results of mammalian laboratory indicate that malathion is slightly toxic to mammals on an acute oral basis. Sublethal effects may occur at concentrations as low as 100 mg/Kg for certain species. Reproductive effects are not expected unless concentration remain at 500-1000 mg/Kg for extended periods of time.

## Non Guideline Studies with Mammals

### Toxicity of Malathion to Mammals, Aquatic Organism and Tissue Culture Desi, I. et al 1976.

Results: In white rats alteration of EEG (elevated excitability of nervous system) at 75 and 438 mg/kg, and reduced acetylcholinesterase levels were observed. The test rats exposed to malathion showed a reduced ability to run a maze with increased numbers of errors.

### Comparative Metabolism of Malathion - C<sub>14</sub> in Plants and Animals. Bourke, J.B. et al New York State Agricultural Experiment Station, Cornell University, 1968.

Methods: Red Kidney bean plants were exposed to radio labeled malathion solution by passing air over foliage for 20-30 minutes (with solution mixed into air stream) C<sup>14</sup> was traced in tissues of plants for 14 days. Various intermediates (metabolites) were deposited within tissues. Rats were treated with malathion via oral ingestion.

Results: Plants appeared to store various metabolites in tissues whereas the rats began excreting the radio labeled malathion in their urine within 2 hours after ingestion. By 24 hours 83.4% of the radio labeled material was excreted in the urine.

**In the 1970 study by R.H. Giles (see previous summary)** certain species of small mammals(white-footed mice and chipmunks) showed significant population reductions(30-55%) after treatment of a forest area with 2 lb ai/A of malathion. Larger mammals and interestingly, shrews which are often sensitive due to high metabolisms, were not observed to have been effected. Population reductions were not observed to be related to acute adult mortality, but rather to reduced reproductive success or possibly effects on survival of young.

### **Nontarget Reptile Toxicity**

The Agency has not reviewed extensive laboratory toxicity data pertaining to toxicity of malathion to reptiles. In general, the Agency uses avian toxicity thresholds in the determination of hazard to reptiles. One study was found regarding acute effects to the Carolina anole and another study which indicated possible effects to developing embryos of snapping turtles.

**Responses of the Iguanid Lizard, *Anolis carolinensis* to Four Organophosphorous Pesticides, Hall R.J. and D.R. Clark, 1982. Environmental Pollution (Ser. A) 28:45-52**  
Methods: Oral ingestion of organophosphate pesticides and the resulting percent mortality was measured for Carolina anoles.

Results For malathion the acute LD50 was determined to be 2324 mg/Kg

**Teratogenic Effects of Malathion and Captan in the Embryo of Common Snapping Turtle. Mitchell and Yutema, 1973.**

Results: Observations of the abnormal development in of embryos of the common snapping turtle exposed to malathion is summarized.

### **Non-target Beneficial Insect Toxicity**

#### **Terrestrial Insects**

A honey bee acute contact study using the TGAI is required for malathion insecticide products because uses will result in honey bee exposure. A honey bee foliar residue contact toxicity study is required using the typical end-use product because many uses will result in potentially adverse honey bee exposure to vegetative surfaces after application. Results are tabulated below.

**Table 19. Nontarget Pollinator Insect Acute and Foliar Contact Toxicity**

Species	%ai	LD50 (ug ai/Bee)	Toxicity Category	MRID	Author/ year	Classification
Honey bee	57EC	8 HR Foliar Contact LD <sub>50</sub> <1.6	Highly toxic	41208001 41284701	1989	Core
Honey bee	Tech	48 HR LD <sub>50</sub> =0.20	Highly toxic	05001991	1978	Core
Honey bee	Tech	96 HR LD <sub>50</sub> =0.709	Highly toxic	0001999	1967	Core
Honey bee	Tech	N.R. LD <sub>50</sub> = 0.27 (0.22-0.29)	Highly toxic	05004151	1968	Core
Honey bee	Tech	48 HR LD <sub>50</sub> =0.38	Highly toxic	05004003	1968	Core

The results indicate that malathion is highly toxic to bees on an acute contact basis either through exposure to direct spray or through foliar residue contact within 8 hours after spray is applied. The guidelines 141-1 and 141-2 are fulfilled by these studies.

**Toxicity to Terrestrial Insects with Aquatic Lifestages**

Though not required, a number malathion studies on toxicity to aquatic insect larvae conducted by the U.S. Fish and Wildlife Service laboratories were reviewed. These studies are indicative of possible population effects to species of insects which spend the early stages of their lives as aquatic larvae. Many of these species form important links in the food chain for insectivorous birds, mammals, fish, amphibians and reptiles as well as predatory aquatic and terrestrial invertebrates.

**Table 20. Toxicity to Aquatic Larvae of Terrestrial Insects**

Species	% ai	LC <sub>50</sub> (C.L.s) in PPB	Toxicity	MRID	Author/Year	Classifi- cation
Stonefly <i>Claasenia sabulosa</i>	95%	LC50=2.8 (1.4-4.3)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Supplemental
Stonefly, <i>Pteronarcella badia</i>	95%	LC <sub>50</sub> =1.1 (0.78-1.5)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Supplemental
Stonefly, <i>Isoperla</i> sp.	95%	LC <sub>50</sub> =0.69 (0.2-2.4)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Supplemental
Damselfly, <i>Lestes congener</i>	95%	LC <sub>50</sub> =10 (6.5-15.0)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Supplemental
Caddisfly, <i>Hydropsyche</i> sp.	95%	LC <sub>50</sub> =5.0 (2.9-8.6)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Supplemental

76

Caddisfly, <i>Limnephilus</i> sp.	95%	LC <sub>50</sub> =1.3 (0.77-2.0)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Supplemental
Snipefly, <i>Atherix variegata</i>	95%	LC <sub>50</sub> =385 (245-602)	High	40098001	Mayer and Ellersieck, USFWS, 1984	Supplemental

Based on the data reviewed to date for aquatic early life stages of terrestrial non-target insects malathion is classified as highly to very highly toxic to larvae of these species, and thus, their successful reproduction and survival to adult stages.

### Field Observations of Effects to Non-Target Insects

Over its long history a great amount of field testing has been conducted with Malathion products in field test situations by agricultural research laboratories, registrants, and government research laboratories. A number of these studies are briefly summarized below along with results that were reported from exposure to malathion to a variety of terrestrial species.

#### California Medfly Report. Oshima, R. S. 1982

Methods: During a six week period - baited sprays were applied with large droplet sizes (200-300  $\mu$  mean diameter).

Results: Malathion and malaoxon were detected in water throughout monitoring period. Rain runoff to storm drains produced concentrations up to 583 ppb in existing streams. Since bait sprays did not attract honeybees it was believed that they would be unaffected. However, nontargeted lacewings and dipterids were attracted (mainly scavenger flies) to the bait and killed.

#### Impact of Mediterranean Fruit Fly Malathion Bait Spray on Honeybees. Gary, Norman E. and Eric C., Mussen, Dept. of Entomology, Univ. of California, Davis 1984.

Methods: The application rate was 175.4 ml malathion technical and 701.8 ml Staley's Protein Bait/hectare applied weekly at predawn in San Francisco Bay area. 48 commercial honeybee colonies were distributed evenly to 4 apiaries-two inside spray zone. Control colonies were located at 5 km and 9 km outside spray zone. Bodies were collected daily at hive entrances. Pollen samples also were taken daily. A similar study was conducted in Stockton, CA. with one apiary located inside the spray zone and one control placed 5.7 km outside spray zone. Each apiary in the Stockton study contained 10 bee colonies.

Results: Significant mortality was observed 48 hours following each application. Cause was determined to be pesticide contaminated pollen (2.06 ppm (mean)) and body residue levels of 0.9 -5.3 ppm. Data from Stockton study also showed increased mortality which was partially attributable to nearby application of Sevin, (alfalfa fields), Kabbate and sulfur dust (tomato fields). Reduced flight activity was observed at both exposed sites after pesticide applications. Other mortality may have occurred in the fields (not measurable).

**Effects of Malathion Bait Spray for Mediterranean Fruit Fly on Non-target Organisms on Urban Trees in Northern California. Dahlsten, Donald L., University of California, 1983.**

Methods: Protein hydrolysate bait spray and malathion applications were monitored. The study measured effects to nontarget beneficial insect predators common in urban trees. Drop cloths were placed under trees and dead fallen insects were collected and identified. 3 Trees were chosen randomly in each of the selected spray areas. 2 trees were chosen in control plots. 2 drop cloths were placed under each tree. Test site locations were in Pleasanton, CA and Woodside, CA.

Results: 17 species of aphids, numerous dipterids, butterfly (lepidoptera) larvae, spiders, cynipoidea, and hemiptera appeared to be heavily effected. Also pscoptera were reduced.

**Effects of Ten Organic Insecticides on Two Species of Stonefly Naiads. Jensen, Loren D. and Anden R. Gaufin, Dept. Zoology, University of Utah, 1964.(MRID 00065497)**

Methods: The 96 hour acute exposures under static conditions included malathion exposure.

Results: Even when the naiads were removed after displaying sublethal effects (convulsions or tremors) and placed in clean water, they generally died within 24 hours.

**Bee Poisoning Hazard of Undiluted Malathion Applied to Alfalfa in Bloom. Johansen, C.A. et al. Washington State University College of Agriculture. 1965.**

Methods: Six colonies of honeybees were placed in a 125 acre alfalfa field 36 hours before aerial application. 2 hives were covered with wet burlap during application and burlap was removed 24 and 48 hours post application. Two hives were left uncovered in the sprayed fields. 2 other colonies were placed 2.25 miles from the application site and one of these was covered with burlap for 48 hours. 8 fluid oz of malathion ULV concentrate was applied per acre by aerial spray at 50 feet altitude in a 125 foot swath on Aug 14. Package bee cages (150-200 bees) were also placed in fields 2 and 7 hours after application for a 3 hour exposure period. Caged bees were also exposed to foliar residue samples at intervals following the application.

Results: Bee mortality was higher than normal for 4 days after application. Those covered with wet burlap suffered the highest mortality 1 day after the covers were removed. Bees caged on treated foliage also exhibited higher than normal mortality. Check (control) colonies showed between 500 and 838 dead bees at hive entrances. Treated hive mortalities ranged from 1298 ((unprotected) to 2582 (entrance covered) honeybees. Bees which contacted treated foliage showed from 100% (2 hours-1 day post application) mortality to 14% mortality (4 day old residues) versus an average of 5% mortality for control bees. Malathion residues on foliage ranged from 28.8 ppm at application to 0.4 ppm 14 days after application. Residues remained over 25 ppm for 4 days following application after which a rainfall event occurred. Grasshopper populations were greatly reduced from 12/sq. yd. to less than 1/sq yd. three days after treatment. Lygus bugs were also controlled for up to 3 weeks. Interestingly, the target organisms, Alfalfa weevils and larvae, were not totally controlled. Lady beetles populations were reduced for up to 3 weeks following applications.

**Terrestrial Wildlife Field Incidents**



The Agency reviews and records all wildlife mortality incidents reported independently or under 6a2 provisions of FIFRA regarding use of pesticides or pesticide mixtures. These incidents are reported to the Agency by a variety of sources including registrants, private organizations and local, state, or federal agencies. A summary of all terrestrial incidents reviewed by the Agency following use of malathion products or mixtures is provided in table 21 below.

**Table 21**

Location and Date	Incident	Description	Probability
Oregon, 1/1/85	I000130	5000 acres of alfalfa treated with malathion by USDA- extensive mortality of honeybees collecting nectar from blossoms reported	Probable
Florida, 1997 Medfly Program, Hillsborough County area	USDA Medfly Incident Report	Three incidents involving mortality of ducks were reported along with over 40 fish kills that were investigated. All occurred where malathion bait formulations were used near ponds. 6/22-10 to 14 Ducks killed-Seminole Hts.-baits used 6/14-Duck kill-NW Hillsborough sector-baits used 6/25-Duck kill-Rodrie pond-baits applied acrially	Possible- but unlikely. Only routes believed to offer logical exposure route- oral ingestion of baits or dermal exposure- residue concentration too low to =LD50.

The incidents where duck mortality was reported in Florida medfly program investigations were determined to be more likely caused by some other toxicant. Though fish kills did occur in the ponds, actual residues were well under those which would be expected to cause oral toxicity in mallard duck(1485 mg/Kg). In the case of the June 14 fish kill an oily substance was observed on the moribund ducks. Park service personnel had also sprayed herbicides near the pond (Glyphosate and Copper). Maximum malathion concentration on vegetation was only 3.0 ppm far below avian toxicity thresholds. The Agency would tend to agree with USDA that malathion was not the primary cause of death in the duck kill incidents.

### Acute Toxicity to Fish

Two freshwater fish acute toxicity studies using the TGAI are required to establish the toxicity of a pesticide to freshwater fish. The preferred test species are rainbow trout (a coldwater fish) and bluegill sunfish (a warmwater fish). Acute toxicity testing with estuarine/marine fish species using the TGAI is required for malathion because the end-use product is intended for direct application to the marine/estuarine environment and the active ingredient is expected to reach this environment because of its use near estuarine environments. The preferred estuarine test species is sheepshead minnow. Results of numerous tests reviewed by EPA are summarized below.

**Table 22. Freshwater(FW) and Marine/Estuarine(SW) Fish Acute Toxicity**

Species Tested	% ai	LC50 and CLs in PPB	Toxicity	MRID	Author	Classification
<b>Freshwater Fish Species</b>						
Bluegill sunfish(FW)	95	96 Hr LC <sub>50</sub> =20 (16-25)	Very High	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Redear sunfish(FW)	95	95 Hr LC <sub>50</sub> =62 (58-67)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Rainbow trout(FW)	95	96 Hr LC <sub>50</sub> =4 (2-7)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Yellow perch(FW)	95	96 Hr LC <sub>50</sub> =263 (205-338)	High	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Largemouth bass(FW)	95	96 Hr LC <sub>50</sub> =250 (229-310)	High	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Carp(FW)	95	96 Hr LC <sub>50</sub> =6590 (4920-8820)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1984	Supl.
Fathead minnow(FW)	95	96 Hr LC <sub>50</sub> =8650 (6450-11500)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Channel catfish(FW)	95	96 Hr LC <sub>50</sub> =7620 (5820-9970)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Species Tested	% ai	LC50 and CLs in PPB	Toxicity	MRID	Author	Classification
Coho salmon(FW)	95	96 Hr LC <sub>50</sub> 170 (160-180)	High	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Cutthroat trout(FW)	95	96 Hr LC <sub>50</sub> =174 (112-269)	High	40098001	Mayer and Ellersieck, USFWS 1984	Core
Brown trout(FW)	95	96 Hr LC <sub>50</sub> =101 (84-115)	High	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Lake trout(FW)	95	96 Hr LC <sub>50</sub> =76 (47-123)	High	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Black bullhead catfish(FW)	95	96 Hr LC <sub>50</sub> =11700 (9600-14100)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1984	Core

Green sunfish(FW)	95	96 Hr LC <sub>50</sub> =1460 (900-2340)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Walleye(FW)	95	96 Hr LC <sub>50</sub> =64 (59-70)	Very high	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Tilapia(FW)	95	96 Hr LC <sub>50</sub> =2000 (N.R.)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1984	Core
Goldfish(FW)	95	96 Hr LC <sub>50</sub> =10700 (8340-13800)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1984	Core
<b>Estuarine Marine Fish Species</b>						
Spot(SW)	95	Flowthrough 48 Hr LC <sub>50</sub> =320(N.R.)	High	40228401	F. L. Mayer, USEPA	Supl.
Striped mullet(SW)	95	Flowthrough 48 Hr LC <sub>50</sub> =330(N.R.)	High	40228401	F. L. Mayer, USEPA	Supl.
Longnose killifish(SW)	95	Flowthrough 48 Hr LC <sub>50</sub> =150(N.R.)	High	40228401	F.L. Mayer, USEPA	Supl.
Sheepshead minnow(SW)	95	Flowthrough 96 HR LC <sub>50</sub> =33.0(14-63)	Very high	41174301	Bowman, J 1989, ABC Laboratories	Core
Striped bass(SW)	95	96 Hr LC <sub>50</sub> =60(N.R.)	Very high	156311	Wellborn, T. 1971 Reference	Supl.
Sheepshead minnow(SW)	57 EC	96 HR LC <sub>50</sub> 55	Very high	41252101	Bowman, J. ABC Labs, 1989	Core

Based on the extensive data reviewed for malathion toxicity to freshwater and estuarine/marine fish the pesticide is classified as very highly to moderately toxic to fish dependent on the sensitivity of the tested species. In many cases these studies were done with static test systems, no measured concentrations, and varying pH levels which can influence the actual toxicity or calculation of toxicity values for a chemical with fate characteristics such as malathion. This is due to the hydrolytic instability of the compound. Thus, actual mean concentrations which caused the mortality may have been much lower after 96 hours of exposure than is indicated. This would have influenced the calculation of LC50 levels if they had been based on measured concentrations instead of nominal 0 hour concentrations. The 72-1 and 72-3 guidelines for acute toxicity testing of fish are fulfilled by the data reviewed above.

### Chronic Toxicity To Freshwater and Estuarine Fish

A freshwater fish early life-stage test and/or an estuarine fish early life stage test using the TGAI is required for malathion because some end-use products may be applied directly to water and other product uses are expected to contribute residues which may be transported to water from the various intended use sites. In addition the following chronic testing guideline conditions are met: the pesticide is intended for use such that its presence in water is likely to be recurrent, aquatic acute LC50 are less than 1 mg/l, the EECs in water are equal to or greater than 0.01 of any acute LC50, studies of other organisms indicate the reproductive physiology of fish may be affected. The preferred test species are the rainbow trout and the sheepshead minnow.

A freshwater fish full life-cycle test using the TGAI is required for malathion because the end-use product is intended to be applied directly to water and is expected to be transported to water from the intended use sites. In addition, the following conditions were met: the EEC is equal to or greater than one-tenth of the NOEL in the fish early life-stage or invertebrate life-cycle test, and studies of other organisms indicate the reproductive physiology of fish may be affected. The preferred test species is fathead minnow. A satisfactory full life cycle test has not been submitted, though a pilot lifecycle study with fathead minnow has been attempted. Results of this test are tabulated below.

An estuarine/marine fish early life stage or life-cycle test using the TGAI is required for malathion due to the application of malathion for mosquito and medfly control near estuarine habitats and use on crops associated with areas near these habitats. This study may be waived if further modeling results indicate that EEC levels in estuaries will not exceed the early life stage NOEC levels for a freshwater species, or if the registrant does not continue to support these uses. The preferred test species is sheepshead minnow.

**Table 23. Freshwater and Marine Fish Chronic Toxicity Test Results**

Species	Guide-line	%ai	LOEC in PPB	NOEC	MRID	Author	Cate-gory
Rainbow trout	72-4	94	97 day LOEC= 44	2 PPB	41422401	Cohle P., ABC Laboratories 1989	Core
Flagfish, <i>Jordanella floridae</i>	72-4	tech	110 day LOEC=11	8.6 PPB	Reference	Hermanutz,R., 1978*	Supple- mental
Fathead minnow	72-5	tech	158 day LOEC=350	N.D.	D234663	ABC Laboratories, 1997	Supple- mental

The guideline (72-4) is fulfilled for freshwater fish.

The guideline (72-4) is **not** fulfilled for a marine/estuarine fish species.  
 The guideline (72-5) is **not** fulfilled. Study aborted early due to malathion degradation problems.

\* Hermanutz, R. 1978. Endrin and Malathion toxicity to Flagfish (*Jordanella floridae*). Arch. Of Environmental Contaminants and Toxicology 7:159-168

### Toxicity of Malathion to Amphibian Lifestages

Though extensive literature has not been reviewed for toxicity of malathion to adult or larval life stages of amphibians, there are data to suggest that malathion may have adverse effect to amphibian early life stages if environmental concentrations exceed 1 ppm. In studies with developing frog embryos, gross abnormalities in skeletal development were noted for tadpoles which had been exposed for several days to malathion concentrations ranging from 1 to 20 ppm. Abnormalities observed included spinal curvature, blister development, and abnormal swimming behavior at concentrations ranging from 5-10 ppm. At concentrations >10 ppm malathion was highly embryo-toxic. (author(unknown) Dept. Of Zoology, University of Poona, India, Bulletin of Environmental Contamination Toxicology, 31:170-176(1983) Please Note: reference unverified-internet article). Mayer and Ellersieck have listed the acute toxicity of malathion to tadpoles of Fowlers toad, *Bufo woodhousei*, and the chorus frog, *Pseudacris triseriata* to be 420 ppb and 200 ppb, respectively. These acute values are considered highly toxic.

### Acute Toxicity to Freshwater and Marine/Estuarine Invertebrates

A freshwater aquatic invertebrate toxicity test using the TGAI is required to establish the toxicity of a pesticide to aquatic invertebrates. The preferred test species is *Daphnia magna*.

Acute toxicity testing with estuarine/marine invertebrates using the TGAI is required for malathion because the end-use product is intended for direct application to the marine/estuarine environment or the active ingredient is expected to reach this environment because of its use near estuarine habitats. The preferred test species are mysid and eastern oyster. Results of freshwater and estuarine invertebrate acute toxicity tests reviewed by the Agency are tabulated below.

**Table 24. Freshwater and Marine /Estuarine Invertebrate Acute Toxicity**

Species Tested FW=Freshwater SW=Marine species	% ai	EC50 or LC50 in PPB	Toxicity	MRID	Author	Classi- fication
<b>Freshwater Invertebrate Species</b>						
Water flea, <i>Daphnia pulex</i> (FW)	95	48 Hr EC <sub>50</sub> =1.8 (1.4-2.4)	Very high	40098001	Mayer and Ellersieck, USFWS, 1986	Core
Scud, <i>Gammarus lacustris</i> (FW)	tech	48 Hr LC <sub>50</sub> =1.8 (1.3-2.4)	Very high	05009242	FWS Laboratories, 1969	Core

Scud, <i>Gammarus fasciatus</i> (FW)	95	96 Hr. LC <sub>50</sub> =0.5 (N.R.)	Very high	40098001	Mayer and Ellersieck, USFWS, 1986	Core
Daphnid <i>Simocephalus serrulatus</i> (FW)	95	48 Hr LC <sub>50</sub> =0.69 (0.44-0.79)	Very high	40098001	Mayer and Ellersieck, USFWS, 1986	Supplemental
Crayfish, <i>Orconectes nais</i> (FW)	95	96 Hr LC <sub>50</sub> =180 (140-230)	High	40098001	Mayer and Ellersieck, USFWS, 1986	Supplemental
Glass shrimp, <i>Palaemonetes kadiakensis</i> (FW)	95	96 Hr LC <sub>50</sub> =12 (N.R.)	High	40098001	Mayer and Ellersieck, USFWS, 1986	Supplemental
Seed Shrimp, <i>Cypridopsis vidua</i> (FW)	95	49 Hr LC <sub>50</sub> =47 (32-69)	High	40098001	Mayer and Ellersieck, USFWS, 1986	Core
Water flea, <i>Daphnia magna</i> (FW)	57EC	48 Hr EC <sub>50</sub> =2.2 (1.9-2.5)	High	410297-01	Burgess, D. ABC Labs, 1989	Core
<b>Species Tested FW=Freshwater SW=Marine species</b>	<b>% ai</b>	<b>EC50 or LC50 in PPB</b>	<b>Toxicity</b>	<b>MRID</b>	<b>Author</b>	<b>Classification</b>
Water flea, <i>Daphnia magna</i> (FW)	95	48 Hr EC <sub>50</sub> =1.0 (0.7-1.4)	High	40098001	Mayer and Ellersieck, USFWS, 1986	Core
Sowbug, <i>Asellus brevicaudus</i> (FW)	95	96 Hr LC <sub>50</sub> =3000 (1500-8500)	Moderate	40098001	Mayer and Ellersieck, USFWS, 1986	Supplemental
<b>Estuarine Marine Species</b>						
Mysid, <i>Mysidopsis bahia</i> (SW)	94	96 Hr LC <sub>50</sub> =2.2 (1.5-2.6)	very high	41474501	Forbis, A., ABC Lab., 1990	Core
Pink shrimp, <i>Penaeus duorarum</i> (SW)	95	48 Hr LC <sub>50</sub> =280 (N.R.)	High	40228401	F.L. Mayer, USEPA, 1986	Supplemental
Eastern oyster, <i>Crassostrea virginica</i> (SW)	95	96 Hr LC <sub>50</sub> >1000	Not conclusive	40228401	F.L. Mayer, USEPA, 1986	Supplemental
Eastern oyster, <i>Crassostrea virginica</i> (SW)	57% EC	96 Hr EC <sub>50</sub> =2960 (N.R.)	Moderate	42249901	Wade, B and J. Wisk, ESE, Inc. 1992	Core
Blue Crab, <i>Callinectes sapidus</i> (SW)	95	48 Hr LC <sub>50</sub> >1000	Not conclusive	40228401	F.L. Mayer, USEPA, 1986	Supplemental

Since the LC50/EC50 values are in the range of 0.5 to 3000 PPB, malathion is classified as very highly to moderately toxic to aquatic invertebrates on an acute basis, dependent on the sensitivity of the tested species. Many of the studies above were conducted under static conditions with no measurement of actual residue levels. Thus, actual LC<sub>50</sub> values might have been even lower than those reported if they had been based on measured concentrations (expected to be lower due to degradation). The guidelines 72-2 and 72-3 for invertebrate acute testing are fulfilled by these studies.

### Chronic Toxicity to Freshwater and Marine Invertebrates

Freshwater and estuarine/marine aquatic invertebrate life-cycle tests using the TGAI is required for malathion since the end-use product may be applied directly to water or is expected to be transported to water from the intended use site, and the following conditions are met: (1) the pesticide is intended for use such that its presence in water is likely to be continuous or recurrent regardless of toxicity, (2) aquatic acute LC50 or EC50 are less than 1 mg/l, and (3) the EEC in water is equal to or greater than 0.01 of any acute EC50 or LC50 value. In addition, testing with other organisms indicate the reproductive physiology of invertebrates may be affected. The preferred test species are *Daphnia magna* for freshwater and *Mysidopsis bahia* for estuarine marine scenarios. Results of these tests are tabulated below.

**Table 25. Freshwater and Marine/Estuarine Aquatic Invertebrate Life-Cycle Toxicity**

Species	%ai	LOEC (parameter)	NOEC	MRID	Author	Classification
Water flea(FW)	94	21D LOEC= 0.10 PPB	0.006 PPB	41718401	Blakemore,G and D.Burgess, 1990	Core
Mysid(SW)		No Data				Required

The guideline (72-4) is fulfilled for freshwater species.

The guideline (72-4) is **not** fulfilled for estuarine species. A full life cycle study for mysid or other acceptable marine/estuarine species is required for the proposed uses of malathion.

## **Non Guideline Laboratory Studies with Aquatic Organisms**

### **Acetylcholinesterase Inhibition in Federally Endangered Colorado Squawfish exposed to Carbaryl and Malathion Beyers, P. & P.Sikoski, 1993.**

Methods: Squawfish were exposed for 32 days in flowthrough systems for the study of early life stage effects to this listed species. AChE inhibition was based on 24 hour exposure.

Results: AChE Activity NOEC=371 ug/L and LOEC=707 ug/l for malathion.

### **Acetylcholinesterase Inhibition in Federally Endangered Bonytail Chub Exposed to Carbaryl and Malathion. Beyers, P. (1993)**

Methods: 32 day static renewal exposure and 4 day static renewal acute exposures employed.

Results: The NOEC for acetocholinesterase inhibition and reduced growth in the Bonytail chub was 990 ug/L. Threshold conc. = 521 ppb

For malathion the squawfish estimated  $LC_{50} = 9.14$  ppm (8.3 - 10) nominal concentration. The Bonytail chub  $LC_{50} = 15.3$  ppm(14.4 - 16.4) based on nominal concentrations. **Reviewers note:** The LC50 levels may have been lower if measured concentrations had been made and used in LC50 calculations. The second Beyers test also mentions an LOEC of 24.7 PPB for flagfish, *Jordanella floridae* in a previous 30 day exposure for survival/growth.

### **Chronic Toxicity of Malathion to the Bluegill (*Lepomis macrochirus*). Eaton, John G., 1970. Duluth EPA Lab., Duluth Minnesota.**

Methods: Fish were wild caught from local ponds. The test fish were exposed at varying concentrations for over 8 weeks. Triton surfactant was added as solvent for stock solutions.

Results: Spinal deformation was observed at <10 ppb. MATC >3.6 <7.4 ppb All fish died at 80 and 40 ppb (within 16 days at 80 ppb and within 54 days at 40 ppb. Reproductive NOEC was <20 ppb. Survival and numbers of eggs produced/female were effected.

### **Sublethal Effects of Malathion to Three Salmonid Species. Post, George and Robert Leasure. Colorado State University, 1974.**

Methods: Test material was 55% EC. Brook trout, rainbow trout and coho salmon were subjected to stamina flow tunnel tests after exposure to 40 - 300 ppb concentrations for 7-10 days. This type of test was used to imitate upstream migration .

Results: Coho salmon was the most sensitive species with AChE levels reduced 75%. The exposed fish were unable to perform 2/3 of work activity (swimming in current) as the unexposed fish. Possible field effect=Less able to hunt, spawn or migrate upstream.

### **Toxicity of Malathion 500 to fall Chinook Salmon Fingerlings. Parkhurst, Zell and Harlan Johnson. USFWS, 1955 (Progressive Fish Culturist)**



Methods: Fingerlings were exposed for 96 hours to concentrations of up to 240 ppb.

Results:  $LC_{50}$  = 120 ppb after 96 hrs and 170 ppb after only 24 hours. 95% mortality after 24 hours at 240 ppb.

**The Toxicity of the Hydrolysis and Breakdown Products of Malathion to Fathead minnow - Bender, Michael E., 1969. University of Michigan.**

Methods: 96 hour static and 14 day flowthrough exposures

Results: Results demonstrated that diethyl fumarate was more toxic than the parent to this species and that synergistic effects occur between the parent and the two major degradates. Toxicity values for 4 confirmed and 9 proposed degradates to fathead minnow are provided. Diethyl fumarate  $LC_{50}$  = 4.5 mg/L. The  $LC_{50}$  for malathion to fathead minnow from Mayer and Ellersieck's publication is 8.65 mg/L.

**Table 26. Toxicity of confirmed and proposed malathion degradates to fathead minnow.**

<u>Degradate</u>	<u>96 hour TLM in mg/L</u>
Dimethylphosphorodithioic acid	23.5
Diethyl fumarate	4.5
2-mercaptodiethyl succinate	35.0
Dimethylphosphorothionic acid	42.5
Maleic acid	5.0
Diethyl maleate	18.0
Dimethyl phosphate	18.0
Dimethyl phosphite	225.0
Thioglycolic acid	30.0
Diethyl succinate	140.0
Diethyl dl-tartarate	650.0
Bis(hydroxymethyl) phosphinic acid	29.0
Ethylene phosphite	34.0

**Uptake and Retention of Malathion by the Carp. Bender, Michael E. University of Michigan, 1969.**

Methods: *Cyprinus carpio* used as test species. Fish were exposed for 4 days at 2.5 ppm.

Results: Liver, flesh, blood, gills and brain were areas of highest concentration (in that order).

**The Toxicity of Malathion and its Hydrolysis Products to Eastern Mudminnow *Umbra pygmaea* Bender, Michael E., Virginia Institute of Marine Science, 1976.**

Methods: Fish were exposed for 96 hour using static systems and 14 days using flow through systems. The fish were wildcaught. The laboratory used dechlorinated tapwater in 5 gallon tanks with 10 fish exposed/concentration. Five concentrations were tested.

Results: 2 mecapto diethyl succinate shown to be most toxic of 4 degradates tested to this species. ( $LC_{50}$  = 0.32 ppm). Diethyl fumarate  $LC_{50}$  = 1.47 ppm (was least toxic).

**Abnormal Locomotion Associated with Skeletal Malformations in Sheepshead minnow. Weis, Peddrick and Judith Weiss, Rutgers University and New York Ocean Science Laboratory, Montauk, New York, 1975.**

Methods: Embryos were exposed at 3 and 10 ppm with 25 eggs/concentration.

Results: Exposure appeared to produce skeletal malformations which impeded swimming ability of fry. NOEC was 1 ppm. 25% and 41% of the larvae were effected at 3 and 10 ppm, respectively. Delayed hatch was observed in the 10 ppm test group.

**Woodward, Dan F., Sport Fisheries Research USFWS publication 77, 1969.**

Results: The study reports an observed loss of avoidance response in goldfish after 72 hr exposure to malathion at 1 and 5 ppm.

**Quarterly Report - USFWS Research Laboratory, Columbia, Mo. 1967.**

Results: Acute Toxicity Studies

- 1.3 gm Walleye  $LC_{50} = 62$  ppb Raw water
- 0.9 gm Largemouth bass  $LC_{50} = 80$  ppb raw water
- 1.4 gm - Bluegill  $LC_{50} = 110$  ppb raw water
- Bluegill  $LC_{50} = 130$  ppb in pond water
- Gammarus faciatius*  $LC_{50} = 0.8$  ppb

Chronic Exposure: 1/4 acre ponds containing bluegill and catfish and were used to measure chronic effects when applications reached 0.002 and 0.020 ppm. No clear indication of significant growth or hematocrit count differences were seen.

**USFWS Sport Fisheries Research Report Publication 106, 1970 Wash. DC.**

Reported Results of Acute Testing: Rainbow trout  $LC_{50} = 93.5$  ppb at 96 hours. Korean shrimp *Palaemon macrodactylus*  $LC_{50} = 33.7$  ppb (21.3 - 53). Striped bass at 30 ppt. salinity,  $LC_{50} > 1000$  ppb.

**Effects of Insecticides on Feeding Activity of the Guppy, a Mosquito-eating Fish in Thailand - Rong, Suriyam, Y. et al, 1968. World Health Organization.**

Results: 24 hour  $LC_{50}$  mosquito larvae = 50 ppb  
24 hour  $LC_{50}$  Guppy = 50 ppb

**Effects of Field Applied Rates of Five Organophosphorous Insecticides on Thermal Tolerance, Orientation and Survival of *Gambusia affinis*. Johnson, C.R. 1977.**

Results: Ability to survive thermal change  $EC_{50} < 100$  ppb. Mortality of 100% at 500 ppb malathion.

**Effects of Mirex, Methoxychlor and Malathion on Development of Crabs. Bookhout, Cazlyn G. and John D. Costlow Jr., 1976, Duke University and Gulfbreeze Laboratory (USEPA) Pg. 53-69.**

Methods: The study was designed to investigate effects to larvae development of crabs. Mudcrabs, *Rhithropanopeus horrisii* and Blue crabs, *Callinectes sapidus* were studied.

Results-Malathion Exposure: Mudcrab larval survival was significantly reduced (100% to 12%) at 11 to 20 ppb, respectively. They did not survive past 2nd local stage at 50 ppb. Development time was also delayed between stages. Blue crab larvae development was slightly delayed but significant reduction in survival to megalopa stage was seen at concentrations of 50 ppb and less significantly reduced at 20 ppb. Total mortality in all stages was high in concentrations of 20 ppb or above compared to acetone controls.

**Toxicity of Malathion to Mammals, Aquatic Organisms and Tissue Culture Cells. Desi, I. et al, Division of Hygienic Biology, Budapest, Hungary, 1976**

Methods: AChE was measured in rat brains. Adductor muscle activity was measured in the freshwater mussel *Anodonta cygnea*. Guppies were exposed to 100, 1000, 10,000, 1000 and 100,000 ppb of Malathion. 30 Daphnia were exposed to 100,000, 10,000, 1000, 100, 10 and 1 ppb, in 300 ml of water (test repeated 3 times). Freshwater mussel larvae (*Glochidium*) shell closing activity was measured for 30 larvae over 3 min period after exposure to concentrations ranging between 100 and 100,000 ppb of malathion.

Results: Rats displayed inhibited levels of AChE and abnormal EEG's 90 days post exposure at concentrations of 75 and 38 mg/kg of body wt. *Anodonta cygnea* showed significantly reduced activity during 48 hour exposure at 10,000 ppb. No change noted at 1000 ppb or less. Guppies were killed at 1000 ppb or above. The LC<sub>50</sub> for guppies was calculated to be 819 ppb. For Daphnia 100% mortality was observed at 10 ppb or above. No effects were observed at 1 ppb. *Glochidium* were less active at 1 ppb but showed similar activity as the controls at 0.1 ppb (NOEL).

**Malathion, Chronic Effects on Estuarine Fish - Holland, H.T. and Jack Lowe, Gulfbreeze Biological Laboratory, 1966.**

Methods: Atlantic Spot, *Leiostomus xanthurus* were exposed to constant concentration of 10 ppb of Malathion for 26 weeks.

Results: Though brain AChE levels in treated fish were 70% of that in untreated fish, no other adverse effects on the spot were noted. After 1 week AChE levels returned to normal.

**Microbial-Malathion Interaction in Artificial Saltmarsh Ecosystems. Bourquin, Al W. Gulfbreeze Laboratory, USEPA. 1975.**

Methods: Natural bacteria samples from uncontaminated marsh were added along with 10 ml of sea water and 10 gm of sediment to 250 ml flasks. 10 mg. aliquots of malathion in acetone were

added every 7 days. Cultures were analyzed for malathion levels and compared to control vial residue levels.

Results: Increased salinity sped up the degradation process. Malaoxon levels remained constant. Monocarboxylic acid and dicarboxylic acid levels increased. Conclusion was that chemical and microbiological processes will act to degrade the levels of parent malathion in saltmarsh environments.

**A Method for Establishing Acceptable Toxicant Limits for Fish - Malathion and Butoxyethanol Ester of 2, 4-D - Mount, Donald I. and Charles Stephan, 1967 U.S. Dept of Interior.**

Methods: Fathead minnows exposed under flowthrough conditions for 9 months. Fish were 1" long fry. Four malathion test concentrations and a control were employed. Ten fish were initially tested per tank with 2 tanks per concentration. Later this was reduced to 5 fish per tank to better stimulate spawning and decrease mating competition.

Results: Proposal was to divide the NOEC by the  $LC_{50}$  and use this fraction as a multiplication factor with other fish species  $LC_{50}$  levels to obtain acceptable LOC levels for other species groups. In this case of fathead minnow exposed to malathion the fraction was 1/45. 20% of the fish exposed to 580 ppb died after 7 weeks. However, survivors at this concentration were able to spawn and reproduce.

**Determination of Malathion, Malaoxon, and Mono-and Dicarboxylic Acids of Malathion in Fish, Oyster, and Shrimp Tissue. Cook, Gary H. and James C. Moore, 1976. USEPA Gulfbreeze Laboratory.**

Methods: Pinfish were exposed to 75 ppb for 24 hours

Results: Greatest tissue concentrations were the MCA and DCA metabolites. Malaoxon and parent malathion were not detected.

**Toxicity of Malathion to Native Freshwater Mussels. Keller, Anne E. , 1995. National Biological Survey Laboratory, Gainesville, Florida.**

Methods: A two year study was conducted with several species of endangered or threatened mussels in various lifestages (glochidial, juvenile, and adult). Malathion was tested at concentrations as high as 500,000 ppb.

Results: Adults were not significantly effected at up to 350,000 ppb.  $LC_{50}$  values for glochidia were determined to range from 133,000 to 494,000 ppb. Juvenile  $LC_{50}$  values ranged from 36,000 to 523,000 ppb.

**Observations of Effects to Aquatic Organisms From Field Applications**

Over its history Malathion has been used extensively over or near freshwater habitats and marshes for mosquito and medfly control. A number of aquatic field studies have been independently conducted and are still being conducted for this type of use pattern. A summarization of the studies contained in Agency files is provided below with a brief discussion of the reported results.

**Dibrom/Malathion Formulation Use as a Piscicide (Hoff, James and Westman, James, 1965)**

Methods: Various combinations tested to determine kill ratio for fish eradication efforts

Results: 3pt/2pt ratio of dibrom and malathion was found to be an effective combination for use as a piscicide.

**Brain Acetylcholinesterase Inhibition in fish as a Diagnosis of Environmental Poisoning by Malathion (Coppage, D et al 1975) Gulfbreeze Environmental Research Lab.**

Methods: Pinfish were wildcaught and acclimated for 3 weeks. A flowthrough exposure was conducted for 72 hours at up to 500 ppb of malathion dissolved in acetone.

Conclusions: Malaoxon is the active AChE Inhibitor they believe since the parent was not present 2 wks later. Inhibition of over 70% generally indicates impending death. This occurred at about 58 ppb. At 25 ppb AChE inhibition of about 34% observed, but with eventual recovery of the test fish.

**Malathion Toxicity to Killifish in Delaware. Darsie, Richard and Coraiden, F. Eugene, 1958. Delaware Agricultural Research Station.**

Methods: This study involved spraying of Delaware marsh near Odessa, Delaware. 381 fish (*Fundulus ocellaris*) were wildcaught and used in testing procedures. 25 fish were placed in each of several metal tubs containing 7 gallons of natural marshwater. An aerial spray was applied at 0.51 lb ai/acre of malathion mixed with 2 qts of diesel oil. Monitored rate was 167 droplets/inch<sup>2</sup>.

Results: After 4 hours 26.3% of the fish died, 42.4% showed sublethal effects and 31.2% appeared unaffected. 26% of the sublethally effected fish died later even though placed in clean water while 74% recovered.

The study report also mentions field observation of extensive mortality to species of killifish during medfly spraying in Florida in 1958. (Applic.Rate was 0.2 - 0.75 lbs ai/A)

**Effects of Malathion on Two Warmwater Fishes and Aquatic Invertebrates in Ponds. Kennedy, Harry D. and David Walsh. USFWS, Fish Pesticide Research Laboratory, Columbia, Mo., 1970.**

Methods: Bluegills and Channel catfish were exposed. 4 applications were made at concentrations of 0.02 and 0.002 ppm over an 11 week summer period. Twelve ponds were treated. Pond surface areas were 688 m<sup>2</sup> with average depth of 0.76 m. and volume of 602 m<sup>3</sup>.

Results: The 8-44% fish loss was not felt to be treatment related as controls also had similar losses. Treatment effects appeared to be reductions of aquatic insects particularly midges at high and low doses (0.02 ppm and 0.002 ppm). Mayflies were reduced also with a significant reduction occurring after the 3rd application.

**Effect of Aerially Applied Malathion on Juvenile Brown and White Shrimp, *Penaeus aztecus* and *Penaeus setiferus*. Conte, Fred S. and Jack C. Parker - Texas A&M University 1975 (Am. Fisheries Society)**

Methods: 3 Bayous and an estuarine lake were monitored. Mean water depth was 61 cm. Wild caught shrimp placed in cages were aerially sprayed at a rate of 85.7 g/hectare by aircraft at a speed of 145 km/hr. 7 to 3 passes were made at each site.

Results: In Test I within 9 hours after treatment 73% of all mortality occurred (24 of 50 shrimp died). Test II produced 50% mortality in 49 hours after application. Only 12% mortality occurred in Test III (estuarine lake). Water concentrations ranged from 2.0 to 3.2 ppb immediately after application.

**Effects of Ground Application of Malathion on Saltmarsh Environments In Northwestern Florida. Tagatz, M.E., 1974. USEPA Gulfbreeze, Environmental Research Laboratory, Gulfbreeze, Florida.**

Methods: Thermal fog and ULV application in Northwestern Florida were monitored. Malathion was applied during low tide with 2 week intervals between applications. Thermal fog was applied at 6 oz/Acre (Sept. & Oct 1972) to a saltmarsh pond with fuel oil carrier. Blue crabs, juvenile sheepshead minnow and adult grass shrimp were exposed.

Results-Thermal Fog: 1 application produced high mortality of shrimp after 7 days. Some reduced AChE levels were observed in fish. No mortality of fish or crabs occurred.

Methods: ULV application was made at 0.64 fl oz/Acre. Three applications were made by truck mounted aerosol generator, with a 330 foot swath. Grass shrimp, blue crabs, and sheepshead minnow were exposed in 18" diameter polyethylene tubs.

Results: No effects noted to animals. No treatment related mortality was observed. Residue levels were 0.28 - 0.34 ppb after the 3rd application.

**Effects of Pesticides in Estuaries Along the Gulf and Southeast Atlantic Coasts.**

**Coppage, D.L. and T. W. Duke, 1971, USEPA, Gulfbreeze Laboratory**

Methods: AChE inhibition measurements were made in spot, croaker and mullet in Louisiana after spraying malathion for mosquito control. Collected fish were frozen and shipped on ice to Gulfbreeze. Normal levels were measured and reported to range from 1.08 to 1.45 before spraying commenced.

Results: After spraying AChE levels were reduced to as low as 0.09. Range of inhibition was measured at 97% to 11% inhibition. Inhibition in spot ranged from 97% to 11% inhibition. Inhibition in spot lasted over 1 week (still 36% inhibition). Second spray was made 18 days after first and mullet were killed while spot and croaker suffered further reduction in AChE. Inhibition may remain over one month after spraying.

**Mortality of Post larval and Juvenile Shrimp Caused by Exposure to Malathion -A Laboratory and Field Study. Proctor, Raphael R. Jr., Jane P. Corliss, and Donald Lightner, National Marine Fisheries Service, Galveston Laboratory, 1966.**

Laboratory Methods: Postlarval white shrimp and brown shrimp were exposed for 48 hrs. in laboratory tanks.

Results: Calculated 50% lethality levels for adults were 25.5 to 21.3 ppb for post larval brown shrimp and 100% mortality of larvae was seen at concentrations as low as 18 ppb.

Field Methods: Caged shrimp were exposed in estuarine areas to application of malathion(95% ai) at 77.8 ml/acre. Water depth was about 1.2 meters (high-tide) for the first application and 0.3 meters at the time of the second application (mean tide).

Field Monitoring Results: Environmental concentrations reached 8.9 ppb at high tide and 69 ppb at mean tide level. Some contamination of control areas occurred possibly from drift. 14% mortality was observed in controls and 80% mortality was seen in the test marsh. In the second application 65-69 ppb residue levels were seen 6 hours after treatment. Mortality was 48% in treated area and 4% in control area. After 10 hours white shrimp mortality increased to 96% in treated area and 7% in control area at mid depth levels. By 24 hours the residue levels had decreased to 1.08 ppb. White shrimp caged on the bottom level showed a similar trend after second application. Brown shrimp mortality results were inconclusive as treated areas showed 55% mortality while controls showed 44% mortality.

**Impact on Fish and Wildlife From Broadscale Aerial Malathion Applications in South San Francisco Bay Region, 1981. Finlayson, B.J., G. Faggella, H. Jong, E. Littrell, and T.Lew, Pesticide Investigation Unit, Water Pollution Control Laboratory, California Fish and Game Department.**

Methods: This 120 page report summarizes extensive monitoring performed during 1981 Medfly control programs around the San Francisco Bay region. In general, most of the 200 fish and invertebrate tissue samples taken contained no detectable levels of malathion residues (<0.5 ppm). This was not true in the case of samples taken at fish kill sites. Steelhead trout populations were monitored in the San Lorenzo drainage area. Aquatic insect populations in the drainage were also monitored (number per sq. Cm).

Results: No discernable effects were noted for steelhead trout populations or appearance or size measurements when compared to control sites. There were significant reductions in either diversity or population counts for aquatic insects (33-50% reduction). Eight fish kills were associated with malathion spraying efforts, while 15 were either not determined as to cause or not attributed to malathion (see incident report section of this document). Many of the fish losses were sticklebacks (highly sensitive to malathion) while carp and channel catfish appeared unaffected at the same locations.

**Impact of Malathion on Fish and Aquatic Invertebrate Communities and on Acetylcholinesterase Activity in Fishes in Stewart Creek, Fayette County, Alabama. Kuhajda, B.R. et al, Dept. Of Biological Sciences, University of Alabama, 1996.**

Methods: Creek is located in west-central Alabama near Winfield and has an approximately 11 square mile drainage basin. Samples were taken upstream, at the entry point, and 0.5 mi. downstream from the application site on two small cotton fields ( 7.6 and 11.6 acres). Fields were within 25 feet of the stream bank. There were no trees along the banks, only grasses and kudzu vines. Sample sites were sampled for three years-the first two during malathion applications, the last during which malathion was not applied. Captured fish were identified, counted , and some analyzed for AChE inhibition. Invertebrates were captured (by kicking up

sediments into a dipnet), recorded, and then preserved in ethanol. 39 samples from each location were taken over a 34 month period. Only one sample date represented prespray conditions. Results: Concentrations recorded ranged from ND to 10.89 ppb (mean=3.49 ppb) for the nine 1993 applications and from 0.88 to 31.1 ppb (mean=2.08) during the four 1994 applications.. 11,921 fish of 48 different species were collected during the study. Numbers and diversity of collected fish did not appear to significantly vary. Not all species were equally distributed at the three sites and some population differences may be attributable to the differences in habitat preferences and availability at the three sites. Numerous specimens of rough shiner, *Notropis baileyi* were collected and analyzed for AChE and significant depression was noted during the spray periods when compared to the upstream control site. Of interest is the observation that downstream activity levels were lower than those at the application site.

Aquatic invertebrate populations which were collected included 87 taxa, and a total of 6,088 individual organisms. Some difference is apparent in numbers and diversity of species collected near the spray site when compared to the upstream site, but significant differences were less apparent at the downstream location. The upstream location did have more taxa present, however, than either of the other two sites for all periods of this study. The study author was not certain that this could be attributable to malathion influence as natural variability could also have played some part.

### Freshwater, Estuarine and Marine Aquatic Incidents:

The Agency receives and reviews all wildlife incidents where aquatic organism kills occurred following application of a pesticide or mixture of pesticides. These incidents are sometimes reported under 6a2 provisions of FIFRA while others are independently submitted by local, state, or federal agencies. Those which are directly implicated with Malathion use are summarized below along with factors which are known about events preceding the incident.

**Table 27. Freshwater, Estuarine and Marine Aquatic Incidents.**

Location and Date	Incident #	Description	Probability
Florida Medfly report-1997 Spray operations Hillsborough Area USDA Report	6 reports from 7/29- 8/28	40 Sites of Fish kills investigated-malathion detected in varying amounts in ponds and pools. Fish species effected include-various sunfish, bass, perch, and carp. 3 tropical fish farms hit. Mortality ranged from 5 to 1000 fish per site. Aerial drift generally blamed though some runoff events did occur.	Probable-residues detected in water and sometimes tissues
South Dakota, Minihaha Co., 7/6/87	I000804-025	10,000 dead fish-incl. walleye and yellow perch-aerial-Clean Crop -near Lake Madison	Probable
North Carolina, Wake Co., 5/17/73	B0000-225	10,000 panfish killed from ½ gallon spill of formulation(12.2 % Malathion/12.2% endosulfan into a pond.	Highly probable

88



Mississippi, Silver Creek, 7/6/89	I000389-001	166 fish, mostly carp, were killed-pest control company applied Aqua Malathion 8 in area	Possible
Missouri, 5/5/70	I000636-002	33 fish kill reports-one sick dog from ingestion of contaminated water	Possible
South Dakota, 7/3/87	I000804-025	35 other incidents besides Lake Madison fish kill-birds, fish, bees effected	Possible
Alabama	I002059-002	2 fish kills-Cotton field application of malathion-bass and sunfish killed	Possible
Florida	I000524-008	Turtle and birds mortalities reported	Possible
New Jersey, Delaware River 8/9/91	none-	Malathion distributed in sewage effluent to kill flies-15 gal malathion product /13000 gal effluent-1000 to 5000 white perch killed at discharge point	Probable
Maryland, Cherry Hill- 5/12/80	EPA report	350 fish found dead- 10,000 acre lake- municipal pest control - Malathion	Probable
Missouri, Wentzville.- 6/29/80	EPA report	6,790 dead fish counted-Malathion treated municipal sewage discharge to McCoy Creek	Probable
South Carolina, Hilton Head-5/25/81	EPA report	1500 dead fish-Sea Pine Lagoons-estuary-pesticide spraying operations using Malathion	Probable
Virginia, Norfolk-8/14/81	EPA report	1500 dead fish-Mason Creek-industrial operations using Malathion	Possible
Florida, near Miami - Summer, 1956	Old report from Mr. J.E. McCurdy-Florida Mosquito Control?	Extensive observations of numerous canals, ponds, ditches, and pools after aerial application of Malathion-some species killed others not-mortality to thousands of mojarra silversides was immediate after spraying-snook, mollies, cyprinids, pinfish , bass and killifish also killed in ditch and canal areas-strangely gambusia were not sensitive	Probable
Massachusetts-four incidents White Island Pond near Plymouth Glen Charles Pond near Waneham Waneham River-estuary Agawan River-estuary	6a2 Report from American Cyanamid Oct. 4, 1990 #281720	4 fish kills reported from treatment of 700,000 acres of estuarine areas with Malathion for control of mosquitoes. Many of the dead fish were estuarine killifish species.	Probable
New York, Thornwood- 5/14/84	EPA report	500 dead fish-Pond in Carroll park-agriculture operations using malathion adjacent to pond	Possible

<p>California-Monitored aquatic incidents during broadscale aerial application over San Francisco, Bay area, 1981. Administrative Report 82-2, Dept. Of Fish and Game, Environmental Services Branch, 1982.</p>	<p>Medfly Control</p>	<p>23 fish poisoning incidents were investigated-8 were confirmed as caused by malathion -10 were listed as undetermined causes-2 were caused by chlorine discharge at sewer plants. Malathion incidents included observed mortality of over 2300 fish including stickleback(Stevens, San Tomas Aquino, Pescadero Creeks), carp(Adobe and Mission Creeks), mosquitofish(Mission Creek), topmelt, flounder, striped bass, and gobies(Seal and Redwood Creek, and Mayfield Sloughs), and largemouth bass and crappie in San Jose Pond.</p>	<p>Probable- Malathion residues detected in water-tissue concentrations in gill filaments, liver, skeletal muscle and whole body tissues</p>
<p>Alabama, Tennessee 1995 Southeast Bollweevil Eradication Program, Environmental Monitoring Report</p>	<p>USDA /APHIS 1995 report</p>	<p>Leighton, Alabama-Catfish Farm-dead catfish-600 ft from aerially treated field(#295) Lincoln Co., TN.-2 acre stream fed pond-4 cotton fields upstream-dead bass, catfish, sunfish.</p> <p>Lighten, AL.-Big Nance Creek-30,000-40,000 fish Colbert Co.,AL.-Donnegan's Slough-fish kill- both followed heavy rains 8/4-8/8 resulting from hurricane</p> <p>Fish pond near Site 139-dead sunfish, catfish-malathion residues in water-5 to 6 ppb</p> <p>Catfish Farm-2 ponds-dead catfish near field #19-150 feet from ponds-9 old day samples did not show high concentration levels-only trace levels</p> <p>Fishkill-1/10 acre pond near field #303-dead adult catfish sampled-malathion detected in water.</p> <p>Fish, turtle, frog, and crayfish kill-5 acre wetland-2 to 3 ft. Depth-cotton field 503 located 600 ft. away-drainage ditch leads to wetland-6 day old samples-malathion still detected in water and fish tissues.</p> <p>Fish Kill(bass, sunfish, catfish)-8 acre pond-20 ft. From application site(cotton field # 1180)-residues of 77.8 ppb in one water sample. Other chemicals used in area-Larvin and Pyrat</p> <p>Fishkill -1/4 acre farm pond near cotton fields #118 and 119-malathion residues in all 4 water samples-fish tissue sample contained 351ppb malathion.</p> <p>Fishkill(catfish)-1/4 acre pond near field#166-70 ft from pond-malathion detected in 8 day post-application samples-</p>	<p>Probable- inspection was too late in many cases-1 week after</p> <p>Possible-Endosulfan, malathion and methyl parathion all suspect. Probable-</p> <p>Probable</p> <p>Possible</p> <p>Probable-fish tissue residues 35-85 ppb.</p> <p>Probable-though not likely from bollweevil aerial treatment, 6 weeks previous</p> <p>Probable-residue levels in tissues were high</p> <p>Possible-sampling too late-cotton field treated 8 days earlier</p>
<p>California-4 Incidents near Fremont, Loma Mar, San Jose, and San Mateo Co. 9/30/81-10/9/81</p>	<p>EPA report</p>	<p>2000 dead fish-Fremont Creek-crop treatment 200 dead fish-Pescadero Creek-crop treatment 75 dead fish-pond near San Jose-crop treated 12 dead fish-Adobe creek-crop treatment</p>	<p>Possible</p>

### **Significance of Reported Incidents**

Though malathion has been used for many years, the greatest numbers of detailed reports of fish kill incidents involved heavily monitored programs such as USDA's boll weevil eradication program and the mediterranean fruit fly eradication efforts. Other incidents appeared linked to uses near aquatic habitats where direct drift may have occurred, such as mosquito control. In many of the incidents use rates and residue levels following the incidents are not clear and kills are investigated days after the event probably occurred. In two of the incidents sewage discharge was treated with malathion to control flies and then released directly into tributaries. In all cases where residue levels are provided they are within the limits expected to prove toxic to sensitive fish species (>4 ppb). One of the points that should be included when discussing fish kill incidents is that invertebrates are likely to have been more severely effected since fish are less sensitive to malathion than a majority of the invertebrate species tested in laboratories to date. In most of the fish kill incidents there appears to have been no effort to investigate the effects to the other components of the ecological community in the adversely effected sites.

### **Toxicity to Plants**

To date the Agency has received no data from malathion registrants regarding the toxicity of malathion to non-target plants. This is not normally a requirement for insecticidal use pesticides. However, the direct application of malathion to aquatic habitats does raise concerns regarding possible phytotoxicity of the product impurities or inert ingredients to non-target aquatic plants or semiaquatic plants. Based on the following study and also results observed in field studies (see previous study, MRID 00104629-Giles, 1970) malathion is expected to be taken up and stored for some time in plant tissues. Metabolites may later show up in new stem and leaf growth.

#### **Comparative Metabolism of Malathion - C<sub>14</sub> in Plants and Animals. Bourke, J.B. et al-New York State Agricultural Experiment Station, Cornell University, 1968.**

Methods: Red Kidney bean plants were forced to imbibe radio labeled malathion solution by passing air over foliage for 20-30 minutes (with solution mixed into air stream?) C<sup>14</sup> was traced in tissues of plants for 14 days. Various intermediates (metabolites) were deposited within tissues.

Results: Plants appeared to store various metabolites in tissues..

### **Toxicity of Degradates and Impurities**

Malathion may contain impurities which account for up to 5% of the pesticide content. These impurities include diethyl fumarate, diethylhydroxysuccinate, O,O-dimethylphosphorothioate, O,O,O-trimethyl phosphorothioate, O,O,S-trimethyl phosphorodithioate, ethyl nitrite, diethyl-bis (ethoxycarbonyl)mercaptosuccinate, S-1, 2-ethyl-O,S-dimethyl phosphorodithioate (isomalathion), S-(1-methoxycarbonyl-2-ethoxycarbonyl)ethyl-O, O-dimethyl phosphorodithioate, Bis-(O,O-dimethyl thionophosphoryl) sulfide, Diethyl methylthiosuccinate, S-ethyl-O,O-dimethyl phosphorodithioate, S-1,2-bis(ethoxycarbonyl)ethyl-O,O,-dimethyl

phosphorothioate (malaaxon), diethyl ethylthiosuccinate, and sulfuric acid. These impurities may range from 0.5% to 1.0 % of the content and have been shown to be toxic alone and may even potentiate the toxicity of the parent. Pellegrini and Santi, 1972, found that purified malathion ( 98% ai) was actually less toxic to laboratory rats than technical malathion of 92.2% purity with corresponding LD50 levels of 1580 mg ai/Kg versus 8000 mg ai/Kg, respectively.

Several studies regarding the toxicity and retention of degradates in fish were reviewed from literature. In 1976 studies at the EPA Gulfbreeze Laboratory, Cook and Moore found that the monocarboxylic and dicarboxylic acids of malathion were detected in fish tissues after 24 hours, but malaaxon and malathion were not. Studies by Dr. Michael Bender at the University of Michigan (1969) and Virginia Institute of Marine Sciences (1976) showed that [diethyl fumarate] and 2 mercapto diethyl succinate were more toxic than the parent compound to fathead minnow and eastern mudminnows (see pages 71-72). However the percentage of these degradates in the environment is expected to be low enough (<10% of original parent) to prevent additional toxicity to fish. Unfortunately, the testing reported for degradate toxicity was not performed on fish species considered highly sensitive to malathion (fathead minnow and eastern mudminnow). No degradate toxicity to invertebrate species has been reviewed. Toxicity from accumulation of degradates following multiple applications is unclear without further fate and chemistry data to characterize their potential to degrade in the environment.

## **Toxicity of Dual Active Mixtures**

### **Mixture Toxicity to Terrestrial Wildlife**

Malathion and Methoxychlor mixtures are marketed by Cheminova and Platte Chemical Company. There is some data regarding the possibility of increased toxicity of combinations of pesticides to rats and mice (M.L. Keplinger and Deichmann, 1967). In their paper entitled Acute Toxicity of Combinations of Pesticides (Toxicology and Applied Pharmacology 10, 586-595(1967), Keplinger and Deichman tested numerous mixtures of pesticides commonly used at the time. Rats and mice were orally intubated with pesticides mixed in corn oil. Generally 5-7 dosages were administered to five animals at each level, with separate chemicals administered within 10 seconds of each other. When methoxychlor was mixed with malathion there was a slight additive effect to expected toxicity, based on the author's computation, which assumed that the expected LD50 of the combined chemicals would be equivalent to the midpoint between the known oral toxicities of the two compounds alone. The expected LD50 of methoxychlor/malathion 50-50 mix was estimated to be 1850 mg/Kg for mice whereas the actual observed LD50 was 1620 mg/Kg. This may be an inadequate difference on which to base any gross assumptions of synergistic effect for these two chemicals in combination. However, it should be noted that certain other combinations of malathion did show additive effects (toxaphene and carbaryl) whereas a protective effect was noted with certain other combinations (aldrin, dieldrin, chlordane, and endrin) when the same assumptions for predicted LD50 levels were made.

Malathion is formulated with several other active ingredients which also may display some levels of toxicity to birds. Among these is Fertilome A-C-G Insecticide and Fungicide Mix marketed

by Voluntary Purchasing Group Inc., Blackleaf Liquid Fruit Tree Spray with Fungicide (Sureco Inc.). These formulations may have additional toxicity over single active formulations containing only malathion and should be tested separately. At this time the Agency has no data on which to predict potential effects to birds from aggregate exposure to these multi-active formulations. Comparison of methoxychlor and malathion avian toxicity values (see table below) indicates that methoxychlor displays low acute values similar to malathion. The mixture of these compounds may provide some additional exposure time due the increased persistence of methoxychlor over malathion. The reviewer was unable to locate data to indicate that the mixture will or will not be more toxic due to synergistic effects of the two insecticides. The fungicide/insecticide mixtures may also add additional toxicity to avian species. This is based on slightly elevated plasma butyryl cholinesterase (BChE) levels in quail when malathion was administered in combination with vinclozolin and ketoconazole and elevated BChE levels in rats when malathion was administered in combination with propiconazole, vinclozolin, and clomitrizazole (Martin, J.J.R. and Thomas Badger, University of Arkansas, Toxicology and Pharmacology 130, 221-228,1995). In studies with Japanese quail, red-legged partridge, and pigeons pretreatment with the fungicide prochloraz resulted in enhanced toxicity of malathion (Riviere J.L. et al, Arch. Environmental Toxicology 14, 1985 and Johnston, G. et al., 1989 Pesticide Biochem. Physiology 35, 107-118)

**Table 28. Comparative Toxicity of Malathion and Methoxychlor to birds.**

Species	% ai	Malathion	% ai	Methoxychlor
Mallard	95%	LD50=1485 mg/Kg	Tech	LD50=2000 mg/Kg
Ring-necked Pheasant	95%	LC50=2639 ppm	Tech	LC50>5000 ppm
Bobwhite	95%	LC50=3497 ppm	Tech	LC50>5000 ppm

Mixtures containing malathion and methoxychlor may produce similar or more pronounced chronic effects if additional persistence results from addition of the organochlorine insecticide.

**Mixture Toxicity to Aquatic Organisms**

Studies published by the Bureau of Sport Fisheries and Wildlife(later USFWS) in the June 1970 issue of Progress in Sport Fishery Research 1969, explored the synergistic activity of combinations of pesticides on toxicity levels for bluegill and rainbow trout. Synergism was observed when malathion was mixed with baytex, EPN, Parathion, Perthane, and Carbaryl. Additive effect was noted when combined with DDT and Toxaphene.

Review of toxicity data for methoxychlor indicates that this chemical may provide additional toxicity over that of malathion to most species of aquatic organisms. A brief, but not comprehensive, comparison table is presented below for comparison of acute toxicity values for the two insecticides to fish and invertebrates. The Agency has not received data to support the registration of methoxychlor/malathion mixtures.

93

**Table 29. Malathion / methoxychlor comparative toxicities to aquatic organisms**

Species	%ai	Malathion	%ai	Methoxychlor
Waterflea, <i>Daphnia pulex</i>	95	EC50=1.8 ppb	98	EC50=0.78 ppb
Scud, <i>Gammarus fasciatus</i>	95	EC50=0.5 ppb	98	EC50=1.9 ppb
Scud, <i>Gammarus lacustris</i>	95	EC50=1.8 ppb	98	EC50=0.8 ppb
FW shrimp, <i>Palaemonetes kadiakensis</i>	95	LC50=12 ppb	98	LC50=1.05 ppb
Waterflea, <i>Simocephalus serrulatus</i>	95	EC50=0.59 ppb	98	EC50=5.0 ppb
Seed shrimp, <i>Cypidopsis vidua</i>	95	LC50=47 ppb	98	LC50=32 ppb
Blue crab, <i>Callinectes sapidus</i>	95	LC50>1000 ppb	100	LC50=320 ppb
Oyster, <i>Crassostrea virginica</i>	95	EC50>1000 ppb	100	LC50=90 ppb
Sowbug, <i>Asellus brevicaudus</i>	95	LC50=3000 ppb	98	LC50=34 ppb
Cutthroat trout, <i>Oncorhynchus clarki</i>	95	LC50=1740 ppb	98	LC50=6.2 ppb
Yellow perch, <i>Perca flavens</i>	95	LC50=263 ppb	98	LC50=17.5 ppb
Channel catfish, <i>Ictalurus punctatus</i>	95	LC50=7620 ppb	98	LC50=52 ppb
Bluegill sunfish, <i>Lepomis macrochirus</i>	95	LC50=20 ppb	98	LC50=32 ppb
Rainbow trout, <i>Oncorhynchus mykiss</i>	95	LC50=4 ppb	98	LC50=11 ppb
Spot, <i>Leiostomus xanthurus</i>	95	LC50=320 ppb	100	LC50=23 ppb

Based on the data reviewed thus far for the two chemicals it would appear that the mixture may prove more toxic to most species of aquatic organisms if based on an equivalent active ingredient % of malathion alone. There will, however, be species sensitivity differences in some instances.

## Ecological Risk Assessment

### Exposure and Risk to Nontarget Terrestrial Wildlife

The acute risk quotients for broadcast applications of nongranular products are tabulated below. They are based on estimated acute and chronic residue levels calculated in the terrestrial exposure portion of this document divided by the LC50 or chronic NOEC of the most sensitive species tested.

#### 1. Birds

Avian Acute and Chronic Risk Quotients for Single Application of Nongranular Products (Broadcast or Foliar Spray) are based on the most sensitive species ringneck pheasant LC50 of 2639 ppm and the chronic NOEC for bobwhite quail of 110 ppm.

Multiple application scenarios were estimated using a first order dissipation program incorporating Fletcher values in conjunction with the appropriate half-life values. The worst case scenario for each application rate is reflected in the table, that is the minimum interval and maximum number of applications permitted under tolerance testing for this crop group. A mean foliar dissipation half-life of 5.5 days was inputted into the program, based on monitored values from several studies including USDA bollweevil and medfly programs and research efforts by Willis and McDowell, 1987 (referenced in previous terrestrial exposure section). Samples of the actual outputs are included as addendums to this document.

The risk quotient results indicate that for a single broadcast application of nongranular products, avian acute high (0.5), restricted use (0.2), and endangered species (0.1) levels of concern are exceeded at registered multiple application rates equal to or above 3.75 lb ai/A , 2.0 lb ai/A and 0.94 lb ai/A , respectively.

Table 30

## Avian Acute Dietary Risk Quotient Ranges

Cheminova and IR4 Supported Maximum Tolerance Rates and Scenarios on Grasses-Seed

	Foliar Dissipation T1/2=5.5 Days			Number of Applications									
	Rate	Int Day	1 grass-seed	2	3	4	5	6	7	8	9	10	12-25
A	0.175	7D	0.01-0.0004									0.02-0.001	
B	0.50	NA	0.04-0.001										
C	0.61	5D	0.05-0.001				0.11-0.007						
C	0.61	7D	0.05-0.001	0.07-0.003	0.09-0.005								
C	0.61	14D	0.05-0.001	0.06-0.004									
D	0.76	10D	0.06-0.002				0.09-0.006						
E	0.94	3D	0.08-0.002										
E	0.94	6D	0.08-0.002					0.15-0.01					
E	0.94	7D	0.08-0.002		0.13-0.008					0.15-0.009			
F	1.0	7D	0.09-0.002					0.15-0.009					
G	1.25	3D	0.1-0.003	0.19-0.01				0.32-0.02					
G	1.25	5D	0.1-0.003				0.23-0.014						
G	1.25	7D	0.1-0.003		0.17-0.01	0.19-0.01	0.19-0.01	0.19-0.01	0.19-0.01	0.19-0.01	0.19-0.01	0.19-0.01	0.19-0.01
G	1.25	14D	0.1-0.003	0.13-0.008									
H	1.50	7D	0.13-0.003		0.21-0.01			0.23-0.01					
I	1.56	7D	0.14-0.004	0.16-0.006			0.23-0.01						
J	1.88	5D	0.17-0.004					0.36-0.02					
J	1.88	7D	0.17-0.004		0.27-0.02	0.28-0.02		0.29-0.02					
J	1.88	14D	0.17-0.004	0.2-0.01									
K	2.03	6D	0.18-0.005					0.34-0.02					
K	2.03	7D	0.18-0.005		0.29-0.02	0.30-0.02							



L	2.5	3D	0.22-0.006		0.5-0.02								0.7-0.04
<b>Avian Acute Dietary Risk Quotients(continued)</b>													
L	2.5	5D	0.22-0.006		0.42-0.03								
L	2.5	7D	0.22-0.006		0.36-0.02		0.38-0.02						
M	3.43	5D	0.31-0.009				0.42-0.02						
N	3.75	7D	0.34-0.01			0.56-0.02		0.58-0.04					
N	3.75	14D	0.34-0.01										
O	4.7	30D	0.42-0.01	0.43-0.01									
P	5.0	7D	0.45-0.01	0.64-0.01	0.72-0.3	0.75-0.03							
Q	6.25	30D	0.57-0.02		0.58-0.01								

- 0.175 lb ai/A A10=Orange, Grapefruit, Lemon, Lime, Tangerine, Tangelo, and Kumquat
- 0.50 lb ai/A B1=Flax
- 0.61 lb ai/A C5(5D)=Sweet Corn , C2(7D)=Hops, C3(7D)=Beans, Corn, Rice, Sorghum, Wheat, and Rye  
C2(14D)=Alfalfa, Clover, Lespedeza, Lupine and Vetch
- 0.76 lb ai/A D5=Blueberry
- 0.94 lb ai/A E1(3D)=Grass for hay, E4(3D)=Mushroom, E6(6D)=Strawberry, E3(7D)=Pepper and spearmint,  
E7(7D)=Macadamia
- 1.0 lb ai/A F6(7D)=Melons, Watermelon, Pumpkin and Squash
- 1.25 lb ai/A G1(3D)=Grass for hay, G2(3D)=Field corn , G6(3D)=Mustards, G25(3D)=Cotton,  
G5(5D)=Watercress, G3(7D)=Rice, Sorghum, Wheat, Rye, Barley, Oats and Corn, G4(7D)=Blueberry(ULV),  
G5(7D)=Turnip, Broccoli, Apple, Sweet Corn, Beet, Chayote, Horseradish, Parsnip, Radish, Rutabaga, Salsify, Sweet potato,  
G6(7D)= Cabbage and Cherry(ULV), G7(7D)=Carrot , G8(7D)=Mango and Passion fruit , G9(7D)=Asparagus G10(7D)=Pears and Quince , G12(7D)=Guava and Papaya,  
G2(14D)=Alfalfa, Clover, Lupine, Vetch nd Lespedeza
- 1.5 lbs ai/A H2(7D)=Celery, H6(7D)=Okra
- 1.56lbs ai/A I2(7D)=Potato, I5(7D)=Onion, Garlic, Shallot, Leeks
- 1.88 lb ai/A J6(5D)=Lettuce, J4(7D)=Blackberry, Raspberry, Loganberry, Boysenberry, Dewberry, Currant, Gooseberry,  
J3(7D)=Cucumber, J6(7D)= Strawberry, J2(14D)=Grapes
- 2.03 lbs ai/A K6(6D)=Strawberry(50%WP), K3(7D)= Spinach, Dandelion, Endive, Parsley and Swiss Chard,  
K4(7D)=Blackberry, Raspberry, Gooseberry, Loganberry, Dewberry, Currant and Boysenberry
- 2.50 lb ai/A L25(3D)=Cotton, L3(5D)=Figs, L3(7D)=Mustards, Walnuts, and Pecans, L5(7D)=Peas
- 3.43 lb ai/A M5(5D)=Tomato, Pepper, Eggplant
- 3.75 lb ai/A N4(7D)=Apricots, N6(7D)=Cherry, N4(14D)=Peach and Nectarine
- 4.7 lb ai/A O2(30D)=Avocado
- 5.0 lb ai/A P3(7D)=Pineapple, P4(7D)=Chestnuts
- 6.25 lb ai/A Q3(30D)=Oranges, Grapefruit, Lemon, Lime, Tangerine and Tangelo

Chronic risk quotients can be calculated based on the average residues on food items. Average residues result from the pesticide being applied repeatedly, but degrading over the course of time from the first application to the last application. Due to rapid malathion degradation characteristics, high numbers of applications and minimal intervals in many cases, birds are expected to be exposed to continuous peaks

at 3, 5, 6, 7, or 10 day intervals. Avian chronic risk quotients based on average residues for multiple, broadcast applications of non-granular products may not be as pertinent under this type of scenario, therefore maximum peaks were compared against the NOEC for bobwhite quail chronic test results. The results, depicted in the table which follows, indicate that for multiple broadcast applications of nongranular products based on expected peak residues, the avian chronic level of concern is exceeded at a registered maximum application rate equal to or above 0.5 lb ai/A on grasses (based on the assumption of chronic effects due to repeated exposure to peak residues with less than one week intervals). This chronic level could be maintained by continuous and repetitive applications during a crop season.

**Table 31. Avian Chronic Risk Quotient Ranges**

Cheminova and IR4 Supported Maximum Tolerance Rates and Scenarios on Grasses-Seeds

	Foliar Dissipation T1/2=5.5 Days			Number of Applications									
	Rate lb ai/A	Int. Day	1	2	3	4	5	6	7	8	9	10	12-25
A	0.175	7D	0.4-0.01									0.7-0.04	
B	0.50	NA	1.09-0.03										
C	0.61	5D	1.3-0.03				2.7-0.17						
C	0.61	7D	1.3-0.03	1.7-0.08	2.1-0.12								
C	0.61	14D	1.3-0.03	1.6-0.1									
D	0.76	10D	1.7-0.05				2.3-0.14						
E	0.94	3D	2.0-0.06										
E	0.94	6D	2.0-0.06					3.8-0.05					
E	0.94	7D	2.0-0.06		3.2-0.2				3.5-0.2				
F	1.0	7D	2.18-0.06					3.7-0.22					
G	1.25	3D	2.72-0.08	4.6-0.3				7.8-0.5					
G	1.25	5D	2.72-0.08				614-38						
G	1.25	7D	2.72-0.08		4.3-0.3	4.4-0.3	4.6-0.3	4.6-0.3	4.6-0.3	4.6-0.3	4.6-0.3	4.6-0.3	4.6-0.3
G	1.25	14D	2.72-0.08	3.2-0.2									
H	1.50	7D	3.3-0.09		5.2-0.33			5.5-0.34					
I	1.56	7D	3.4-0.1	3.9-0.14			5.7-0.35						
J	1.88	5D	4.1-0.11					8.6-0.5					
J	1.88	7D	4.1-0.1		6.5-0.4	6.8-0.4		7.0-0.4					
J	1.88	14D	4.1-0.1	4.8-0.3									
K	2.03	6D	4.4-0.1					8.2-0.5					
K	2.03	7D	4.4-0.1		7.0-0.4	7.3-0.45							

L	2.5	3D	5.45-0.2		11.7-0.5								17.3-1.1
---	-----	----	----------	--	----------	--	--	--	--	--	--	--	----------

Table 32. Avian Chronic RQ Table for Shortgrass and Seeds(continued)

	Rate lb ai/A	I	1	2	3	4	5	6	7	8	9	10	12-25
L	2.5	5D	5.45-0.2				10.1-0.6						
L	2.5	7D	5.45-0.2				8.6-0.5		9.1-0.6				
M	3.43	5D	7.6-0.2						9.2-0.6				
N	3.75	7D	8.1-0.24				13.5-0.6			13.9-0.9			
N	3.75	14D	8.1-0.24										
O	4.7	30D	10.2-0.3	10.4-0.3									
P	5.0	7D	10.9-0.3	15.4-0.4	17.3-0.6	18.1-0.7							
Q	6.25	Q1	13.6-0.4			14.0-0.4							

- 0.175 lbai/A A10=Orange, Grapefruit, Lemon, Lime, Tangerine, Tangelo, and Kumquat
- 0.50 lb ai/A B1=Flax
- 0.61 lb ai/A C5(5D)=Sweet Corn , C2(7D)=Hops, C3(7D)=Beans, Corn, Rice, Sorghum, Wheat, and Rye,  
C2(14D)=Alfalfa, Clover, Lespedeza, Lupine and Vetch
- 0.76 lb ai/A D5=Blueberry
- 0.94 lb ai/A E1(3D)=Grass for hay, E4(3D)=Mushroom, E6(6D)=Strawberry, E3(7D)=Pepper and spearmint,  
E7(7D)=Macadamia
- 1.0 lb ai/A F6(7D)=Melons, Watermelon, Pumpkin and Squash
- 1.25 lb ai/A G1(3D)=Grass for hay, G2(3D)=Field corn , G6(3D)=Mustards, G25(3D)=Cotton,  
G5(5D)=Watercress, G3(7D)=Rice, Sorghum, Wheat, Rye, Barley, Oats and Corn, G4(7D)=Blueberry(ULV),  
G5(7D)=Turnip, Broccoli, Apple, Sweet Corn, Beet, Chayote, Horseradish, Parsnip, Radish, Rutabaga,  
Salsify, Sweet potato, G6(7D)= Cabbage and Cherry(ULV), G7(7D)=Carrot , G8(7D)=Mango and Passion fruit ,  
G9(7D)=Asparagus G10(7D)=Pears and Quince , G12(7D)=Guava and Papaya,  
G2(14D)=Alfalfa, Clover, Lupine, Vetch nd Lespedeza
- 1.5 lbs ai/A H2(7D)=Celery, H6(7D)=Okra
- 1.56lbs ai/A I2(7D)=Potato, I5(7D)=Onion, Garlic, Shallot, Leeks
- 1.88 lb ai/A J6(5D)=Lettuce, J4(7D)=Blackberry, Raspberry, Loganberry, Boysenberry, Dewberry, Currant,  
Gooseberry, J3(7D)=Cucumber, J6(7D)= Strawberry, J2(14D)=Grapes
- 2.03 lbs ai/A K6(6D)=Strawberry(50%WP), K3(7D)= Spinach, Dandelion, Endive, Parsley and Swiss Chard,  
K4(7D)=Blackberry, Raspberry, Gooseberry, Loganberry, Dewberry, Currant and Boysenberry
- 2.50 lb ai/A L25(3D)=Cotton, L3(5D)=Figs, L3(7D)=Mustards, Walnuts, and Pecans, L5(7D)=Peas
- 3.43 lb ai/A M5(5D)=Tomato, Pepper, Eggplant
- 3.75 lb ai/A N4(7D)=Apricots, N6(7D)=Cherry, N4(14D)=Peach and Nectarine
- 4.7 lb ai/A O2(30D)=Avocado
- 5.0 lb ai/A P3(7D)=Pineapple, P4(7D)=Chestnuts
- 6.25 lb ai/A Q3(30D)=Oranges, Grapefruit, Lemon, Lime, Tangerine and Tangelo

## Mammals

Birds and mammals have similar responses to xenobiotics. Birds have lower hepatic microsomal mono-oxygenase and A-esterase activity than do mammals. Therefore, birds are more susceptible than mammals to both organophosphate and carbamates in general. Malathion does not present an acute risk to mammals based on the low toxicity observed in exposure studies conducted with laboratory rats, rabbits and mice.

However, malathion does present a potential for long-term dietary exposure to mammals if multiple applications are repeated with inadequate intervals to allow for complete degradation. Malathion does appear to offer potential chronic hazard to birds, but hazard to mammals appears to be less likely. In 2 year oncogenic studies with laboratory rats (Food and Drug Research Labs, 1980-ACC 248179-180) the animals were fed diets containing 0, 1000 and 5000 ppm of 92.1 % malathion. No gross adverse effects were noted, however decreased cholinesterase levels and body weight were noted at 1000 ppm test levels. In another study male and female rats were fed 4000 ppm of malathion in their diets (equivalent to 240 mg/kg/day) for five months. Reduced litter size and survival of young was observed in this study (Kalow and Marton, 1965). These effect levels are above those expected on vegetation from the highest rate scenario (1500 ppm on vegetation surrounding citrus at 6.25 lb ai/A). However, temporary reduction of acetylcholinesterase levels is expected at higher rates of application.

Estimating the potential for adverse effects to wild mammals is based upon EEB's draft 1995 SOP of mammalian risk assessments and methods used by Hoerger and Kenaga (1972) as modified by Fletcher *et al.* (1994). The concentration of malathion in the diet that is expected to be acutely lethal to 50% of the test population (LC50) is determined by dividing the LD50 value (usually rat LD50) by the % (decimal of) body weight consumed. A risk quotient is then determined by dividing the EEC by the derived LC50 value. Risk quotients can then be calculated for three separate weight classes of mammals (15, 35, and 1000 g), each presumed to consume four different kinds of food (grass, forage, insects, and seeds). The acute risk quotients for broadcast applications of nongranular products are tabulated using the equations below. The reviewer calculated quotients based on a full range of application scenarios, but not on every possible scenario. In addition larger mammals were not included as data suggests that toxicity thresholds will not be attained for the higher weight classes of mammals. As with the risk quotient for birds, the driving influence on how high field residues (and thus the risk quotients) will be from multiple applications appears to be determined by the interval between applications more than the total number of applications of malathion. Chronic exposure to malathion is more a matter of continuous exposure to peak levels on a 3, 5, 6, 7, or 14 day cycles. Intervals of less than 7 days may allow buildup of malathion residues levels over time. Unfortunately, many of the mammalian chronic studies conducted for human health analysis are two year studies which are not truly comparable to a single season exposure period expected for wild mammals. In many of the chronic mammal studies less noticeable sublethal effects were noted such as reduced acetylcholinesterase levels, in brain, blood and plasma (Hazelton Labs, AMA Arc. Occ. MED:8; 1953) or gastric ulcers (Nat. Cancer Institute, 1979). These types of effects would go unnoticed during field use in all probability.

**Mammalian (Herbivore/Insectivore) Acute Risk Quotients for Single and Multiple Application of Nongranular Products (Broadcast) Based on rat LD50 of 390 mg/Kg**

$$RQ = \frac{EEC \text{ (ppm)}}{LD50 \text{ (mg/kg)} / \% \text{ Body Weight Consumed}} \quad \text{or} \quad \frac{EEC}{NOEC}$$

**Table 32. Worst Case RQ's for Dietary Consumption by Small Mammals:**  
 Small Mammal-15 gram Wt consuming 95% of Food Matter as Shortgrass or Fruit  
 Small Mammal of 35 gm Wt consuming 66% of Food Matter as Shortgrass or Fruit

Site/App. Method	Rate (lbs ai/A)	Single Application Day 0 Max. EEC Range for Short grass to Fruits	Highest # Applic. (Minimum Interval)	Multi-App. Max. EEC Range-Shortgrass to Fruit	Acute RQ Range 15 g Body Wt Consuming 95%	Acute RQ Range 35 g BodyWt Consuming 66%
Citrus/Aerial	0.175	42 to 1.2 ppm	10X(7D)	43 to 2.7	0.10- 0.006	0.07 - 0.0045
Corn/Aerial	0.61	146 to 4.3 ppm	3X(7D)	195 to 12	0.47 - 0.029	0.33 - 0.02
Blueberry/Aerial	0.76	182 to 5 ppm	5X(10D)	195 to 12	0.47 - 0.029	0.33 - 0.02
Strawberry/Ground	0.94	226 to 7 ppm	6X(6D)	278 to 17	0.67 - 0.04	0.47 - 0.02
Melons/Ground	1.0	240 to 7.5 ppm	6X(7D)	280 to 17	0.68 - 0.04	0.48 - 0.02
Cotton/Aerial	1.25	300 to 8.8 ppm	25X(3D)	531 to 33	1.3 0 - 0.08	0.90 - 0.056
Onion/Ground	1.56	374 to 11 ppm	5X(7D)	437 to 27	1.06 - 0.07	0.74 - 0.046
Lettuce/Ground	1.88	451 to 13 ppm	6X(5D)	601 to 37	1.46 - 0.09	1.01 - 0.06
Strawberry/Ground	2.03	487 to 14 ppm	6X(7D)	526 to 33	1.3 - 0.08	0.90 - 0.056
Cotton/Aerial	2.50	600 to 18 ppm	25X(5D)	733 to 35	1.78 - 0.09	1.24 - 0.05
Tomato/Ground	3.43	823 to 24 ppm	5X(5D)	1096-68	2.67 - 0.16	1.86 - 0.11
Cherry/Ground	3.75	900 to 26 ppm	6X(7D)	1050-60	2.56 - 0.14	1.78 - 0.10
Avocado/Ground	4.7	1128 to 33 ppm	2X(30D)	1128-35	2.75 - 0.08	1.91 - 0.08
Pineapple/Ground	5.0	1200 to 35 ppm	4X(7D)	1400-40	3.41 - 0.09	2.37 - 0.068
Citrus/Ground	6.25	1500 to 44 ppm	2X(30D)	1500-44	3.65-0.074	2.54 - 0.07

**Table 33.**

**Chronic RQ Ranges for Mammals - Exposure to Multiple/Continuous Residue Peaks**

Based on Chronic Studies with Mice(MRID 242903) and rats (Document # 000389, Karlow and Marton, 1965)  
 Reduced Body Wt. -Mice At 500 PPM      Reduced Pup Survival for Rats at 4000 ppm

Site/Method Application	Rate (lb ai/A)	# of Apps. (Interval)	Maximum EEC Range in PPM	Chronic RQ Growth	Chronic RQ Reproduction
Citrus/Aerial	0.175	10X(7D)	43 to 2.7	0.09-0.005	very low
Corn/Aerial	0.61	3X(7D)	195 to 12	0.39-0.024	very low
Blueberry/Aerial	0.76	5X(10D)	195 to 12	0.39-0.024	very low
Strawberry/Ground	0.94	6X(6D)	278 to 17	0.55-0.03	very low
Melons/Ground	1.0	6X(7D)	280 to 17	0.56-0.03	very low
Cotton/Aerial	1.25	25X(3D)	531 to 33	1.06-0.07	0.13-0.008
Onion/Ground	1.56	5X(7D)	437 to 27	0.87-0.05	very low
Lettuce/Ground	1.88	6X(5D)	601 to 37	1.2-0.07	very low
Strawberry/Ground	2.03	6X(7D)	526 to 33	1.05-0.07	very low
Cotton/Aerial	2.50	25X(5D)	733 to 35	1.46-0.07	0.18-0.009
Tomato/Ground	3.43	5X(5D)	1096-68	2.19-0.13	0.27-0.017
Cherry/Ground	3.75	6X(7D)	1050-60	2.10-0.12	0.26-0.02
Avocado/Ground	4.7	2X(30D)	1128-35	2.26-0.07	0.28-0.008
Pineapple/Ground	5.0	4X(7D)	1400-40	2.80-0.08	0.35-0.01
Citrus/Ground	6.25	2X(30D)	1500-44	3.0-0.09	0.38-0.01

**Malathion Used in Bait Applications**

Though no granular malathion products are proposed for reregistration, malathion is used in a number of bait application uses. These liquid bait applications may be similar to granules in their route of ingestion by exposed wildlife. Mammalian species also may be exposed to bait droplets containing concentrated (95% ai) malathion. This would be applicable to such programs as the medfly eradication programs where malathion protease baits are employed to attract the target organisms (Mediterranean fruit fly). They also may be exposed by other routes, such as by walking on exposed bait and drinking water contaminated by malathion baits. The number of lethal doses (LD50's) that are available within one square foot immediately after application can be used as a risk quotient (LD50's/ft<sup>2</sup>) for the various types of exposure to bait pesticides. Risk quotients are calculated for a small mammal and for the ringneck pheasant.

102

### Mammalian Acute Risk Quotients for Bait Products (Broadcast).

<u>Use Site</u>	<u>Application Method</u>	<u>Rate in lbs ai/A</u>	<u>% Surface Residues</u>
Medfly Control	Aerial	0.18	80% efficiency est.

Body Weight (g) Rat =100 gm  
Based on a rat LD50 of 390 mg/Kg

Ringneck Pheasant=1135 gm  
Ringneck LD50=167 mg/Kg

Mammalian Acute RQ<sup>1</sup> (LD50/ft<sup>2</sup>)= 0.0000004

$$^1 \text{ RQ} = \frac{\text{Rate (lbs ai/A)} * (453,590 \text{ mg/lbs}/43,560 \text{ ft}^2/\text{A}) * 80\%}{\text{LD50 mg/kg} * \text{Weight of Animal (g)} * 1000 \text{ g/kg}} = \frac{65316 \text{ mg}/43560}{390 * 100 * 1000} = \frac{1.5 \text{ mg/sqft}}{3900000}$$

$$\text{Avian Acute Oral RQ (Pheasant)} = \frac{1.5 \text{ mg/sq ft}}{189545000} = 0.00000001$$

The results above indicate that for aerial application of protease bait products at 0.18 lbs ai/A, no mammalian or avian acute levels of concern are exceeded. Currently, EFED has no procedure for assessing chronic risk to mammalian species for protein bait products.

### Hazard to Non-Target Insects

Currently, EFED does not quantify risk to nontarget insects. Results of acceptable studies and actual field use observations are used for recommending appropriate label precautions. Acute toxicity to honeybees from acute contact or foliar contact with malathion residues is very high. Based on these acute studies and observations from field studies presented under the previous toxicity to insects section of this document, acute hazard is expected for non-target pollinator insects (honeybees, etc) exposed to direct spray droplets, to residues on foliage, or to residues which are transported on pollen back to the hives or nests (Gary, N.E., 1984). This hazard can extend to pollinators with hives located several kilometers away from the application site, dependent on the distance range of flight paths associated with the particular species in question. Several field studies have shown increased mortality for colonies located as much as two kilometers away from application sites. Many other beneficial species of insects and arachnids (lacewings, butterfly larvae and adults, spiders, beetles etc) are vulnerable to non-crop spray applications which are used to control other pests of public concern such as medflies, mosquitoes, and flies (Dahlsten, D.L., 1983; Johansen, C.A., 1965). Spraydrift to aquatic habitats may produce adequate residue levels to prove hazardous to aquatic larvae of insects which later become important terrestrial members of the insect community (eg. dragonflies, mayflies, damselflies, snipeflies, caddisflies, stoneflies etc.). Mortality to these types of larvae may occur at aquatic concentrations as low as 1 PPB. Studies by L.D. Jenson, 1965 showed that even after stonefly larvae were removed from exposure areas and placed in clean water mortality could still occur within 24 hours. Many of these larvae also serve as important food sources for juvenile fish.

## Risk to Nontarget Freshwater or Estuarine Aquatic Organisms

Based on actual monitored concentrations, predicted modeling results, and actual fish kill incidents, there is acute hazard from contamination of aquatic habitats adjacent to or within target application areas. Tables presented below represent risk quotients for various application scenarios for agricultural and public health uses of malathion. Risk quotients which exceed 0.5 are considered to present acute hazard to the species in question. Risk quotients which exceed 0.1 are considered to offer potential hazard to endangered species within these groups (fish, crustacea, molluscs, amphibia, etc). The tables below present risk quotients for invertebrates and fish in the same table. The first number in each scenario cell pertains to the RQ associated with the acute EC50 (1 ppb) or chronic NOEC (0.1 ppb) associated with *Daphnia magna*. The second number in the cell represents the RQ for fish based on the LC50 of the bluegill sunfish (20 ppb) or the chronic NOEC for the rainbow trout early life stage test (4 ppb). The RQs are derived using predicted EECs from GENEEC (tables 35 and 36) or PRZM/EXAMS (table 37) and dividing them by the acute or chronic toxicity endpoints.

**Table 34. Aquatic Organism Acute Risk Quotients-Invertebrate RQ/Fish RQ**  
Cheminova and IR4 Supported Maximum Tolerance Rates and Crop Scenarios

Number of Applications													
	Rat e	Int.	1	2	3	4	5	6	7	8	9	10	12-25
A	0.175	7D	Inv/Fish								8.2/0.4		
B	0.50	NA	11.4/0.57										
C	0.61	5D					*						
C		7D		23.2/1.2	27.7/1.4								
C		14D		26.8/1.3									
D	0.76	10D					40.6/2.0						
E	0.94	3D	21.7/1.1										
E		6D						45.4/2.3					
E		7D			42.2/2.1				42.5/2.1				
F	1.0	7D						45.2/2.3					
G	1.25	3D	28.5/1.4	54.3/2.7				90.4/4.5					91.9/4.5
G		5D					66/3.3						
G		7D			56.1/2.8	56.5/2.8	56.5/2.8	57.2/2.9-A	56.5/2.8	56.6/2.8	56.6/2.8	56.6/2.8	
G		14D		47.1/2.4									



H	1.50	7D			67.3/3.4			67.3/3.4					
I	1.56	7D		67.8/3.4			70.8/3.5						
J	1.88	5D						99.4/5					
J		7D			84.4/4.2	84.9/4.2		85/4.2					
J	1.88	14D		70.8/3.5									
K	2.03	6D						98/4.9					
K		7D			91.1/4.6	91.7/4.6							
L	2.5	3D											181/9
L		5D			128/6.4								
L		7D			112/5.6		113/5.6						
M	3.43	5D					181/9						
N	3.75	7D				169/8.4		169/8.4					
N		14D					142/7.1						
O	4.7 lb	30D		171/8.5									
P	5.0	7D			224/11.2	225/11.2							
Q	6.25	30D			226/11.2								

**Table 35. Aquatic Organism Chronic Risk Quotient (Invertebrates RQ/Fish RQ)**  
 (Cheminova and IR4 Supported Max. Tolerance Rates and Scenarios- 0.175 to 1.25 lbs ai/Acre)

		Number of Applications											
	Rate lb ai/A	Int. Day	1	2	3	4	5	6	7	8	9	10	12-25
A	0.175	7D	inv/fish									20/0.2	
B	0.50	NA	28/0.26										
C	0.61	5D		57/0.55	68/0.65								
C		7D		66/0.65									
C		14D											
D	0.76	10D					100/0.85						
E	0.94	3D	100/0.5										

E		6D						112/1.06					
E		7D			104/0.98				10.5/0.99				
F	1.0	7D						112/1.05					
G	1.25	3D	70/0.66	134/1.26				223/2.13					23/8.7
G		5D					163/1.54						
G		7D			138/1.31	139/1.32	139/1.32	141/1.35 (Aerial)	139/1.32	139/1.3	139/1.3	139/1.3	
G		14D		116/1.13									
H	1.50	7D			166/1.57			166/1.57					
I	1.56	7D		167/1.58			175/1.65						
J	1.88	5D						246/2.32					
J		7D			208/1.96	21/1.98		21/1.99					
J	1.88	14D		175/1.65									
	Rate lb ai/A	Int. Day	1	2	3	4	5	6	7	8	9	10	12-25
K	2.03	6D						242/2.29					
K		7D			225/2.12	226/2.14							
L	2.5	3D											447/4.2
L		5D			316/3.0								
L		7D			277/2.6		279/2.7						
M	3.43	5D					200/4.23						
N	3.75	7D				417/3.95		417/3.95					
N		14D				350/3.3							
O	4.7 lb	30D		422/3.98									
P	5.0	7D			553/5.23	556/5.28							
Q	6.25	30D			558/5.3								

0.175 lbai/A

0.50 lb ai/A

0.61 lb ai/A

0.76 lb ai/A

0.94 lb ai/A

1.0 lb ai/A

A10=Orange, Grapefruit, Lemon, Lime, Tangerine, Tangelo, and Kumquat

B1=Flax

C5(5D)=Sweet Corn , C2(7D)=Hops, C3(7D)=Beans, Corn, Rice, Sorghum, Wheat, and Rye

C2(14D)=Alfalfa, Clover, Lespedeza, Lupine and Vetch

D5=Blueberry

E1(3D)=Grass for hay, E4(3D)=Mushroom, E6(6D)=Strawberry, E3(7D)=Pepper and spearmint, E7(7D)=Macadamia

F6(7D)=Melons, Watermelon, Pumpkin and Squash

**1.25 lb ai/A** G1(3D)=Grass for hay, G2(3D)=Field corn , G6(3D)=Mustards, G25(3D)=Cotton,  
 G5(5D)=Watercress, G3(7D)=Rice, Sorghum, Wheat, Rye, Barley, Oats and Corn,  
 G4(7D)=Blueberry( ULV), G5(7D)=Turnip, Broccoli, Apple, Sweet Corn, Beet, Chayote,  
 Horseradish, Parsnip, Radish, Rutabaga, Salsify, Sweet potato, G6(7D)= Cabbage and  
 Cherry(ULV), G7(7D)=Carrot , G8(7D)=Mango and Passion fruit , G9(7D)=Asparagus  
 G10(7D)=Pears and Quince , G12(7D)=Guava and Papaya, G2(14D)=Alfalfa, Clover, Lupine,  
 Vetch and Lespedeza  
**1.5 lbs ai/A** H2(7D)=Celery, H6(7D)=Okra  
**1.56lbs ai/A** I2(7D)=Potato, I5(7D)=Onion, Garlic, Shallot, Leeks  
**1.88 lb ai/A** J6(5D)=Lettuce, J4(7D)=Blackberry, Raspberry, Loganberry, Boysenberry, Dewberry, Currant,  
 Gooseberry, J3(7D)=Cucumber, J6(7D)= Strawberry, J2(14D)=Grapes  
**2.03 lbs ai/A** K6(6D)=Strawberry(50%WP), K3(7D)= Spinach, Dandelion, Endive, Parsley and Swiss Chard,  
 K4(7D)=Blackberry, Raspberry, Gooseberry, Loganberry, Dewberry, Currant and Boysenberry  
**2.50 lb ai/A** L25(3D)=Cotton, L3(5D)=Figs, L3(7D)=Mustards, Walnuts, and Pecans, L5(7D)=Peas  
**3.43 lb ai/A** M5(5D)=Tomato, Pepper, Eggplant  
**3.75 lb ai/A** N4(7D)=Apricots, N6(7D)=Cherry, N4(14D)=Peach and Nectarine  
**4.7 lb ai/A** O2(30D)=Avocado  
**5.0 lb ai/A** P3(7D)=Pineapple, P4(7D)=Chestnuts  
**6.25 lb ai/A** Q3(30D)=Oranges, Grapefruit, Lemon, Lime, Tangerine and Tangelo

**Table 36.**

<b>PRZM-EXAMS Derived Aquatic RQ's</b> (EECs Based on 1 in 10 Year Events)									
Crop	% of total a.i. applied / year <sup>1</sup>	Use rate (lbs a.i./A)	Interval (days)	No. of applications	Model Results (ppb)				
					PRZM-EXAMS			Acute/Chronic RQ's	
					peak <sup>2</sup>	21d ave	60 d ave. <sup>3</sup>	Inv.	Fish
Cotton	41.6%	Max: 2.5	3	25	291	67.4	47.7	291/674	14.5/24
		Typ: 0.3	[3] <sup>4</sup>	4	7.9	1.48	0.50	8/15	0.4/0.25
Alfalfa	23.5%	Max: 1.25	14	2					
		Typ: 1.4	[14]	1					
Sorghum	7.4%	Max: 1.25	7	3	26.7	5.01	1.95	27/50	1.3/1
		Typ: 0.8	[7]	1	2.94	0.50	0.18	3/5	0.15/0.1
Apple	2.14%	Max: 1.25	7	5	0.80	0.33	0.19	0.8/3	0.04/0.1
		Typ: 0.7	[7]	3	0.59	0.24	0.09	0.59/2	0.03/0.3
Citrus	0.49%	Max: 6.25	30	3	162	25.2	11.1	162/252	8.1/5.6
		Typ <sup>5</sup> : 2.5	[30]	1	47.3	7.38	2.59	48/74	23.6/1.3
Lettuce	0.45%	Max: 1.88	5	6	15.4	6.26	2.98	15/63	0.8/1.5
		Typ: 2.0	[5]	1	5.63	1.58	0.56	6/16	0.3/0.3

**PRZM EXAM Runs Correspond to the Following Use Scenario Numbers :**

- 1.25 lb ai/A** G3(7D)=Rice, Sorghum, Wheat, Rye, Barley, Oats and Corn, G5(7D)=Turnip, Broccoli, Apple, Sweet Corn, Beet, Chayote, Horseradish, Parsnip, Radish, Rutabaga, Salsify, Sweet potato, G2(14D)=Alfalfa, Clover, Lupine, Vetch and Lespedeza
- 1.88 lb ai/A** J6(5D)=Lettuce
- 2.50 lb ai/A** L25(3D)=Cotton
- 6.25 lb ai/A** Q3(30D)=Oranges, Grapefruit, Lemon, Lime, Tangerine and Tangelo

**Table 37. MALATHION NON AGRICULTURAL USE-Maximum Labeled Rates  
Worst Case EECs and RQs Direct Application to Water or 100% Drift\***

<b>USE LOCATION (Predicted U.S. Acreage)</b>	<b>Max. Rate lbs ai/A</b>	<b>Max # Applic.</b>	<b>Min. Interv</b>	<b>Acute Aquatic EEC s 0.5-6 ft</b>	<b>Acute Aquatic Risk Quotient</b>
Nonagricultural rights of way/fencerows/hedgerows (17,000 acres)	0.598	NS	NS	438 to 36 ppb	Inv.-438 to 36 Fish-22 to 1.8
Mosquito Control (8,227,000 acres) Lakes/Ponds/Reservoirs(human use)(0.5985) Nonag. Uncultivated Areas/Soils (0.6) Polluted Water (0.6)	0.630 ULV Aerial	NS	NS	462 to 38 ppb	Inv.-462 to 38 Fish-23 to 1.9
Lakes/Ponds/Reservoirs (No Human Use) (0.628) Swamps/Marshes/Wetlands/Stagnant Water (0.628) Intermittently Flooded Areas/Water (0.628)	0.630 Grnd Fogger	NS	NS	92.4 to 7.6 ppb (20% drift)*	Inv.-92 to 8 Fish-4.62 to 0.4
Woodland Use (17,000 acres) Pine Forest/Shelterbelt (0.9375) Eastern White Pine (Forest) (0.9375)	0.94	NS	NS	688 to 57 ppb	Inv.=688 to 57 Fish=34.4 to 2.8
Rangelands/Pastures/Set Aside Acreage/Summer Fallow (1,625,000 acres) Canarygrass (1.2) Rangeland or Pastures (1.25) Grass Forage/Fodder/Hay (1.25)	1.25	NS	NS	917 to 76 ppb	Inv.=917 to 76 Fish=46 to 3.8
Ornamental Plant Uses-Nurseries-Homeowner (175,000 acres) Ornamental trees and Herbaceous Plants	1.746	NS	NS	direct drift unlikely	Not computed
Ornamental Nonflowering Plants Ornamental Woody Shrubs and Vines (2.5)	2.50	NS	NS	direct drift unlikely	Not computed
Commercial Tree Production (no est. acreage) Christmas Tree Plantations, (3.125) Ornamental and/or Shade Trees (3.125) Slash Pine (forest) (3.125)	3.125	NS	NS	2293 to 190 ppb	Inv.=2293 to 190 Fish=114.6 to 9.5
Public Parks (67,000 acres)	NS			Rates not specified	Not computed
Turf Use/ Golfcourses/Commercial Lawncare Ornamental Lawns and Turf	5.1	NS	NS	direct drift unlikely	Not computed
No Non-Ag uses at higher rates (>5.1 lb ai/A)	Not supported				Not supported

NS=Not Specified

\*Drift from truck mounted foggers is not expected to exceed 20% deposition due to continuous drift of micro droplets on air currents.

### Endangered Species

Endangered species LOCs are exceeded for malathion for acute hazard to endangered fish, aquatic invertebrates, and insects for most outdoor uses. Chronic hazard LOC's to threatened birds, mammals, amphibians and reptiles are potentially exceeded for certain uses. Chronic hazard LOC's for endangered fish and invertebrates are exceeded by most uses. The magnitude

of malathion use and the numbers of potentially exposed endangered species will require more extensive analysis by the OPP Endangered Species Branch.

The Endangered Species Protection Program is expected to become final in the future. Limitations in the use of malathion will be required to protect endangered and threatened species, but these limitations have not been defined and may be formulation specific. EPA anticipates that a consultation with the U.S. Fish and Wildlife Service will be conducted in accordance with the species-based priority approach described in the Program. After completion of consultation, registrants will be informed if any required label modifications are necessary. Such modifications would most likely consist of the generic label statement referring pesticide users to use limitations contained in county Bulletins.

## Risk and Exposure Characterization

The following section identifies major routes of exposure expected to lead to effects on ecological resources and the highest exposure levels for drinking water sources.

### Summary of Expected Paths of Potential Exposure for Wildlife

#### Direct Application Public Health Use

Aerial and ground spray applications of malathion allow for coverage of large areas of urban, suburban, and rural areas. For instance malathion may be applied aerially "...over cities, towns, and other areas...." (Fyfanaon ULV Insecticide label) for adult mosquito control. Rates of malathion use for mosquito control are up to 9.9 lbs ai / mi<sup>2</sup> / year in the following states: Washington, Oregon, Utah, Wyoming, Texas, Louisiana, Mississippi, Michigan, Alabama, Florida, Georgia, South Carolina, North Carolina, New Jersey, and Virginia. In some areas urban and agricultural use of malathion may overlap. Terrestrial wildlife, insect, and adult amphibian exposure from this type of use is expected to be through a multitude of food sources receiving residue including vegetative food matter, insects, drinking water and also through direct dermal and inhalation exposure to spray applications. Aquatic exposure to fish, crustacea, mollusca, arthropoda, and larval amphibia is expected to be primarily from drift with lesser amounts contributed by runoff.

Malathion for adult mosquito control is applied by fogging and aerial methods. Thermal aerosols or fogs create very small droplets of malathion (<20µm) that can be carried on air currents for long distances before contacting plants, water, or soil. Because of this tendency, malathion in fogs is expected to dissipate largely through atmospheric diffusion with relatively little deposition onto water or soil. Wang *et al* (1987) studying fenthion fog deposition on water measured 5-6% deposition of the applied insecticide. EFED has used a more conservative estimate of 20%. For aerially applied ULV formulations higher deposition is expected because droplet size is larger (up to 100µm) and there are no specified protective buffer zones. Thus it is assumed by EFED that 100% deposition occurs in shoreline areas. Based on low toxicity thresholds of 0.5 ppb (invertebrates) to 10 ppb (fish), 20% to 100% drift scenarios, actual monitored residue levels and observed adverse effects in actual use situations risk quotients for aquatic invertebrates and fish are expected to be exceeded.

In areas close to the fogging apparatus or beneath aerial applications inhalation may be an important route of acute exposure for terrestrial wildlife. Mammalian toxicity data provided to HED show severe effects to rats in the lowest exposure group of 0.1 mg/l (96 h). The no effect level is not defined and it is not known what interspecies differences in sensitivity exist. Although it is not clear what atmospheric malathion concentrations are acceptable, the levels generated during mosquito control are expected to be very transient. Direct exposure to flying non-target insects is not only likely, but probably unavoidable as fogging type applications are designed to contact flying insects.

#### Ground and Airblast Application to Agricultural Sites

Normal malathion ground application usage in agricultural field settings presents chronic risks for terrestrial wildlife, but lower risk to aquatic life. Key issues identified are possible chronic effects resulting from repeated 3 to 7 day pulse exposure of birds to malathion at certain rates of application, increased risk to wildlife resulting from exposure to malathion products containing

mixtures of other insecticides and fungicides, and direct contact toxicity to beneficial insects from off target drift from agricultural target areas or later contact with residual malathion residues on the target crops. Some toxic exposure to aquatic organisms from small amounts of runoff is expected, though exposure from spraydrift will be less than for aerial application.

Synergistic toxicity resulting from coexposure to malathion and other pesticides has been noted in previous portions of this review. Not only are malathion mixtures with other pesticides marketed as end-use products, but agricultural use of malathion is commonly accompanied by the use of other pesticides in the same field leading to mixtures of residues in the field. Runoff and drift of malathion also mixes with residues of many other pesticides used in fields adjacent and in the same drainage basin. Some cholinesterase inhibiting insecticides are expected to result in additive toxicity when combined with malathion, although some other pesticides been shown to potentiate malathion toxicity. Greater than expected toxicity has been noted with certain cholinesterase inhibiting insecticides (including carbaryl and EPN) and some fungicides (including clotrimazole). The environmental and ecological effects of mixtures is poorly understood, but in many instances, increased sensitivity of organisms is expected. More data concerning the toxicities of these end-use product mixtures as well as mixture with other pesticides in normal agricultural use is needed to assess these concerns for malathion.

#### Spray Drift from Agricultural Uses

Monitoring results show that spray drift can be a major source of aquatic contamination. Drifting malathion applications carried by air movement will reach unintended sites. More than half of the malathion usage in the United States is applied in ULV formulations which are highly prone to drift when applied aurally. ULV formulations are popular with aerial applicators because they are very concentrated and allow the treatment of large acreages without returning to the airfield for refilling or refueling.

An assessment of drift as a result of malathion use methods for the Boll Weevil Eradication Program (presently 60% of all malathion use in U.S.) was conducted by measuring off-target drift adjacent to aerial ULV malathion applications (Penn State 1993). Application conditions were the same as those used in the eradication program. The spray system was a conventional boom and nozzle system fitted with Spraying Systems stainless steel 8002 Flat Fan spray tips. The nozzle position was straight down and the flying height was a nominal 5 feet above the crop canopy. Drift was measured from single aircraft passes delivering 1 lb/A. Wind direction was perpendicular to the flight path. Seventeen runs were conducted under varying meteorological conditions. Maximum depositions were 21, 12, 2.8, and 0.7% of the expected maximum at 100, 200, 300, and 1000m downwind (Penn State 1993). The highest amount of drift at 1 km occurred when atmospheric conditions were stable, meaning vertical air mass movements are dampened. Higher drift levels at shorter distances occurred under unstable, windy conditions. Averages of results under different atmospheric conditions show deposition of 9.4% at 100 m while at 1000m the deposition rate was 0.08%.

Using deposition rates from the Penn State study it is possible to calculate aquatic EECs for varying depths of water based on direct application of the expected % of drift and using a 6 inch to 6 foot depth range for the standard 1 acre farm pond scenario.



water depth (in)	100 m (21% deposition)			200 m (12% deposition)			300 m (2.8% deposition)		
	EEC	RQ (fish)	RQ (daphnid)	EEC	RQ (fish)	RQ (daphnid)	EEC	RQ (fish)	RQ (daphnid)
6	154	7.7	154	88	4.4	88	20.5	1.0	20.5
72	25.6	1.28	25.6	7.3	0.37	7.3	1.7	0.1	1.7

Based 1 lb/acre (used in the Penn State drift study) drifting to a one hectare pond. Risk quotients are based on fish LC50 for bluegill (20 ppb) and *Daphnia magna* LC50 (1 ppb). The effects of reducing spray drift was examined two ways: first, varying the drift parameters in PRZM-EXAMS modeling and, second, by comparing the effect different application practices and buffer strips on measured drift in monitoring studies.

Levels of concern for fish are exceeded at 100 and 200 m distances with these maximum drift values. At 300 m a level of safety is achieved for fish, but daphnids are still at risk. Monitored values of malathion drift to streams suggest that table above is conservative in estimating aquatic risk. A typical range of monitored values is shown in Table 13a ranging from non-detected to almost 11 ppb 25 feet from the field. In these applications wind direction was away from the water.

The Boll Weevil Eradication Program mostly uses ultra low volume (ULV) formulations in its program in several states. ULV which is ~95% malathion is the most cost effective formulation in the treatment of cotton for boll weevil because it is concentrated and enables aerial applicators to treat large areas before refueling and refilling. ULV applications results in the formation of small droplets of the pesticide which are prone to drift long distances. The speed by which droplets fall is exponentially dependent upon their size such that small droplets fall very slowly. Smaller droplets result in more nontarget deposition of pesticide through drift caused by wind, thermal air currents, and turbulence from applicator planes. Presently at least 14 different crops receive aerial ULV applications of malathion. These crops include alfalfa, blueberries, clover, cotton, dry beans, corn, sorghum, grass, lima beans, oranges, rice, snap beans, wheat, and cherries. A potential risk management measure is the use of formulations other than ULV for aerial applications. Drift from non-ULV formulations is significantly lower under analogous application conditions.

Monitoring studies suggest that reducing drift dramatically reduces aquatic EECs. Boll weevil treatments were examined for drift to surface water in the Southeast and Texas. Table 12a shows the effects of ground versus aerial application and varying buffer strips on malathion drift by measuring concentrations before and at 15 minute increments after application. Four different sites are examined with buffer strips ranging from 700 feet with 30-60 foot trees to 25 feet with low-lying vegetation. Five other sites shown in previous tables related to actual field monitoring results provide additional information on drift but lack site and application information which led to the monitored residue levels.

Monitoring data suggests that wider buffer strips and ground applications reduce drift. Aerial applications to two fields with 125 foot or greater buffers resulted in no measured drift. Aerial applications to a 95.3 acre field with a 100 foot buffer containing mature hardwoods (Pursley Creek site) resulted in minor drift: only three measurements of six total were greater than 3 ppb above background. Four aerial applications to a 19.2 acre field with a 25 foot buffer containing low kudzu vegetation (Stewart Creek) resulted in significant drift: all four events resulted in aquatic concentration exceeding 3 ppb. Five ground applications to the same site resulted in low drift: four of five ground applications resulted in aquatic concentration of less than 0.33 ppb. Additional measures that can be taken to reduce spray drift are placing limitations on application height (10 feet), wind speed (7 mph), boom pressure (40 psi), and jet nozzle size.

### Runoff in Urban Scenarios

Though initial exposure of non-target aquatic habitats is expected to be primarily through spray drift, monitored residue levels in residential storm water runoff events have yielded high residue levels, despite the short terrestrial half-life values that are reported for malathion. It should be noted that monitored runoff events in urban areas reflect aggregate malathion residues resulting from all uses of malathion in that particular drainage basin such as homeowner use, commercial turf use, municipal mosquito control use, and commercial agricultural use. There are approximately 60 home and garden products containing malathion and malathion/methoxychlor on the market.

Monitoring data of runoff from urban-use sites is frequently high probably due to increased runoff from impermeable surfaces and increased persistence on microbially inactive, dry surfaces. The fastest routes of malathion degradation are through aerobic metabolism and hydrolysis. Residential surfaces such as asphalt and concrete, which malathion is likely to contact in urban use, do not provide microbes and moisture required for these degradation pathways. A CalEPA study and monitoring data also suggest that the toxic degradate malaaxon is more likely to form on residential surfaces and occur more frequently in urban runoff. Anthropogenic surfaces are less likely to retain malathion during rainfall which would lead to pulses of malathion in storm water runoff which drain into urban streams. USGS NAQWA data show higher levels of malathion and more detects in urban streams than were monitored in rural and agricultural counterparts. In medfly control efforts south of San Francisco in 1984 residue levels increased significantly after rainfall events. Fish kills coincided with high levels of malathion (80-800 ppb) after rainfalls. Application rates and methods for mosquito control and medfly programs are similar, thus runoff resulting from urban mosquito control operations may be similar to those observed from medfly applications. Malathion is used in community mosquito control programs in at least 15 states up to 9.9 lbs / mi<sup>2</sup> / year. This assessment indicates that risk to aquatic life from runoff transported residues will be high in urban use scenarios.

### Runoff in Agricultural Scenarios

Agricultural field runoff of malathion to nontarget aquatic habitats has been observed to be generally low, probably due to rapid degradation on soil. Runoff monitored in the Boll Weevil Eradication Program suggests that a majority of malathion levels in receiving waters will not present a significant risk for fish. The risk level for invertebrates from runoff is less clear. Malathion in runoff from cotton fields ranged from none detected to 146 ppb in undiluted drainage. Dilution of runoff and therefore the degree of risk for invertebrates will vary with the

size of the body of receiving water. A dilution factor of nearly 300 is necessary to reduce the daphnid RQ to below 0.5. This would be expected in larger receiving water bodies but agricultural field runoff to small streams and ponds will result in higher risk.

#### Malathion Non-Crop Usage in Rural Scenarios

Malathion is used in a variety of settings which are rural in nature, but not related to a particular crop. Malathion ULV uses include aerial and ground application to control grasshopper and beet leafhopper on pasture lands, rangeland, "non-agricultural" lands (wasteland and roadsides), fencerows, feed-lots, clover (usually a cover crop) and summer fallow. In addition malathion ULV labels list woodland uses via aerial application to control forest insects on douglas fir, true fir, spruce, hemlock, pine, and larch trees to control budworms, looper, sawfly, spittlebug and larch casebearer. Registrants have stated an intention to remove forest uses, but this may not apply to privately owned wood lots and wooded lands. These use patterns are similar to mosquito control scenarios in that they are not directed at any particular field site, yet labeling language does not include specific instructions to aid in protection of sensitive aquatic habitats contained within these areas nor do they specify maximum seasonal application restrictions. They are also similar to agricultural sites in that soil degradation is likely to be more pronounced than in urban scenarios. The total acreage of these types of use sites will total over 2 million acres on a yearly basis.

Exposure risk to avian and mammalian species from repeated applications with narrow intervals is therefore concluded to exist for these use scenarios. Exposure risk from runoff is likely to be equivalent to or perhaps less than agricultural crop sites which presumably might have more bare soil surfaces. Danger of off target drift and to some extent runoff (grasslands) to aquatic habitats may be reduced by foliar intercept in some cases. However, without precautions such as buffer zones to protect bogs, potholes, streams, marshes and other aquatic habitats common to these areas it is assumed that direct drift contamination to these habitats may occur with detrimental effects to aquatic vertebrates and invertebrates. Protection of beneficial or endangered insects with such applications would appear to be impossible. It is therefore assumed that acute risk to non-target insects will occur.

### **Spatial Distribution of Potentially Effected Habitats and Species Groups**

#### Terrestrial Wildlife Utilization of Major Malathion Usage Areas

The following summary of potential major exposure areas for malathion usage is based on EPA Quantitative Usage Analysis data prepared in 1997. Maximum usage estimates were used to allow for potential shifts in market usage of malathion products. Species expected in various crop scenarios were drawn from Wildlife Utilization of Croplands, Gusey, William F. And Z. Maturgo, 1973. The purpose of this portion of the document is not to categorize every species type that could conceivably be exposed to the vast number of potential malathion use sites, but instead to provide a general overview of the species types which might be present for crop and non-crop use sites and to categorize which areas of the country (where possible to predict) may be most heavily impacted by the type of use pattern. Aquatic species are too numerous to list so habitat types common to use sites were listed instead.

**Table 39. Terrestrial Wildlife Utilization of Major Malathion Usage Areas**

Crop Group	Maximum Usage(acres)	Major States for Malathion Usage	Species Common to Usage Locations
Berry Crops (Blueberry, blackberry, strawberry, etc)	70,000	OR, MI, NJ, WA, CA	waterfowl, quail, pheasant, crows, blackbirds, songbirds(finches, robins, starlings, cedar waxwing), grouse, rabbits, deer, racoon, woodchuck, skunk, opossum,
Citrus Crops	14,000	FL, CA, AZ	doves, roadrunner, screech and horned owls, hummingbirds, gilded flicker, laderbacked woodpecker, western kingbird, verdin, cactus wren, mockingbird, thrashers, orioles, cardinals, grosbeaks, goldfinch, linnet, deer, raccoon
Pome Fruits(apple, pear), Avocado, Figs, Grapes	102,000	WA, MO, MI, TX, GA, CO, CA, TN, FL, MS, OH, AZ	grouse, pheasant, songbirds(bluebird, cardinal, catbird, flicker, blue jay, kingbird, magpie, mockingbird, phainopepla, robin, fox sparrow, thrashers, thrushes, vireos, cedar waxwing, woodpeckers), hawks, bear, fox, marmot, porcupine, rabbit, deer, quail, flicker, racoon, opossum, partridge
Stonefruits (apricots, cherries, peach, nectarine)	64,000	OR, WA, GA, TX, AL, MS, MO, CA, AZ	doves, songbirds(blackbirds, grosbeaks, cedar waxwings, robins, starlings western tanager, brown thrasher, titmouse, orioles, jays, finches, etc), pheasant, wild turkey, rabbit, deer, fox, opossum, raccoon, squirrel,
Nut Trees	57,000	CA, TX, LA, GA, OK	
Bulb vegetables (onion, garlic, etc)	37,000	CA, UT, MI, ID, GA	pheasant, rabbit, deer, songbirds, dove
Leafy, Legume, Tuber and Root, Fruiting, Cucurbit and Other Vegetables	315,000	CA, TX, AL, MI, FL, OH, NY, IL, AZ, MS, MO, MN, WI, ID, IN, WA, OR, VA, NC, WV, UT, NJ, GA	turkey, California, scaled, valley, and bobwhite quails, songbirds(buntings, larks, pidgeon, sparrows, roadrunner, grosbeak, ground doves, pipits), shorebirds, coots, ducks, geese, crows, doves, sandhill crane, prairie chicken, partridge, owls and hawks(feeding on field rodents), coyote, muskrat, gray squirrel, groundhog, elk, skunk, rabbits, raccoon, opossum, woodchuck, deer,
Cereal Grain Crops (barley, corn, rice, wheat, sorghum, oats, rye, rice)	697,000	GA, CO, TX, AZ, KY, VA, MN, MT, NC, ND, CA, NY, NC, PA, TX, AR, MS, LA, KS, MO, NE, SD, TN, OK,	rabbits, pheasant, pigeon, doves, ducks( black, canvasback, mallard, pintail, ringnecked, shoveler, teal, wood), coots, rails, egrets, herons, ibis, and gallinules(rice fields), geese, swan, songbirds( blackbirds, towhees, thrasher, sparrows, junco, magpie, snow buntings, grosbeaks, jays, cardinal, bobolink, meadow and horned lark), woodpeckers (eat seeds), ravens, grackles, crows, partridge, grouse, scaled and bobwhite quail, sandhill crane, Attwater prairie chicken(TX), deer, elk, antelope, wild turkeys, gray, fox and ground squirrel, woodchuck, fox, porcupine, coyote, moles, whitefooted and pocket mice, kangaroo rat, muskrat, javelina(TX)
Cotton + USDA Bollweevil	796,000	TX	deer, turkey, squirrel, rabbit, quail, dove, pheasant, prairie chicken, raccoon, opossum, sandhill crane, antelope

Grass and Non-grass Forage Crops (alfalfa, clover, hay)	605,000	CA, ID, MT, OK, AZ, KS, TX, MO, SD, KY	pheasant, mourning dove, partridge, quails, ducks, Canada geese, elk, deer, antelope, grouses, prairie chickens, rabbits, turkey, songbirds, cranes, skunk, small mammals, marmot, ground squirrels,
Hops		OR, WA	pheasant, quail, songbirds, doves, owls and hawks feeding on small mammals
Mint	31,000	IN, WI, OR, WA(90%)	pheasant, quail, doves, songbirds, partridge
Pasture lands	47,000	LA, MO, FL, GA, TX, MS	field and vesper sparrows, bobolink, meadow and horned lark, goldfinch, swallows, pipit, cowbird, red polls, juncos, longspurs, blackbirds, crows, nighthawk, whippoorwill, yellow, palm and prairie warblers, grackles, flickers, bluebirds, and indigo bunting.
Private Lots/Farmsteads	66,000	FL, CA, SD, AL, OK, KS (60%?)	No definitive state surveys were reviewed.
Set Aside Acreage	665,000	MT, MN =(90%?)	No definitive state surveys were reviewed
Summer Fallow	893,000	MT, TX =(100%)	No definitive state surveys were reviewed
Rangeland	20,000	TX, FL, CO =(85%)	No definitive state surveys were reviewed
Woodlands	17,000	AL, LA, TN=(81%)	No definitive state surveys were reviewed
<b>NON Agricultural</b>			
Roadways and fencerows	17,000	Nationwide	sparrows, kingbirds, flycatchers, yellowbreasted chat, indigo bunting, bluebird, goldfinch, brown thrasher, catbird, robin, woodpeckers, yellow and palm warblers, and vireos.
Golf Courses	>12,000	Nationwide	Waterfowl including snow and Canada geese (may feed on treated turf), squirrels and other small mammals(in rough areas), ground feeding songbirds, ie robins,
Nurseries	175,000	Nationwide	
Parks	67,000	Nationwide	Many types of songbirds, small and large mammals,
Landscape Contractors-Bldg Perimeters/Grounds	No estimates	Nationwide-Urban	Songbirds
Cemeteries	21,000	Nationwide -Urban	Same as parks
Mosquito Control	8,227,000	Nationwide near population centers, particularly those surrounded by static aquatic settings ie beach resorts, lake shore communities, low water or flood prone areas	Saltmarshes: Bank and tree swallow, fish and common crow, savanna, seaside, and Sharp tailed sparrow, redwing blackbird, horned lark, egrets, rails, shorebirds, gulls, herons, gallinules, other waterfowl, muskrat, otter,  Freshwater marshes and wet woodlands: marsh, winter and Carolina wrens, swamp, Savanna, sharp-tailed sparrows, swallows, water thrush, ovenbird, phoebe, wood pewee, veery, bluegray gnatcatcher, yellow breasted chat, warblers(hooded, yellowthroat, blackcapped, and Wilson's), racoon, muskrat, beaver,

Proposed for Revocation of Uses Soybean, Peanut, Sunflower	226,000	GA, OK, NC, FL, TX, TN, MN, MO, IN, AR, KS, SD, ND	Species groups not categorized due to impending revocation of use on these crops
---	---------	---	--

Aquatic Organisms: Utilization of Habitats Exposed to Malathion Usage

Numerous types of agricultural uses of malathion may border valuable aquatic habitats such as streams, rivers, lakes, and freshwater marshes. Many of these tributaries may drain to estuarine areas. A few of the crop uses may actually involve sites which border estuarine areas (ie citrus). In some cases, irrigation canals near crop sites will contain fish and shrimp populations and also drain to natural water sources. In general, malathion incidents have involved pulse loading of malathion to streams and ponds following heavy rainfall events or aerial spraydrift of residues directly to the surface of standing water bodies. Residue detection in sediments has been rare. In urban scenarios, storm water runoff has provided a point-source type of residue contribution to streams which drain these areas. Malathion poisonings of aquatic organisms are most likely to occur in the early hours of the exposure period immediately after rainfall or spray applications to specific sites. The numbers of species potentially effected is large and the types of habitat exposures quite varied. The following table provides a very general overview of the types of aquatic habitats that are expected to be exposed from various uses of malathion.

<b>Table 40. Aquatic Habitats - Use Associations</b>			
<b>Crop Group</b>	<b>Maximum Use(acres)</b>	<b>Major States for Malathion Usage</b>	<b>Habitats Common to Usage Locations</b>
Berry Crops (Blueberry, blackberry, strawberry, etc)	70,000	OR,MI,NJ,WA,CA	FW Marshes, ponds, and streams:
Citrus Crops	14,000	FL, CA, AZ	Irrigation canals, rivers, freshwater springs, some estuaries
Pome Fruits(apple, pear), Avocado, Figs, Grapes	102,000	WA, MO, MI, TX, GA, CO, CA, TN, FL, MS, OH, AZ	FW streams, rivers, ponds, marshes, and lakeshore
Stonefruits (apricots, cherries, peach, nectarine)	64,000	OR, WA, GA, TX, AL, MS, MO, CA, AZ	FW streams, rivers, ponds and marshes
Nut Trees	57,000	CA, TX, LA, GA, OK	Streams, irrigation canals, and rivers
Bulb vegetables (onion, garlic, etc)	37,000	CA, UT, MI, ID, GA	Streams, rivers, bogs
Leafy, Legume, Tuber and Root, Fruiting, Cucurbit and Other Vegetables	315,000	CA, TX, AL, MI, FL, OH, NY, IL, AZ, MS, MO, MN, WI, ID, IN, WA, OR, VA, NC, WV, UT, NJ, GA	Irrigation canals, streams, rivers, bogs, marshes
Cereal Grain Crops (barley, corn, rice, wheat, sorghum, oats, rye, rice)	697,000	GA, CO, TX, AZ, KY, VA, MN, MT, NC, ND, CA, NY, NC, PA, TX, AR, MS, LA, KS, MO, NE, SD, TN, OK,	Streams, rivers, ponds, prairie potholes, marshes, saltmarshes, estuarine bays

Cotton + USDA Bollweevil	796,000	TX	Rivers, streams, possibly marshes
Grass and Non-grass Forage Crops (alfalfa, clover, hay)	605,000	CA, ID, MT, OK, AZ, KS, TX, MO, SD, KY	Ponds, bogs, marshes, streams, prairie potholes
Hops		OR, WA	rivers and streams
Mint	31,000	IN, WI, OR, WA(90%)	Streams
Pasture lands	47,000	LA, MO, FL, GA, TX, MS	Streams, rivers, ponds, prairie potholes, marshes, swamps
Private Lots/Farmsteads	66,000	FL, CA, SD, AL, OK, KS (60%?)	Streams, ponds, bogs, potholes, FW springs
Set Aside Acreage	665,000	MT, MN =(90%?)	Streams, ponds, lakes, marshes and potholes
Summer Fallow	893,000	MT, TX = (100%)	Streams, rivers, and potholes
Rangeland	20,000	TX, FL, CO =(85%)	Streams, rivers, swamps, FW springs
Woodlands	17,000	AL, LA, TN =(81%)	Streams, bogs, rivers, wooded wetlands
<b>NON Agricultural</b>			
Roadways and fencerows	17,000	Nationwide	Drainage ditches, crossing or adjacent streams and rivers, swamps, saltmarshes, ponds
Golf Courses	>12,000	Nationwide	Ponds, streams, marshes, some saltmarsh areas
Nurseries	175,000	Nationwide	Ponds, drainage areas to streams
Parks	67,000	Nationwide	Streams, ponds, and lakes(inland) , saltmarshes and ocean shorelines(coastal)
Landscape Contractors-Bldg Perimeters/Grounds	No estimates	Nationwide-Urban areas	Retention and natural ponds, streams, drainage from storm sewers to tributaries
Cemeteries	21,000	Nationwide-Urban areas	Ponds and streams
Mosquito Control	8,227,000	Nationwide near population centers, particularly those surrounded by static aquatic settings ie beach resorts, lake shore communities, low water or flood prone areas	Saltmarsh mosquito control-saltmarshes, estuarine bays, mangrove swamps, shoreline areas  Freshwater mosquito control: freshwater marshes, bogs, and wet woodlands:(Inland areas near population centers
Proposed for Revocation of Uses Soybean, Peanut, Sunflower	226,000	GA, OK, NC, FL, TX, TN, MN, MO, IN, AR, KS, SD, ND	

### Adequacy of Malathion Toxicity Data

The toxicological data, though extensive for malathion, is not complete in several key areas. In addition, much of the data is over twenty years old, and, to some extent, was not conducted in accordance with stricter standards which are required of studies presently submitted to support registration of pesticides. One example would be the fact that most of the acute toxicity endpoints for aquatic organisms are based on nominal concentrations which, due to malathion's short aquatic persistence, may not be appropriate since this could lead to calculated LC50 values which are higher than would have been estimated if based on mean measured concentrations.

There are also some other areas where the data set is weak. In formulation testing only one presently employed product formulation (57% EC) was tested on 4 species (daphnid, oyster, honeybee, and sheepshead minnow). There are no submitted toxicity data on the mixture of malathion and methoxychlor, a possibly highly lethal cocktail for aquatic life. There are no studies regarding the chronic effect levels of malathion to estuarine fish or invertebrates which could conceivably be exposed to repeated pulse load exposures for such uses as citrus and cotton. Further data to elucidate potential effects to non-target insect populations is needed. Acute studies with honeybees indicate that acute contact with direct or latent residues may prove lethal for several days after application. Other beneficial insect populations may also suffer acute losses. There is some indication that amphibian life cycles could be effected by malathion exposure. Though not presently a data requirement requested by the Agency, but given what is known about acute and chronic effect levels observed in frogs, a better understanding of effects to this taxa is needed to improve this assessment.

Sublethal effects caused by temporary disruption of nervous system functions are difficult to use in present risk assessment procedures, because so little is known about their ultimate effect on non-target species populations. However, malathion has been shown to disrupt nesting success in sharp tailed grouse, loss of ability of laboratory mice to navigate a maze, and loss of swimming ability for fish swimming against a current. All of these effects theoretically could lead to reduced survival of certain species groups, when combined with the normal stress factors associated with survival (eg. successful rearing of young, escape from predators, and navigation to spawning grounds).

#### **Limitations of Monitored Effects**

Though a large number of incidents associated with adverse effects to aquatic vertebrates near malathion use sites have been reported, very little information regarding effects to invertebrate populations in the same sites was provided. Given the lower toxicity thresholds for invertebrates exposed to malathion, it is expected that lethal effects to these populations are now occurring from present uses, but, due to the difficulty in observing these effects, go unreported. Many of the monitored residue levels in aquatic habitats near malathion use sites have far exceeded 0.5 ppb which is considered a toxicity threshold for acute effects to aquatic invertebrates. Chronic effects were observed in laboratory studies on daphnids at concentrations which are considered the limitation of detection in field monitoring studies (0.1 ppb). Another consideration is that many of the adverse effects reported for malathion are not investigated within the first 48 hours of exposure, thus allowing substantial degradation of the initial peak concentrations which caused the acute reaction observed in the effected organisms.

#### **Characterization of Predicted Effects to Nontarget Species from Malathion**



### **Ecological Risk to Birds and Mammals**

Based on estimated risk quotients for dietary exposure scenarios malathion is not expected to offer significant acute hazard to birds even at the proposed maximum application scenarios of 6.25 lbs ai/acre on citrus.

Sublethal effects to birds (reduced AChE levels) will, in all probability, result from exposure to malathion residues. The effects may not in themselves prove lethal, but the ultimate result may prove to be reduced survival when exposed birds are subjected to other stress factors in the environment. When radio-tagged sharp-tailed grouse were sublethally dosed with dieldrin or malathion and released back into the wild significant reductions in ability to nest, reproduce and possibly escape predators were observed up to 12 days after dosing (McEwen and Brown, 1966). Control birds all survived and reproduced successfully. In field exposures of birds to malathion applications singing activity was reduced or ceased for up to 2 days following the application.

Chronic exposure for birds presents another matter. In general, malathion is not deemed to be a persistent compound. However, because of the fact that there are no clear restrictions on most of the present labels regarding numbers of consecutive applications, intervals, or avoidance of nesting birds it is conceivable that birds may be subjected to repeated peak levels within very short time intervals. The chronic effects to egg hatch and viability were observed in bobwhite quail at 350 ppm dietary levels. The NOEC for this study was 110 ppm. This threshold would be crossed at application rates above 1.0 lb ai/acre, particularly with short intervals between applications. As this type of effect is usually observed early in the study it might be surmised that the effects in the field would result from early initial exposures to malathion as opposed to growth effects which might require a longer exposure period.

Acute and chronic reproductive effects to mammals are not expected at the proposed tolerance rates. Sublethal effects to nervous system functions caused by acetylcholinesterase blockage may lead indirectly to reduced survival. In studies where test rats were exposed to malathion, reduced ability to navigate a maze was observed (Desi, 1976). This could be serious if a small mammal's ability to relocate its shelter left it exposed to predators or unable to return to its young.

### **Risk to Invertebrates**

The modeling results and field monitored residues indicate that aquatic acute high risk, restricted use, and endangered species levels of concern are exceeded by 8 to 80 times for freshwater and marine invertebrates at application rates at 0.175 lb ai/A, the lowest rate labeled for malathion. For the higher rates the acute risk LOC's are exceeded by over 400 times. The chronic level of concern is far exceeded at all application rates for malathion. Monitored levels of malathion have frequently (though not always) been observed at concentrations which would far exceed the 0.5 ppb level of concern for acute toxicity to invertebrates. In Florida, monitored background levels in urban ponds sometimes exceeded this level of concern **before** aerial applications for medfly control were made. During 1994-95 Medfly spraying efforts in Ventura County, California samples were taken from streams in the spray area after rainfall events and subsequently used in toxicity studies with the freshwater cladoceran, *Ceriodaphnia dubia* and the estuarine mysid, *Neomysis mercedis* (Fujimura, 1995-see summary in appendices). Samples taken during a storm event proved 100% toxic to all exposed test organisms within 2 to 24 hours. These results indicate that concerns for invertebrate survival in exposed urban streams and estuaries are warranted. Monitoring programs related to bollweevil eradication efforts in southeastern states have also yielded residue levels which would be considered to offer acute risk

to invertebrates. In general, levels monitored in agricultural settings appear to be lower than in urban settings and therefore exceedances may be less severe and less frequent under agricultural scenarios. However, predicted EEC's still indicate potential hazard to invertebrates from most crop uses from spraydrift (when applicable) or runoff.

### **Risk for Fish**

Risk quotients indicating levels of concern for acute risk to fishes, restricted use, and endangered species are exceeded for freshwater and estuarine fish at registered application rates of 0.5 lb ai/A, <0.175 lb ai/A, and <0.175 lb ai/A, respectively. Based on monitored residues, this will prove more likely if no protective restrictions are employed. The labels presently do not include actual protective methods (eg. buffer zones) for prevention of drift to aquatic habitats. Due to malathion's low persistence characteristics in water, chronic exposure risk for fish is less likely for single applications. Repeated applications could lead to continual exposure to peaks within one week periods, allowing for mean levels to remain above the chronic threshold of 2 ppb for early life stage effects. In actual uses of malathion (both urban and agricultural) many fish kills have been reported and confirmed. These incidents generally involve drift from aerial applications to small ponds, inland lakes, and rivers. In most cases the residues have not remained at high levels following these fish kills, indicating that fish are severely effected early in the exposure period. Fish kills resulting from runoff have also occurred several days after applications have been completed. These kills generally involve concentration of residues from a watershed into small feeder tributaries or stormwater feeder pipes which then open into retention ponds or farmponds within the drainage basin. Effects to estuarine fish have generally involved shallow lagoons or tidal waters at low tide following mosquito control uses.

### **Risk for Amphibians**

Routes of exposure for amphibians are expected to be through direct contribution of residues to aquatic habitats where adults or their offspring reside or through dermal adsorption from spraydrift to terrestrial areas where they might reside. Based on risk criteria for fish ( $\frac{1}{2}$  the LC50 = Level of Concern) risk to tadpoles of sensitive frog species will occur with aquatic EEC's of 100 ppb. This could occur from direct drift of less than 0.5 lbs ai/acre, or, using a worst case GENEEC scenario, runoff and/or drift from an approximate 2.0 lb ai/acre application of malathion. EFED has limited information on possible effects to amphibians from dermal adsorption of residues. In actual reports of adverse effects to aquatic organisms mortality of adult amphibians (usually frogs) has been reported as well as presence of malathion residues in tissues following non-lethal exposures. These adverse effects generally involve malathion contamination of shallow wetland areas where flush rates are slow. Exposure of aquatic eggs or larvae of amphibians to malathion residues in surrounding water is also a potential route of exposure which could lead to adverse effects to developmental stages of amphibians.

### **Risk for Reptiles**

Acute risk for adult reptiles is not expected from most malathion uses. Oral ingestion or dermal adsorption of residue laden water might be the most likely route of exposure for aquatic reptiles. In several of the reported fish kills for malathion, adverse effects to aquatic turtles was also observed. However, confirmation that the turtles were killed by malathion alone is not provided. Effects to developing eggs of reptiles from direct exposure is also of concern when malathion uses provide potential exposure to shoreline nesting sites.

### **Risk to Nontarget Plants**

Malathion is not expected to pose a serious hazard to terrestrial plants or aquatic algae as the mode of action (effects to nervous system) would not apply to plants. Malathion is expected to be systemically absorbed into plant tissues based on field study analyses of plant tissues after malathion applications. The Agency has received no reports of adverse reactions of crops or plants to malathion itself though label advisories for forest use do caution against application to certain species of trees.

### **Risk to Non-Target Insects**

Malathion has been shown to be lethal to many species of beneficial insects at rates routinely employed in agricultural settings. The routes of exposure may be direct contact, contact with foliar residues, and contact with residue coated pollen transported back to nests or hives. Aquatic larvae of terrestrial species may also be acutely effected for limited time periods through residue drift or runoff to streams or other aquatic habitats. In Giles' review of effects of malathion application to a hardwood forest (see previous summary) the author made a pertinent summary of the predictability of what may occur to insect populations. "Effects will range from none to near complete extermination of species on the area. Insecticidal effects on certain populations may be obscured by drastic predator-prey-host-parasite shifts caused by the insecticide. The resistance of natural populations and the immediate recharge and stabilization of populations will obscure effects of insecticides. Egg and larval stages, unmeasured by sampling techniques may be affected, the results of which may be postponed or may remain unrecognized. Aquatic populations may be affected with subsequent effects on insect eggs, larvae and later, adults. The result is a multidimensional web of action and interaction between and within species and their natural environment and an unnatural environmental hazard, malathion insecticide." This same summary may also be applicable to malathion effects when used near or over other non-agricultural areas containing beneficial insect populations, such as salt marshes, riverbanks, meadows and natural grasslands.

## References Environmental Fate

- Ali Fouad, A.F. and Fukuto, T.R. (1982). Toxicity of O,O,S-trialkyl phosphorothioates to the rat. *J Agric Food Chem* **30**, 126-130.
- Aldridge, W.N., Miles, J.W., Mount, D.L., and Verschoyle, R.D. (1979). The toxicological properties of impurities in malathion. *Arch Toxicol* **42**. 95-106.
- Bourquin, A.W. (1977). Effects of malathion on microorganisms of an artificial salt-marsh environment. *J. Environ. Qual.* **4**. 373-378.
- Chukwudebe, A., March, R.B., Othman, M., and Fukuto, T.R. (1989). Formation of trialkyl phosphorothioate esters from organophosphorus insecticides after exposure to either ultraviolet light or sunlight. *J Agric Food Chem.* **37**. 539-545.
- Eichelberger, J.W. and Lichtenberg, J.J. (1971). Persistence of pesticides in river water. *EnvironSci Technol* **5**. 541.
- Fukuto, T.R. (1983). Toxicological properties of trialkyl phosphorothioate and dialkyl alkyl- and arylphosphonothioate esters. *J Environ Sci Health* **B18**. 89-117.
- Guerrant, G.O., Fetzer, L.E. Jr., and Miles, J.W. (1970). Pesticide residues in Hale County, Texas, before and after ultra-low volume aerial application of malathion. *Pesticides Monitoring Journal* **4**. 14-20.
- Handbook of Environmental Fate and Exposure Data for Organic Chemicals*<sup>1</sup> volume 3. P.H. Howard, Ed., Lewis Publishers. 1991.
- Howard, P.H. ed. *Handbook of Environmental Fate and Exposure Data for Organic Chemicals* Volume 3. Lewis Publishers Chelsea Michigan. 1991.
- Kearney, P.C., Plimmer, J.R., and Helling, C.S. 1969. Encyc. Chem. Technol. **18**. 515 as reported in Matsumura, Fumio. *Toxicology of Insecticides*. Second Edition. Plenum Press, New York. 1985.
- Lichtenstein, E.P. and Schulz, K.R. (1964). The effects of moisture and microorganisms on the persistence and metabolism of some organophosphorus insecticides in soil with special emphasis on parathion. *J Econ Entomol* **57**, 618.
- Matsumura, Fumio. Toxicology of Insecticides. Second Edition. Plenum Press, New York. 1985.
- Miles, C.J. and Takashima, S., 1991. Fate of malathion and O,O,S-trimethyl phosphorothioate by-product in Hawaiian soil and water. *Arch Env Contam and Toxicol.* **20**. 325-329.
-

- Mulla, M.S., Mian, L.S., and Kawecki, J.A. (1981). "Distribution, transport, and fate of the insecticides malathion and parathion in the environment" in *Residue Reviews* volume 81. Gunther, F.A. and Gunther, J.D. eds., Springer-Verlag New York.
- Muhlman, V.R. and Shrader, G. (1957). Hydrolyse der insektiziden phosphorsauresster. *Z. Naturf.* 12. 196-208.
- Paschal, D.C., and M.E. Neville. (1976) Chemical and microbial degradation of malaoxon in an Illinois soil. *J. Environ. Qual.* 5:441-443.
- Penn State. Study of Off-Site Deposition of Malathion Using Operational Procedures for the Southeastern Cotton Boll Weevil Eradication Program. Aerial Application Technology Laboratory. Department of Entomology. December 1993.
- Roberts, J.E., Chisholm, R.D., and Koblitsky, L. (1962). Persistence of insecticides in soil and their effects on cotton in Georgia. *J Econ Entomol* 55. 153.
- State of California Environmental Protection Agency. A Characterization of Sequential Aerial Malathion Applications in the Santa Clara Valley of California. California Department of Food and Agriculture (presently CalEPA). Division of Pest Management, Environmental Protection and Worker Safety. EH-82-01. 1981.
- State of California Environmental Protection Agency. Environmental Monitoring Results of the Mediterranean Fruit Fly Eradication Program, Riverside County 1994. Department of Pesticide Regulation. Environmental Hazards Assessment Program. September 1996.
- State of California Environmental Protection Agency. Environmental Monitoring Results of the Mediterranean Fruit Fly Eradication Program, Riverside County 1994. Department of Pesticide Regulation. Environmental Hazards Assessment Program. September 1996.
- State of California Environmental Protection Agency. Assessment of Malathion and Malaoxon Concentrations and Persistence in Water, Sand, Soil and Plant Matrices Under Controlled Exposure Conditions. Department of Pesticide Regulation. Environmental Hazards Assessment Program. (Report EH 93-03) February 1993.
- Toia, R.F., March, R.B. Umetsu, N., Mallipudi, N.M., Allahyari, R., and Fukuto, T.R. (1980). Identification and toxicological evaluation of impurities in technical malathion and fenthion. *J Agric Food Chem* 28. 599-604.
- U.S. Department of Agriculture. National Boll Weevil Cooperative Control Program: Final Environmental Impact Statement-1991 volume 1. Animal Plant Health Inspection Service. 1991.
- U.S. Department of Agriculture. Environmental Monitoring Report: Boll Weevil Cooperative Eradication Program: Texas Lower Rio Grande Valley. Animal Plant Health Inspection Service. 1995.
- U.S. Department of Agriculture. Environmental Monitoring Report: Southern Rolling Plains Boll Weevil Eradication Program Animal Plant Health Inspection Service. 1994-1995.

U.S. Department of Agriculture. Environmental Monitoring Report: Texas High Plains Boll Weevil Diapause Control Program. Animal Plant Health Inspection Service. 1995.

U.S. Department of Agriculture. Environmental Monitoring Report: Southeast Boll Weevil Eradication Program. Animal Plant Health Inspection Service. 1993.

U.S. Department of Agriculture. Environmental Monitoring Report: Southeast Boll Weevil Eradication Program. Animal Plant Health Inspection Service. 1996.

U.S. Department of Agriculture. Environmental Monitoring Report: Southeast Boll Weevil Eradication Program Sensitive Sites. Animal Plant Health Inspection Service. 1997.

U.S. Department of Agriculture. Environmental Monitoring Report: Cooperative Medfly Project Florida. Animal Plant Health Inspection Service. 1997.

U.S. Environmental Protection Agency. Pesticides in Ground Water Database - A Compilation of Monitoring Studies:1971- 1991. Office of Prevention, Pesticides, and Toxic Substances, EPA 734-12-92-001, September 1992.

U.S. Environmental Protection Agency. GENEEC: A Screening Model for Pesticide Environmental Exposure Assessment. The International Symposium on Water Quality Monitoring, April 2-5 1995. American Society of Agricultural Engineers. p 485. 1995.

U.S. Environmental Protection Agency. PRZM-3, A Model for Predicting Pesticide and Nitrogen Fate in the Crop Root and Unsaturated Soil Zones: Users Manual for Release 3.0. R.F. Carsel, J.C. Imhoff, P.R. Hummel, J.M. Cheplick, and A.S. Donigian, Jr. National Exposure Research Laboratory, U.S. Environmental Protection Agency, Athens, GA 30605-2720. AQUA TERRA Consultants, Mountain View, CA 94043. Waterborne Environmental, Inc. Leesburg, VA 22075. 1997.

U.S. Environmental Protection Agency. EXPOSURE ANALYSIS MODELING SYSTEM (EXAMS II) User's Guide for version 97.2 Lawrence A. Burns, Ph.D., Research Ecologist, Ecosystems Research Division, National Exposure Research Laboratory, Athens, Georgia. 1997.

U.S. Geological Survey. Pesticides in Surface and Ground Water of the United States: Preliminary Results of the National Water Quality Assessment Program. Pesticides National Synthesis Project. National Water Quality Assessment. 1997.

Umetsu N., Grose, F.H., Allahyari, R., Abu-El-Haj, S., Fukuto, T.R. (1977). Effect of impurities on the mammalian toxicity of technical malathion and acephate. *J Agric Food Chem* **25**. 946-955.

Wang, T.C., Lenahan, R.A., and Tucker, J.W. 1987. Deposition and persistence of aerially-applied fenthion in a Florida estuary. *Bull Environ Contam Toxicol.* **38**. 226-231

Willis and McDowell, 1987. Pesticide Persistence on Foliage. *Reviews of Environmental Contamination and Toxicology*, Vol. 100.

## REFERENCES: Ecological Effects

- Bender, Michael E., 1969. The Toxicity of the Hydrolysis and Breakdown Products of Malathion to Fathead minnow - University of Michigan.
- Bender, Michael E. 1969. Uptake and Retention of Malathion by the Carp. University of Michigan.
- Bender, Michael E. 1976. The Toxicity of Malathion and its Hydrolysis Products to Eastern Mudminnow *Umbra pygmaea*, Virginia Institute of Marine Science.
- Beyers, P. & P.Sikoski, 1993. Acetylcholinesterase Inhibition in Federally Endangered Colorado Squawfish exposed to Carbaryl and Malathion.
- Beyers, P. 1993. Acetylcholinesterase Inhibition in Federally Endangered Bonytail Chub Exposed to Carbaryl and Malathion.
- Bookhout, Cazlyn G. and John D. Costlow Jr., 1976. Effects of Mirex, Methoxychlor and Malathion on Development of Crabs., Duke University and Gulfbreeze Laboratory (USEPA) Pg. 53-69.
- Bourke, J.B. et al, 1968. Comparative Metabolism of Malathion - C<sub>14</sub> in Plants and Animals. New York State Agricultural Experiment Station, Cornell University.
- Bourquin, Al W. 1975. Microbial-Malathion Interaction in Artificial Saltmarsh Ecosystems. Gulfbreeze Laboratory, USEPA.
- California-Administrative Report 82-2, 1982. Monitored aquatic incidents during broadscale aerial application over San Francisco, Bay area, 1981., Dept. Of Fish and Game, Environmental Services Branch.
- Cook, Gary H. and James C. Moore, 1976. Determination of Malathion, Malaoxon, and Mono- and Dicarboxylic Acids of Malathion in Fish, Oyster, and Shrimp Tissue. USEPA Gulfbreeze Laboratory.
- Conte, Fred S. and Jack C. Parker, 1975. Effect of Aerially Applied Malathion on Juvenile Brown and White Shrimp, *Penaeus aztecus* and *Penaeus setiferus*. - Texas A&M University (Am. Fisheries Society).
- Coppage, D et al 1975. Brain Acetylcholinesterase Inhibition in fish as a Diagnosis of Environmental Poisoning by Malathion. Gulfbreeze Environmental Research Lab.
- Coppage, D.L. and T. W. Duke, 1971. Effects of Pesticides in Estuaries Along the Gulf and Southeast Atlantic Coasts. USEPA, Gulfbreeze Laboratory.
- Dahlsten, Donald L. 1983. Effects of Malathion Bait Spray for Mediterranean Fruit Fly on Non-target Organisms on Urban Trees in Northern California., University of California.

Darsie, Richard and Coraiden, F. Eugene, 1958. Malathion Toxicity to Killifish in Delaware. Delaware Agricultural Research Station.

Desi, I. et al , 1976. Toxicity of Malathion to Mammals, Aquatic Organism and Tissue Culture Cells. Division of Hygienic Biology, Budapest, Hungary.

Dieter, Michael P., 1975 Use of Enzyme Profiles to Monitor Residues in Wildlife. USFWS, Patuxent Wildlife Research Center.

Dunachie and Fletcher, 1969. ( Walker (1971)Khera and Lyon(1968)  
A number of studies were conducted where malathion or malaaxon were injected into chick embryos,). Malaaxon caused reduced survival of embryos at a concentration of 30 micromoles, and those that did survive had severe abnormalities. Malathion at 30 and 15 micromoles produced less severe abnormalities.

Desi, I. et al 1976. Toxicity of Malathion to Mammals, Aquatic Organism and Tissue Culture .

Eaton, John G., 1970. Chronic Toxicity of Malathion to the Bluegill (*Lepomis macrochirus*). Duluth EPA Lab., Duluth Minnesota.

Engbring, John. 1989. Fluctuations in Bird Populations on the Island of Bota as Related to an Experimental Program to Control The Melon Fly. U.S. Fish and Wildlife Service, Honolulu, Hawaii.

Finlayson, B.J., G. Faggella, H. Jong, E. Littrell, and T.Lew , 1981. Impact on Fish and Wildlife From Broadscale Aerial Malathion Applications in South San Francisco Bay Region, , Pesticide Investigation Unit, Water Pollution Control Laboratory, California Fish and Game Department.

Gary, Norman E. and Eric C., Mussen. 1984. Impact of Mediterranean Fruit Fly Malathion Bait Spray on Honeybees. Dept. of Entomology, Univ. of California, Davis.

Giles, S. and Robert H., Jr., 1970. The Ecology of a Small Forested Watershed Treated with the Insecticide Malathion S<sup>35</sup>., Published by the Wildlife Society.

Greenburg, J. and N. Latham, 1968. Malathion - Induced Teratisms in the Developing Chick. University of Ottawa.

Gusey, William F. And Z. Maturgo, 1973. Wildlife Utilization of Croplands. Dept. Of Environmental Affairs, Shell Oil Company.

Hall R.J. and D.R. Clark, 1982. Responses of the Iguanid Lizard, *Anolis caroleninsus* to Four Organophosphorous Pesticides, Environmental Pollution (Ser. A) 28:45-52

Hill, Elwood F. et al., 1971. Effects of Ultra-Low Volume Applications of Malathion in Hale County Texas. Journal of Med Entomology.

Hoff, James and Westman, James, 1965. Dibrom/Malathion Formulation Use as a Piscicide.



- Holland, H.T. and Jack Lowe , 1966. Malathion, Chronic Effects on Estuarine Fish -, Gulfbreeze Biological Laboratory.
- Jensen, Loren D. and Anden R. Gaufin. 1964. Effects of Ten Organic Insecticides on Two Species of Stonefly Naiads. , Dept. Zoology, University of Utah. (MRID 00065497)
- Johansen, C.A. et al. 1965. Bee Poisoning Hazard of Undiluted Malathion Applied to Alfalfa in Bloom. Washington State University College of Agriculture.
- Johnson, C.R. 1977. Effects of Field Applied Rates of Five Organophosphorous Insecticides on Thermal Tolerance, Orientation and Survival of *Gambusia affinis*.
- Keller, Anne E. , 1995. Toxicity of Malathion to Native Freshwater Mussels. National Biological Survey Laboratory, Gainesville, Florida.
- Kennedy, Harry D. and David Walsh. 1970. Effects of Malathion on Two Warmwater Fishes and Aquatic Invertebrates in Ponds. USFWS, Fish Pesticide Research Laboratory, Columbia, Mo.,
- Kucera, Emil. 1987. Brain Cholinesterase Activity in Birds After a City-Wide Aerial Application of Malathion. Manitoba Dept. of Environment and Workplace Safety and Health.
- Kuhajda, B.R. et al 1996. Impact of Malathion on Fish and Aquatic Invertebrate Communities and on Acetylcholinesterase Activity in Fishes in Stewart Creek, Fayette County, Alabama. , Dept. Of Biological Sciences, University of Alabama,
- Marliac and Mutchler, 1963. In other studies where malathion was injected into eggs at 50 mg/egg chicks showed shortening of legs and bleaching of feathers .
- McEwen, Lowell and Robert L. Brown. 1966. Acute Toxicity of Dieldrin and Malathion to Wild Sharp-tailed Grouse. Journal of Wildlife management. Vol. 30, No. 3, July 1966. MRID 113233
- Mehrotra, K. N. et al, 1966. Physiological Effects of Malathion on the House Sparrow *Passer domesticus*. Indian Agricultural Research Institute Delhi, India.
- Mitchell and Yutema, 1973. Teratogenic Effects of Malathion and Captan in the Embryo of Common Snapping Turtle.
- Mount, Donald I. and Charles Stephan, 1967. A Method for Establishing Acceptable Toxicant Limits for Fish - Malathion and Butoxyethanol Ester of 2, 4-D -U.S. Dept of Interior.
- Oshima, R. S. 1982 California Medfly Report.
- Parkhurst, Zell and Harlan Johnson. 1955. Toxicity of Malathion 500 to fall Chinook Salmon Fingerlings. USFWS, (Progressive Fish Culturist) .

Parsons, Jack K. and Billy Don Davis, 1971. The Effects on Quail, Migratory Birds and Non-Game Birds from Application of Malathion and Other Insecticides. Texas Parks and Wildlife Dept., Study conducted from 1964 to 1968.

Post, George and Robert Leasure. , 1974. Sublethal Effects of Malathion to Three Salmonid Species. Colorado State University .

Proctor, Raphael R. Jr., Jane P. Corliss, and Donald Lightner 1966. Mortality of Post larval and Juvenile Shrimp Caused by Exposure to Malathion -A Laboratory and Field Study. National Marine Fisheries Service, Galveston Laboratory.

Rong, Suriyam, Y. et al, 1968. Effects of Insecticides on Feeding Activity of the Guppy, a Mosquito-eating Fish in Thailand - World Health Organization.

Tagatz, M.E., 1974. Effects of Ground Application of Malathion on Saltmarsh Environments In Northwestern Florida. USEPA Gulfbreeze, Environmental Research Laboratory, Gulfbreeze, Florida.

USDA, Environmental Monitoring report. Cooperative Medfly Project Florida, 1997. Spray Operations Hillsborough Area. USDA Report.

Weis, Peddrick and Judith Weiss, 1975. Abnormal Locomotion Associated with Skeletal Malformations in Sheepshead minnow. Rutgers University and New York Ocean Science Laboratory, Montauk, New York.

Walker, W.W. and Stojanovic, B.J., 1974. Malathion degradation by an *Arthrobacter* species. *J. Environ Quality*. 3. 4-10.

Walker, W.W. and Stojanovic, B.J., 1973. Microbial versus chemical degradation of malathion in soil. *J. Environ Quality*. 2. 229-232.

Woodward, Dan F., , 1969. Sport Fisheries Research USFWS, Publication 77.

Author Anonamous, 1967. Quarterly Report - USFWS Research Laboratory, Columbia, Mo..

Author Anonamous, 1970. USFWS Sport Fisheries Research Report Publication 106, Wash. DC.

**References: Use Information**

Biological and Economic Analysis Division, OPPTS/OPP/USEPA. Malathion Quantitative Usage Analysis data prepared in 1997.



## APPENDICES

### APPENDIX 1

#### DATA REQUIREMENTS FOR Malathion

Chemical No:057701

#### 158.490 Wildlife and Aquatic Organisms

Guideline	Data In	MRID(s)	Data Req. Fulfilled
71-1(a) Acute Avian Oral, Quail/Duck	Yes	00160000	Yes
71-2(a) Acute Avian Diet, Quail	Yes	00022923	Yes
71-2(b) Acute Avian Diet, Duck	Yes	00022923	Yes
71-3 Wild Mammal Toxicity	No	Not Required	NA
71-4(a) Avian Reproduction Quail	Yes	43501501	Yes
71-4(b) Avian Reproduction Duck	Yes	42782101	Yes
71-5(a) Simulated Terrestrial Field Study	No	Not Required	NA
71-5(b) Actual Terrestrial Field Study	No	Not Required	NA
72-1(a) Acute Fish Toxicity Bluegill	Yes	40098001	Yes
72-1(b) Acute Fish Toxicity (TEP)	No	Required*(5,6)	No
72-1(c) Acute Fish Toxicity Rainbow Trout	Yes	40098001	Yes
72-1(d) Acute Fish Toxicity Rainbow Trout (TEP)	No	Required*(5,6)	No
72-2(a) Acute Aquatic Invertebrate	Yes	40098001	Yes
72-2(b) Acute Aquatic Invertebrate (TEP)	Yes	41029701	Partially**
72-3(a) Acute Est/Mar Toxicity Fish	Yes	41174301	Yes
72-3(b) Acute Est/Mar Toxicity Mollusk	Yes	40228401	No
72-3(c) Acute Est/Mar Toxicity Shrimp	Yes	41474501	Yes
72-3(d) Acute Est/Mar Toxicity Fish (TEP) **(5,6)	Yes	41252101	Partial
72-3(e) Acute Est/Mar Toxicity Mollusk (TEP)	Yes	42249901	Partial**(5,6)
72-3(f) Acute Est/Mar Toxicity Shrimp (TEP)	Yes	Required*(5,6)	No
72-4(a) Early Life Stage Fish(Freshwater)	Yes	41422401	Yes
72-4 Early Life Stage Estuarine Fish	Yes	Required*(5,6)	No
72-4(b) Life Cycle Aquatic Invertebrate	Yes	41718401	Yes
72-5 Life Cycle Fish	Yes	Reserved	No
72-6 Aquatic Organism Accumulation	Yes		

<b>Guideline</b>	<b>Data In</b>	<b>MRID(s)</b>	<b>Data Req. Fulfilled</b>
72-7(1) Simulated Aquatic Field Study	No	Reserved	No
72-7(b) Actual Aquatic Field Study	No	Reserved	No
<b>§158.540 PLANT PROTECTION</b>			
122-1(a) Seed Germ.,Seedling Enmergence	No	Not required	NA
122-2 Aquatic Plant Growth	No	Not required	NA
122-1(a) Seed Germ./Seedling Emerg.	No	Not required	NA
122-1(b) Vegetative Vigor	No	Not required	NA
123-1(a) Seed Germ./Seedling Emerg.	No	Not required	NA
123-1(b) Vegetative Vigor	No	Not required	NA
123-2 Aquatic Plant Growth	No	Not required	NA
124-1 Terrestrial Field Study	No	Not required	NA
124-2 Aquatic Field Study	No	Not required	NA
<b>§158.490 NONTARGET INSECT TESTING</b>			
141-1 Honey Bee Acute Contact	Yes	05001991,001999, 05004151, 05004003	Yes
141-2 Honey Bee Residue on Foliage	Yes	4120800, 41284701	Yes
141-5 Fuel Test for Pollinators	No	Reserved	No

**FOOTNOTES:**

\*Required to support Malathion/Methoxychlor mixture products

\*\*Though formulation tests on 57EC were submitted the Agency requires formulation toxicity testing of Malathion/Methoxychlor mixture products

1. 1=Terrestrial Food; 2=Terrestrial Feed; 3=Terrestrial Non-Food; 4=Aquatic Food; 5=Aquatic Non-Food(Outdoor);6=Aquatic Non-Food (Industrial);7=Aquatic Non-Food (Residential);8=Greenhouse Food; 9=Greenhouse Non-Food;10= Forestry; 11=Residential Outdoor; 12=Indoor Food; 13=Indoor Non-Fo d; 14=Indoor Medicinal;15=Indoor Residential

## Environmental Fate Data Requirements

	<u>Status</u>	<u>MRID Number</u>
<b><u>Degradation</u></b>		
161-1 Hydrolysis	Fulfilled (RJM, 12/15/92 RJM, this review)	40941201 43166301
161-2 Photo. - water	Fulfilled (RJM, 12/15/92 RJM, this review)	41673001 43166301
161-3 Photo. - soil	Fulfilled (RJM, 12/15/92 RJM, this review)	40658009 41695501 43166301
161-4 Photo. - air	Not required <sup>1</sup> (RJM, 12/15/92 RJM, this review)	40969301 43166301
<b><u>Metabolism</u></b>		
162-1 Aerobic soil	Fulfilled (RJM, 12/15/92 RJM, this review)	41721701 43163301
162-2 Anaerobic soil	Not required	
162-3 Anaerobic aquatic	Fulfilled (RJM, 12/15/92 RJM, this review)	42216301 43163301
162-4 Aerobic aquatic	Unfulfilled (RJM, 12/15/92) (RJM, this review)	42271601 43163301
<b><u>Mobility</u></b>		
163-1 Leaching, Ads./ Desorption	Fulfilled (RJM, 12/15/92) (RJM, this review)	41345201 43163301 43868601
163-2 Volatility-lab	Fulfilled (RJM, 12/15/92)	42015201
163-3 Volatility-field	Not Required <sup>2</sup>	
<b><u>Dissipation</u></b>		
164-1 Soil	Fulfilled (HLM, 7/25/91 RJM, 12/15/92, this review)	41748901 41727701 43042401 43042402 43166301
164-2 Aquatic	Unfulfilled (RJM, 12/15/92)  (RJM, this review)	42058401 42058402 43166301

---

<sup>6</sup>A study was submitted and reviewed but was not acceptable. Based on the laboratory volatility study, volatility does not appear to be an important route of dissipation; therefore, this study is not needed at this time.

164-3 Forest	Not required	
	<b><u>Status</u></b>	<b><u>MRID</u></b>
164-5 Soil, long term	Not required	
<b><u>Accumulation</u></b>		
165-2 Field rotat. crop	Not required	
165-3 Irrigated crop	Fulfilled	
	(RJM, 12/15/92)	42058401
		42058402
	(RJM, this review)	43166301
165-4 Fish	Fulfilled	
	(RJM, this review)	43106401
		43106402
		43340301
<b><u>Spray Drift</u></b>		
201-1 Drop size spec.	Not submitted <sup>3</sup>	
202-1 Drift field eval.	Not submitted	

---

<sup>3</sup> Data is not required at this time pending the results of the spray drift task force.

## APPENDIX 2

### *Tier I Estimated Concentrations for Surface-Water Exposure Assessment*

#### Summary

Based on fate characteristics, model predictions and actual monitoring studies, the Agency predicts malathion will reach drinking water sources from the proposed uses. Surface water concentrations were modeled using the GENEEC model with acute and chronic drinking water levels set with the pesticide use scenarios that produced the highest aqueous pesticide levels. HED has indicated that malathion's degradate, malaoxon, is to be included in the tolerance expression for malathion thus water concentrations are provided in this document for both malathion and, when possible, malaoxon.

Table 1. Expected drinking water concentrations for malathion and malaoxon (Tier I).

compound / exposure type	surface water		ground water	
	estimated concentration (ppb)	source of concentration	estimated concentration (ppb)	source of concentration
malathion / acute	226	GENEEC peak	3.1	Monitoring data
malathion / chronic	21.2	GENEEC 56-day ave.		
malaoxon / acute	96.0	GENEEC peak	3.1	Derived from malathion monitoring data
malaoxon / chronic	75.5	GENEEC 56-day ave.		

EFED recommends that 226 and 96.0 ppb (Table 1) be considered as the highly conservative first tier estimates for acute surface drinking water levels for malathion and malaoxon, respectively. For chronic surface drinking water levels, 21.2 and 75.5 ppb are recommended for malathion and malaoxon, respectively. The chronic malaoxon value exceeds the chronic malathion level because of its longer expected environmental persistence. First tier groundwater concentrations were derived from monitoring data because they were higher than results from the SCIGROW model. The highest detected malathion concentration in groundwater accepted by EFED was 3.1 ppb. Malaoxon was not examined in this study but the same value is expected to be a conservative estimate of malaoxon concentration. EFED recommends exposure estimates of 3.1 ppb for malathion and 3.1 ppb for malaoxon in ground water.

This assessment was conducted under guidelines stated in OPP's Interim Approach for Addressing Drinking Water Exposure (November 1997), however, standard modeling techniques were modified to estimate malaoxon concentrations. Malaoxon levels were estimated with the GENEEC model with the assumption that fate variables which were not known are the same as those for malathion. Acceptable environmental fate studies specifically for malaoxon; including degradation, metabolism, mobility, dissipation, and solubility data; would be very useful for future assessments.

#### Environmental Fate

Based on all the data submitted, EFED concludes that the primary route of dissipation of malathion in surface soils appears to be aerobic soil metabolism. Supplemental data submitted



by the registrant shows malathion degrades in soils with a half-life ( $t_{1/2}$ ) of <1 day on Blackoar loam soil (pH 6.1). For modeling this half-life value was multiplied by a factor of three to estimate a 90th percentile  $t_{1/2}$  value, thus a 3-day half-life was used. This half-life is the same as the value used by USDA in malathion modeling (USDA 1991). EFED notes that longer half-lives (6.9 days) have been reported on sand (CalEPA 1996). Laboratory half-lives for anaerobic aquatic metabolism (<2.5 days) and hydrolysis (6.21 days at pH 7, 12 hours at pH9) indicate that these are also important routes of dissipation. Conversely, the compound is moderately stable to aqueous ( $t_{1/2}$  = 71 and 98 days) and soil photolysis ( $t_{1/2}$  = 173 days) and does not volatilize appreciably ( $\leq 5.1\%$  volatilized after 16 days).

Data presented to the Agency demonstrate that malathion is extremely mobile and thus runoff and leaching may be major routes of dissipation. Acceptable leaching data on parent malathion indicate that it is mobile in all soils tested [ $K_{ds}$  = 0.82-2.47 L/kg,  $K_{oc,s}$  = 151-308 L/kg]. Terrestrial and aquatic field dissipation data indicate rapid dissipation ( $t_{1/2}$  = <2 days). Malathion has been detected in ground water in three states at levels ranging from 0.007 to 6.17 ppb (USEPA 1992). Based on these data, EFED concludes that malathion has the potential to contaminate surface and ground water.

Malathion mono- and dicarboxylic acids, malaoxon, ethyl hydrogen fumarate, diethyl thiosuccinate, and  $CO_2$  are degradates that have been found in malathion laboratory and field studies. Time course studies on malaoxon production on sand and soil have been published (CalEPA 1993) showing levels to increase over time. Maximal measured malaoxon concentration relative to initial malathion concentrations were 1.4% (after ~10 days on sand) and 10.7% (after 21 days on soil). Measurements past 21 days were not made. In the aerobic soil metabolism study submitted by the registrant 1.8% conversion to malaoxon was the maximum level observed on the Blackoar loam soil, thus 10.7% conversion appears to be a conservative conversion value.

EFED does not have a complete environmental fate database for malaoxon but based on its chemical similarity to malathion (sulfur is replaced by oxygen); the parent and its degradate are expected to have similar chemical properties. However, the biological properties of malaoxon are notably different in that it is more toxic than malathion. The aerobic half-life of malaoxon has been reported as 3 and 7 days in basic and acidic soils, respectively (Paschal and Neville 1976). This longer half-life is proposed to be a result of malaoxon's biocidal effect on soil microbes which contribute to malathion's degradation.

### **Surface Water Assessment**

GENEEC (USEPA 1995) is a screening model designed by the Environmental Fate and Effects Division (EFED) to estimate the concentrations found in surface water for use in ecological risk assessment. As such, it provides upper-bound values on the concentrations that might be found in ecologically sensitive environments because of the use of a pesticide. It was designed to be simple and require data which is typically available early in the pesticide registration process. GENEEC is a single event model (one runoff event), but can account for spray drift from multiple applications. GENEEC is hardwired to represent a 10-hectare field immediately adjacent to a 1-hectare pond that is 2 meters deep with no outlet. The pond receives a spray drift event from each application plus one runoff event. The runoff event moves a maximum of 10% of the applied pesticide into the pond. This amount can be reduced due to degradation on the

field and the effects of soil binding in the field. Spray drift is equal to 1 and 5% of the applied rate for ground and aerial spray application, respectively.

Standard GENEEC modeling is inappropriate for malaoxon concentrations because the model assumes initial concentrations are highest which is not the case with malaoxon which increases over a period of weeks. In this case EFED chose a conservative scenario for modeling malaoxon concentrations by assuming that 10.7% of each malathion application is converted to malaoxon, thus for the purposes of GENEEC malaoxon was applied at 10.7% the rate of malathion. Data used for modeling were not ideal. The physical parameters used for malaoxon were those of malathion based on their chemical similarity. For the purpose of modeling EFED has attempted to estimate the upper 90<sup>th</sup> percentile of malaoxon's aerobic soil half-life value by multiplying 7 days (Paschal and Neville 1976) by a factor of three resulting in the model input value of 21 days. The aqueous half-life used was 107 days (based on malathion hydrolysis at pH 5), respectively. Both half-lives are expected to be conservative. The hydrolysis data for malathion is expected to be similar to malaoxon and is used in the absence of a half-life value in water with microbial activity.

Modeling results indicate that malathion and malaoxon have the potential to move into surface waters. Based on the inputs shown in Table 2 the peak GENEEC estimated environmental concentrations (EEC) of malathion and malaoxon in surface water is 226 and 96.0 ppb, respectively (Table 3). This estimate is based on the maximum application rate for citrus which represents the highest application rate for any crop used to support residue tolerances. EFED notes that higher use rates are reported on product labels but the registrant has stated they will not support rates greater than those defined in crop residue studies.

#### Acute exposure

EFED recommends that 226 and 96.0 ppb be adopted as a highly conservative estimates of *acute* first tier drinking-water exposure for malathion and malaoxon, respectively, based on the peak GENEEC value obtained with use on citrus and cotton.

#### Chronic exposure

EFED recommends that 21.2 and 75.5 ppb be adopted as highly conservative first tier estimates of *chronic* drinking-water exposure for malathion and malaoxon, respectively, based on 56-day average GENEEC concentrations obtained with use on citrus and cotton.

Table 2. GENEEC Environmental Fate Input Parameters (values are for malathion unless otherwise stated.)

DATA INPUT	INPUT VALUE	DATA ASSESSMENT	SOURCE
Application Rate	0.18-6.25 lbs ai/A	confirmed	Recommended usage rates
Maximum Number of Applications	1-25	confirmed	Recommended usage rates
Application Interval	3-30 days	confirmed	Recommended usage rates
Batch Equilibrium (Koc)	151 ml/g	Acceptable	MRID 41345201
Aerobic Soil Metabolism	malathion: $t_{1/2}$ = 3 day malaoxon: $t_{1/2}$ = 3-7 day (model input = 21 days)	Supplemental Supplemental	MRID 41721701 Paschal and Neville 1976
Solubility	145 ppm	Acceptable	Reported by registrant
Aerobic Aquatic Metabolism	$t_{1/2}$ = 3.3 day	Acceptable	MRID 42271601, 43163301
Hydrolysis (used for malaoxon aerobic aquatic $t_{1/2}$ )	$t_{1/2}$ = 104 day		MRID 40941201
Photolysis	$t_{1/2}$ = 94 days	Acceptable	MRID 41673001, 43166301

Table 3. GENEEC EECs ( $\mu\text{g/L}$ ) for certain malathion uses. The lowest and highest malathion use-rates and the use scenario for cotton were analyzed by GENEEC modeling.

rate (lbs ai / A)	crop / interval (days)	application		GENEEC EEC ( $\mu\text{g/L}$ )			
				malathion		malaoxon	
		method	max # annually	peak	56-day ave	peak	56-day ave
0.18	Orange/7 grapefruit/7 lemon/7 lime/7 tangerine/7 tangelo/7 kumquat/7	aerial	10	8.24	0.78	3.10	2.44
0.50	Flax	ground	1	11.4	1.07	1.82	1.43
2.5	Cotton/3	ground	25	181	16.9	96.0	75.5
5.0	Pineapple/7	ground	3	224	20.9	47.3	37.2
5.0	Chestnut/7	ground	4	225	21.1	57.1	44.9
6.25	Orange/30 grapefruit/30 lemon/30 lime/30 tangerine/30 tangelo/30	ground	3	226	21.2	37.1	29.2

---

EFED notes that there is limited information available on the conversion of malathion to malaoxon during drinking water treatment. In a limited sampling of water entering and leaving a water treatment plant both malathion and malaoxon levels generally decreased after treatment, however, one sample showed an increase in malaoxon (USDA 1997). Data from sampling and analysis with a lower detection limit show a much higher rate of conversion (summarized further in the second tier assessment and Table )(personal communication, Dr. Marion Fuller, Florida Department of Agriculture and Consumer Services). EFED recognizes that conversion of malathion to malaoxon may be more efficient during water treatment than under conditions in the field, thus malaoxon may be present at a much higher concentration relative to malathion after water processing.

### **Ground Water Assessment**

As EFED noted above, malathion has some mobility characteristics similar to other chemicals that have been detected in ground water. In addition, malathion has been detected in ground water in at levels ranging from 0.03 to 6.17 ppb in California (1 detection out of 499 wells sampled at a concentration of 0.32 ppb), Mississippi (2 detection out of 263 wells sampled at a range of concentrations of 0.03-0.053 ppb) and Virginia (9 detections out of 138 well sampled at a range of concentration of 0.007-6.17 ppb ); as reported in the EPA/OPP/EFED/EFGWB EPA Pesticides in Ground Water Data Base 1971-1991, National Summary. ERB1/EFED believes that malathion has the potential for movement into groundwater, especially on soils with low organic matter and high sand content.

Cheminova disputes the ground-water data reported in the PGWDB. In particular, it calls into question the analytical methods used to generate the data in the Virginia study. In addition, Cheminova indicates that the maximum detection in the study was 3.12 ppb, not 6.17 ppb.

Noting Cheminova's doubts for the Virginia data, EFED suggests a ground-water concentration estimate of 3.1 ppb for malathion. This value is more conservative than SCI-GROW modeling results using use parameters for citrus or cotton as stated above. Since this monitoring result is specific for malathion EFED assumes the concentration of malaoxon will not exceed the concentration of malathion. Thus, EFED suggests conservative ground water concentration estimates of 3.1 ppb for malathion and 3.1 ppb for malaoxon.

## APPENDIX 3

### *PRZM-EXAMS inputs*

#### Chemical-Specific Input

Persistence and mobility numbers used in the first-tier GENEEC simulations were also used for the Tier II assessment. Chemical specific input parameters for PRZM and EXAMS are summarized in Tables X and .x. Certain assumptions were made for chemical dissipation parameters included in PRZM 3.1 but not GENEEC;

1. The aerobic soil-metabolism half-life of 3 days ( see following discussion) was used for the adsorbed and dissolved half-life throughout the soil column. Subsoil layers were assumed not to be anaerobic, as the deepest soil column simulated was only 150 cm deep;
2. A foliar decay rate of  $0.126 \text{ d}^{-1}$  was used based on the 90% upper confidence limit of 37 foliar half-lives reported in Willis and McDowell (1987).
3. Volatilization from the soil or foliage were not simulated (set to zero). Registrant submitted data suggest that volatilization is not an important route of dissipation;
4. Dissipation through plant uptake was not simulated;
5. Foliar wash-off of  $0.5 \text{ cm}^{-1}$  was simulated, although data exists showing complete wash-off of organophosphate pesticides with the first 0.1 cm of rainfall. .
6. An application efficiency of 95% was assumed for all application methods. As for GENEEC, drift from aerial applications was assumed to be 5% of the applied mass of malathion, and that from ground or airblast applications was assumed to be 1% of the applied mass.

The aerobic soil half-life for malathion chosen for modeling purposes was 3 days. This value is consistent with that used for USDA modeling in for malathion in the boll weevil eradication program which is the single greatest consumer of malathion. Degradation rates in soils vary greatly from the registrant supported half-life of 0.2 days to 11 days in rangeland soil with low organic content.<sup>4</sup> Open literature values are mostly greater than those observed in the acceptable submitted aerobic soil metabolism study. However, because the conditions and parameters controlled in the different open literature studies vary greatly it is not possible to calculate an upper 90th percentile limit of the values. In this instance, multiplying the registrant's submitted half-life value of 0.2 days by three to estimate the 90th percentile upper confidence limit did not produce a conservative value relative to published literature (Table 3). Using a single half-life value for modeling multiple scenarios is clearly a simplification in this instance but it is necessary to choose a value that is a conservative estimate of malathion degradation in

---

<sup>4</sup> Buckman , H.O. and Brady, N.C., 1969. The Nature and Properties of Soils. Macmillian Company, Collier-Macmillian Limited, London as referenced in USDA/APHIS National Boll Weevil Cooperative Control Program. Final Environmental Impact Statement Volume 1, 1991.

agricultural settings used in modeling. The 3 day half-life chosen is not the highest available value published but conditions favoring very long persistence (*ie* very low moisture levels and microbial counts) are not expected to commonly occur in agricultural settings.

PRZM and EXAMS require that degradation half-lives be converted into rate constants. The aerobic soil metabolism half-life of 3 days (as explained above) was converted to a daily rate constant for PRZM 3.1 by the equation  $\text{Ln } 2 / (T_{1/2})$ . The aerobic aquatic (input variable KBACW), anaerobic aquatic (KABCS), and photolysis (KDP) half-lives for EXAMS were converted to hourly rate constants using the formula  $\text{Ln } 2 / (T_{1/2} \times 24)$ . Hydrolysis half-lives at pH 7 (KNH) and pH 9 (KBH) were converted to rate constants by solving two simultaneous equations with the stable pH5 (KAH) constant set to zero.

Table 3. PRZM 3.1 input parameters for Malathion		
Input Parameter	Value	Quality of Data
Foliar Volatilization (PLVKRT)	0 d <sup>-1</sup>	Poor
Foliar Decay Rate (PLDKRT)	0.126 d <sup>-1</sup>	Supplemental
Foliar Wash-off Extraction Coefficient (FEXTRC)	0.5 cm <sup>-1</sup>	Poor
Plant Uptake Fraction (UPTKF)	0	Poor
Partition Coefficient (Koc) for all crops	151 L kg <sup>-1</sup>	Acceptable
Dissolved Phase Decay Rate: All Horizons (DWRATE)	0.231 d <sup>-1</sup>	Fair
Adsorbed Phase Decay Rate: All Horizons (DSRATE)	0.231 d <sup>-1</sup>	Fair
Vapor Phase Decay Rate (DGRATE) (all horizons)	0 d <sup>-1</sup>	Poor

Table 4. EXAMS Input parameters for Malathion	
Input Parameter	Value
Aerobic Aqueous Metabolism Constant (KBACW)	8.82 x 10 <sup>-3</sup> h <sup>-1</sup>
Sediment Metabolism Constant (KBACS)	3.78 x 10 <sup>-3</sup> h <sup>-1</sup>
Acid Hydrolysis Rate Constant (KAH)	0 h <sup>-1</sup>
Neutral Hydrolysis Rate Constant (KNH)	4.10 x 10 <sup>-3</sup> h <sup>-1</sup>
Alkaline Hydrolysis Rate Constant (KBH)	5.46 x 10 <sup>3</sup> h <sup>-1</sup>
Photolysis Rate Constant (KDP)	2.95 x 10 <sup>-4</sup> h <sup>-1</sup>
Partition Coefficient (KOC) for all modeled crops	151
Molecular Mass (MWT)	330 g · mol <sup>-1</sup>
Solubility (SOL)	145 ppm
Henry's Law Constant (HENRY)	0
Q10 For The water Column (QTBAW)	2
Q10 For Sediment (QTBAS)	2

Crop-Specific Inputs

Cotton

This input file was adapted from EFED's standard PRZM scenario for cotton grown on the Loring silt loam in Mississippi, dated April 10, 1998. This soil is located in Major Land Use Area (MLRA) 134. However, weather data from Major Land Resource Area (MLRA) 131 is suggested for this standard scenario, as it represents a closer weather station (Jackson, MS). Inputted modeling parameters are as follows:

Crop	Planting Dates	Harvest Dates	Application Dates	Application Method
Cotton	May 1	Sept. 22	June 1 - August 12	Aerial

Local dates for planting and harvesting cotton, and likely dates of malathion application, were from USDA Boll Weevil Eradication reports. This PRZM simulation reflects the maximum label rate (2.5 lb ai/a), number of applications (25/year) and application interval (3 days) sought by the registrants for methyl parathion on cotton. USDA notes that these usage parameters are extreme and only 2 of 1000 fields treated in the Boll Weevil Eradication Program received 25 applications. Quantitative usage statistics show the average application rate of malathion on cotton is 0.3 lbs ai / A.

Sorghum

This input file was adapted from EFED's standard PRZM scenario for sorghum in Kansas grown on a Loring silt loam soil. The weather recorded for MLRA 112 was used for meteorological input. Leroy Brooks of Kansas Agricultural Extension provided information that malathion would be most likely used early in the season to control aphids and greenbugs. Typical application months would be May through July. Inputted modeling parameters are as follows:

Crop	Planting Dates	Harvest Dates	Application Dates	Application Method
Sorghum	May 21	October 1	June 1-15	aerial

Apple

Malathion is used in orchards in the Northwest including apples, cherries, pears and walnuts. The largest use is on cherries and apples but results from these scenarios are expected to similar to each other and other orchard crops. Franz Needleholter of Oregon State Agricultural Extension provided typical malathion application dates for northern Oregon cherries.

Applications may begin in the end of May and typically end the beginning of August. The most intense period of malathion usage is in June. Inputted modeling parameters are as follows:

Crop	Bloom	Harvest Dates	Application Dates	Application Method
Apples	April 1	December 15	June 1-29	air blast

Citrus

A standard Florida citrus scenario was chosen to model malathion on this crop. Although more malathion is used annually in California it is expected that the Florida scenario would result in higher estimated environmental concentration because of the weather, agricultural practices, and soils. Andy Rose of the University of Florida Agricultural Extension provided information that insecticides may be used throughout the year in citrus agricultural but use may be highest in the summer months. Modeling parameters were as follows:

Crop	Bloom	Harvest Dates	Application Dates	Application Method
Citrus	May 11	August 1	June 1 - July 31	air blast

Lettuce

More than 14 California vegetable crops receive malathion treatments. The California lettuce scenario adapted from a cole crop scenario is expected to have similar results to several other California crops. Dr. Bill Chaney in Salinas with Agricultural Extension provided very useful information on lettuce grown in central California. Lettuce may be planted anytime from January through August and harvested anytime from April through November. No lettuce is grown in December to break lettuce mosaic virus life cycle. Insecticides are applied generally starting April 1. Modeling parameters were as follows:

Crop	Planting Dates	Harvest Dates	Application Dates	Application Method
Lettuce	February 10	May 12	April 1-26	aerial



## APPENDIX 4

### **Other Aquatic Monitoring Studies**

#### **Environmental Monitoring for Malathion Residues in Selected APHIS (PPQ) Control/Eradication Programs. USDA, 1990.**

##### (A) Florida Medfly Program

Methods: Water analysis of residue levels in canals, lakes, swimming pools located with 1/4 miles of spray sites in Florida. Samples were taken 24 hours prior to and immediately after malathion sprays (15 minutes) - 1985 - 24 samples taken. 1987 - 42 water samples. 1990 - 62 water samples taken.

##### Results: 15 minutes post application

1985 - 6 of 24 samples showed Malathion residues 0.2 - 2 ppb

1987 - 9 of 42 samples - Residue levels - 0.23 - 1.55 ppb

1990 - 56 of 62 samples Residue levels - mean 0.8 ppb 0.23 - 51 ppb  
mean average fluctuation 6 - 18 ppb - mean 9.4 ppb

After 48 hours residues degraded rapidly - most below detectable limits.

##### (B) Grasshopper Control Programs

These monitoring programs were conducted in 13 states from 1984 through 1989.

Methods: Malathion applied at 8 oz ULV/Acre; Purity was 91% ai. Samples obtained from flowing or impounded waters, natural or man made and standing water within 1/4 mile of spray zone or in the spray zone. Samples collected 24 hours prior to application and 15 minutes after application. Some sites - daily sample up to 72 hours post application. Level of detection = 0.1 ppb.

Results: Residue ranges were from 0.11 to 85.11 ppb - Highest residues in Utah and Wyoming. Two day samples had residues of 0.3 - 18 PPB.

© Bollweevil Control Program: Monitoring was conducted in Alabama, Arizona, California, Florida, Georgia, New Mexico, Mississippi, N. Carolina, So. Carolina and Texas during the 1985, 1988, 1989 and 1990 cotton growth seasons.

Methods: Application rate was 12 oz./Acre of 93-96% malathion using ULV. Interval was as little as one week for multiple applications. Soil, water and vegetation samples were collected. Water samples were take from area within 1/4 mile of application sites. These included recreational areas, houses, buildings, endangered species habitat, cotton fields (multi-applications) and pond sample following rainfall which occurred 3 days post-app. Where runoff occurred daily samples were collected for 3-5 days depending on how soon after application rainfall occurred.

Results: In Alabama 48 of 82 samples had Malathion residues over 0.1 ppb. These ranged from 0.102 to 24.74 ppb. Only 1 site was over 20 ppb (all samples). In Florida 15 samples were taken with 8 having residues ranging from 6 to 48.6 PPB (the high Malathion residues less than 6 PPB. In Georgia post application mean average residues after 12 applications were 12.9 PPB (15 minutes), 5.18 PPB (1-5 days) 1.78 PPB (6-10 days) and 1.86 (11-71 days.) Testing to determine whether distance or amount of rainfall was a more important factor indicated that distance from the water was more influential in effecting residue levels.

**Pesticide Residues in Hale County, Texas, Before and After Ultra-Low Volume Aerial Application of Malathion. Guerrant, G.O. et al Pesticides Monitoring Journal, 1970.**

Methods: Mosquito control June - August 1967 near Plainview and Abernathy Texas. Applied at 3.0 fluid oz/Acre by Air Force Tactical Air Command C 123 cargo planes based with 4500th Air Base Wing in Langley, AFB, Va. Altitude: 150 feet. Speed 150 mph. Winds <10 mph. Filter papers used to collect residues.

Results: Average concentration following 20 applications on June 16-22 was 1.5 MG/FT<sup>2</sup> or 65% of application rate. Hydrolysis studies of water samples stored in various PH solutions up to 20 days after treatment showed residue stability at pH 2 and 72-62% recovery in pH 4-6. At pH 7 and 8, 16% and 11% recovery were observed. Residues in 32 sites (field water) showed malathion residues of 0.00 to 0.50 ppm 4 hours post application which decreased to < 0.006 within 24 hours post application.

**Residue Monitoring Report submitted to the Agency as 6a2 information under Barcode D207975.**

Methods: Surface water residue monitoring data from waters adjacent to California rice fields included bensulfuron methyl, molinate, thiobencarb, carbofuran, methyl parathion, and malathion monitoring results.

Results: For malathion the maximum reported residues were 0.17 ppb.

**Malathion: Special Projects Report No. 84-9SP. Cornacchia, John W., 1984. A report to the California State Water Resources Control Board, Toxic Substances Control Program.** This report not only summarizes fate and toxicity characteristics for the Board, but also reports on findings of the 1981 monitoring program carried out during Medfly eradication efforts in Santa Clara, Santa Cruz, and San Mateo counties.

Spray Methods: Aerial spraying during spring, summer and fall months. 6 or more applications were made per site at a rate of 2.4 oz. Cythion/acre as a malathion laced bait. Helicopter and fixed wing aircraft were used.

Monitoring Program Methods: Monitoring stations were concentrated in aquatic areas based on the value of fisheries resources supported by the watershed. Creeks and rivers were sampled 24 hours prior to spraying to establish background levels of contaminants. Post application samples were taken within 72 hours of spraying in the area. These sample scenarios were taken every 1-2 weeks during the spraying program. Some sediment and biota samples were also taken for

analysis of malathion/malaoxon residues. Some of the creeks were sampled daily. Fish kills were monitored more closely and fish tissues were analyzed as well as the water from the location of the kill. Population estimates were also made before and after spraying in the San Lorenzo River drainage area. Additional monitoring of storm water drains and urban creeks was conducted immediately following rainfall events during the spraying program. The spray zones ranged in size from 43 to 1264 square miles.

Brief Summary of Results:

Santa Clara Valley: Air samples following the spraying did not exceed 1 ug/m<sup>3</sup>. Average coverage efficiency was approximately 76% with an estimated 24% loss attributed to off target spray drift. Monitoring was carried out up through the 6th application only. Dissipation from teflon coated collection surfaces indicated a half-life of 2.96 days for malathion residues. Peak summer water sample residue levels in two creeks ranged from 0 to 152 ppb. Based on sampling efforts aquatic half lives from local creeks were calculated to range from 1.71 to 6.97 days with less than 5% remaining after 1 week. One creek actually showed a slight increase in residue levels following applications 3, 4, and 5. During fall spraying efforts more frequent rainfall events led to levels as high as 800 ppb in Adobe Creek. This sample level coincided with a fish kill in the creek. In one instance a spraying effort was continued during a rain event and sampling showed residue levels as high as 1000 ppb, also leading to a fish kill. In general fall monitoring of residue levels averaging below 30 ppb within 48 hours after spraying. Levels were elevated when rain events occurred as late as 6 days after application. Storm drains acted as point source discharges for concentrated residues into tributaries.

The first fish kill primarily involved sticklebacks, a sensitive creek species. Residue levels at the time of the kill were 81 ppb. Body tissue concentrations of 3.8 to 1.6 mg/Kg were measured. One sediment sample contained 21 ug/L of malathion, but all others fell below 10 ug/L. In one instance residue levels reached 15 ppb in a lagoon and estuarine trough receiving water from spray area tributaries after a storm event and a fish kill involving striped bass and starry flounders was recorded.

Santa Cruz County: Spraying was carried out from August to mid November. Sampling along the San Lorenzo River and it's tributaries produced residue levels of <0.1 ppb to 41 ppb. Half-life estimates based on these sample periods ranged from 1.67-3.67 days. In general, levels dropped to below 1.0 ppb within a week after application. Rainfall events within 36-72 hours after spraying produced 11 to 19% increases in recorded malathion residues.

San Mateo County: During a storm event malathion residues reached 103 ppb in Pascadero Creek. steelhead trout and stickleback fish kills were reported following the event.

**Malathion Residue Detections in Field Studies for Mosquito Control Uses**

Description/Location	Application Data	Detected Residue Range
----------------------	------------------	------------------------

NW Florida near Pensacola Beach, Tagatz, M.E. et al, 1974 Mosquito News test site was saltmarsh plot measuring 85 x 115 ft and control site was similar size. A 10' wide 4' deep canal bordered marsh.	Thermal Fog-95% Technical in fuel oil-applied at 17500 cu. Ft/minute-applied 3 times with two week intervals at low tide.	Marshwater 6 hrs post applic.=5.2 ppb 12 hrs post applic.=<0.1 ppb Canal Water 6 hrs post applic.=0.42 ppb 12 hrs post applic.=<0.1 ppb
Same study as above but applied to different sites=8.5 acre marsh -25 liter tubs also distributed in marsh	ULV application- 3 applications- truck mounted-330 foot swath at rate of 0.64 fl. oz./acre.	After 3rd application: Tubwater: 1 hr.=0.32 -1.52 ppb 6 hr=>0.5 - 0.58 ppb Marshwater: <0.05 ppb(ND)
Texas-near West Galveston. Conte F.S. and J.C. Parker, 1975- applied to 3 bayous and saline lake in a saltmarsh. Depth was 61-91 cm.	Aerial application at 85.7 g/hectare-air-speed 145 km/hr at 9.2 meter altitude.	Post application residues Test I                      Test II and III 9 hrs:3.0 ppb            1 hr:2.0 - 2.5 ppb 24 hrs: 1.5 ppb        3 hr.: 2.0 - 3.2 ppb 33 hrs: 1.0 ppb        8 hr.: 1.5 - 2.4 ppb 48 hrs: 0.8 ppb        24 hr: 1.2 - 2.2 ppb
Texas-Galveston, Proctor, R.R. J.P. Corliss, and D.V. Lightner, NMFS, Gulf Coastal Fisheries Center	Aerial application at 77.8 ml of 95% malathion/acre. Tests were conducted twice at different times for same test site-two weeks apart.	Measured residues post application Test 1- High tide    Test 2 Low Tide 0 hr= 8.9 ppb            65.3 ppb 6 hr=7.0 ppb            69.0 ppb 24 hr=3.1 ppb            1.08 ppb 48 hr=0.5 ppb            0.05 ppb

During the 1981 aerial application program to control medfly outbreaks the California Dept. Fish and Game documented the environmental effects of malathion laced protein bait applications on 23 inland streams during the first 10 spray sequences of the program. Application rate was 196 g/ha. First application was July 14, 1981. One liter samples were collected in amber glass bottles at 0.3 m. depth, closed under water (to prevent air pockets), placed on ice, chilled to 4 deg. C, and transported to the Dept. Of Food and Agriculture's Meadowview laboratory in Sacramento for analysis of malathion residues. Samples were taken within 24 hours before application or within 12 hours after application.

"The aerial applications produced noticeable pulses of malathion concentrations in the streams. The peak malathion concentrations generally diminished to previous pre-spray concentrations before the next application."

"The highest malathion concentrations recorded in streams were those during or immediately following rainstorms. On October 7, 1981, 800 ug/L malathion was recorded at Station 4.1 during a major rainstorm. In Stevens Creek (Station 8.2), the effect of rain runoff on malathion concentrations was dramatic; previously recorded post-spray levels averaged 4.4 ug/liter but increased to 159 ug/liter on October 27, 1981 immediately after a rainstorm. High concentrations of malaoxon were also seen in these water samples. At station 4.5A, malaoxon concentrations exceeded the malathion concentrations."

148

**San Francisco Monitoring Program-Stream Samples Mean Monitored Levels in PPB**

Station # and Location	Dates Monitored	Pre Application Mean(range)	Post application Mean(range)
1.1 Permanente Creek	7/14-8/10	0.3(0.2-0.5)	8.3(0.7-19.0)
1.2 Stevens Creek(El Camino)	7/14-8/9	2.9(1.7-4.0)	40.9(0-82.6)
2.1 Saratoga Creek	7/16-8/9	0	0.2(0.2-0.2)
3.1 San Tomas Aquino Creek	7/15-8/27	2.2(0.9-5.1)	57.4(0.3-157.0)
4.1 Los Gatos Creek	7/16-8/5	0.5(0-1.3)	2.5(0.6-5.4)
4.2 Ross Creek	7/16-8/5	2.2(0.1-4.3)	27.1(6.2-50)
4.3 Coyote Creek(Oakland Rd)	7/16-8/26	0.8(0.2-2.1)	3.1(0.4-10)
5.1 Coyote Creek(Kelly Park)	7/15-8/6	0.4(0.2-0.6)	1.0(0.3-1.6)
7.1 Belmont Creek	8/11-9/16	0.4(0.2-0.6)	20.1(2.8-81.1)
8.1 Adobe Creek	7/15-8/6	1.5(0-4.3)	19.6(1.0-54.0)
8.2 Stevens Creek(S.Creek Blvd)	7/15-9/24	0.3(0-1.5)	4.1(1.2-8.8)
8.3 Los Trancos Creek	7/19-8/26	0.1(0-0.2)	2.5(0.7-5.3)
10.1 Llagas Creek	8/13-9/22	0.3(0-0.5)	1.3(0-3.8)
10.2 Uvas Creek	8/13-9/22	0.5(0-1.4)	1.1(0-2.6)
12.1 San Lorenzo River(Felton)	8/15-9/22	0.3(0-0.5)	6.2(0-15.9)
12.2 San Lorenzo R. (Brookdale)	8/15-9/21	0.2(0.1-0.3)	6.4(0.7-17.0)
12.3 Loch Lomond Reservoir	8/24-9/21	0.1(0-0.1)	4.2(0.1-12.5)
12.4 Bear Creek	8/15-9/22	0.3(0.2-0.4)	12.5(1.0-39.0)
12.5 Boulder Creek	8/15-9/21	0.1(0-0.3)	10.2(0.2-41.0)
12.6 San Lorenzo R.(Saratoga Rd)	8/15-9/21	0.1(0-1.4)	0.3(0-0.6)
13.1 Coyote Creek(Metcalf Rd)	8/12-9/24	0(0-0.3)	0.5(0.1-1.0)
15.1 Alameda Creek	8/27-9/25	0.1(0-0.3)	1.6(0.4-2.9)
16.1 Pajaro River(Hwy 101)	9/1-9/24	0.7(0.4-1.1)	5.6(0.3-11.5)

**Malathion /Malaoxon Concentrations Up and Down Stream of 4 Drainage Culverts**

Station #	October 27 Malathion/Malaoxon ug/L	November 12 Malathion/Malaoxon ug/L
1.3A Upstream At Drain Downstream	449.5 / 164.5 569 / 384 361.5 / 169	328.2 / 30.4 231.2 / 21.2 253.2 / 22.1

4.4 A Upstream	2.0 / 0.8	38.6 / 26
At Drain	142 / 147	37.5 / 13.5
Downstream	23.5 / 22	37.9 / 26.9
4.5 A Upstream	137 / 212.5	32.0 / 18.4
At Drain	188.5 / 250	50.0 / 13.8
Downstream	169.5 / 231	37.3 / 19.3
1.5 A Upstream	159.0 / 68.0	52.3 / 14.8
At Drain	434.0 / 166.5	292 / 57.5
Downstream	156.5 / 68.0	62.5 / 14.8

Six different locations(sampling stations) in the San Francisco Bay estuary were monitored during the 1981 medfly eradication program. The samples represented prespray levels, levels after spraying during the dry season(July 15-Sept. 24), and during the rainy season Oct. 14-Dec. 13). No detections were recorded prior to spraying. Rainy season detections ranged from mean averages of 1.0 to 7.0 ppb whereas dry season detections were much lower and sometimes considered non-detects(below 0.1 ppb f detection). The table below summarizes dry and rainy season detection ranges and means for the six sample stations.

**San Francisco Bay Estuary- Malathion Monitoring Results Dry and Rainy Seasons 1981**

Station #	Monitoring Dates	Mean Avg.	Range
SF.1 Coyote Creek RR Bridge	7/20 to 9/24(dry season) 10/14 to 12/13(wet season)	0.1 1.7	0.0-0.25 0.0-5.3
SF. 2 Alviso Slough at South Bay	7/20 to 9/24(dry season) 10/14 to 12/13(wet season)	0.1 1.4	0.0-0.25 0.0-3.3
SF. 3 Guadalupe Slough at South Bay	7/20 to 9/24(dry season) 10/14 to 12/13(wet season)	0.1 1.7	0.0-0.40 0.0-3.5
SF. 4 Stevens Creek at South Bay	7/20 to 8/7(dry season) 11/3 to 12/13(wet season)	N.D. 1.0	N.D. 0.0-2.0
SF. 5 Palo Alto Yacht Harbor at buoy	7/20 to 8/7(dry season) No samples (wet season)	N.D. No samples	N.D. No samples
SF.6 Coyote Creek at Calaveras Pt.	7/20 to 8/7(dry season) 11/3 to 12/13	N.D. 1.0	N.D. 0.0-2.0
SF.7 Guadalupe Slough midway point	dry season-no samples 11/3 to 12/3	No samples 7.0	No samples 0.0 - 18.0
SF. 8 Alviso Slough midway point	dry season-no samples 11/3 to 12/13(wet season)	No samples 7.0	No samples 0.0-16.0

(Impact on Fish and Wildlife From Broad scale aerial Malathion Applications in San Francisco Bay Region, 1981, State of California Resources Agency, Dept. of Fish and Game , Environmental Services Branch, Administrative Report 82-2)

**Office of Water - Red Book 1976 Jeff, G.**

**Malathion Sensitivity of Freshwater Fish (Salmonids (4), centrarchids (3))**

\* Mentions Estimated pH-6 half-life = 5 months. At pH-8 estimated half-life = 2 wks (Weiss and Gakstatter, 1964)

\*In Miami River monitoring 10% of original residues remained in Little Miami River (pH 7.3 - 8.0) after 2 wks (Eichelberger and Lichtenberg, 1971).

**Environmental Monitoring Results of the Mediterranean Fruit Fly Eradication Program, Ventura County, 1994-1995. A.Bradley, P.Woolford, R.Gallavan, P.Lee, J. Troiano. State of California Environmental Protection Agency, Dept. Of Pesticide Regulation. EH-97-05, December 1997.**

Methods: Air, soil surface and water residues were collected for analysis of malathion and malaoxon deposition levels following aerial application of malathion ULV and Nu-Lure bait mixture at an application rate of 102 gms malathion+ 789 ml of Nu-Lure per hectare. A 41 square kilometer area was dosed 14 times at night by helicopters flying at 130 km/hr 100 meters above the ground and equipped with 6 Tee Jet flat fan nozzles resulting in minimum swath width of 61 meters. Intervals ranged from 14 to 21 days between applications. Ground deposition to soil was recorded on 1 foot square absorbent material at 34 sites which were collected 30 minutes after application. Air samples were taken at 5 sites prior to, during and after applications were made. Water samples were collected at Conejo Creek at one site before entry of stream into the treated area and at a location where the stream exited the treatment area. These samples were stabilized by reducing pH to 3 by addition of hydrochloric acid. In addition, stream samples were collected following 3 heavy rainfall events on Nov. 7, Jan. 20, and March 21. These rainfalls occurred 3 to 12 days after the last application. Due to high readings of malathion residues in runoff after the first two rainfalls several additional sites further downstream and in an estuary 15 km away from the application area were sampled after the third event. In addition bioassays were performed on cladoceran and mysid species using stream samples following the Jan. 20 and March 21 events.

Reported Results: Deposition ranged from 80% (first 2 sprays) to 30% (third spray) of theoretically expected rate based on application rate computations. Ground deposition ranged from non-detectable to 0.07 mg/m<sup>2</sup>. The highest average air sample level recorded was 5.0 ppt (0.067 ug/m<sup>3</sup>) during the 24 hour period following a spray. Water samples collected at the inflow site for Callegus Creek before application averaged 0.09 ppb of malathion and 0.04 ppb of malaoxon. Following application the outflow sample site malathion residues averaged 44 ppb (range=39-50 ppb). During the rainfall events the highest recorded level of malathion was 787 ppb and the highest malaoxon level was 160 ppb (12 days post application). The estuarine collection site (Mugu Lagoon) was sampled when a rainfall occurred six days after an application. The resulting residue concentrations were negligible during the storm, but rose to 11.2 ppb of malathion and 2.62 ppb of malaoxon 4.5 hours later. Fifteen hours later malathion had decreased to 0.16 ppb and malaoxon was not detected. No fish kills were reported, despite the high residue levels. However, bioassays performed with stream samples and *Ceriodaphnia dubia* and with estuarine samples and *Neomysis mercedis* produced 100% mortality within 2 and 24 hours, respectively.

### SAMPLE TERESTRIAL EEC OUTPUT FILES-MALATHION

RUN No. 1 FOR Malathion ON Sweet Corn \*INPUT VALUES\*

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.610( .813) 5 5 AERIAL\_A 28.1 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT GRASS TEEC	TALL GRASS TEEC	BROAD LEAF TEEC	SEED FRUIT TEEC
------------------------	-----------------------	-----------------------	-----------------------

195.01(69.07) 89.38(29.25) 109.69(36.56) 12.19( 5.69)

RUN No. 2 FOR Malathion ON Beans \*INPUT VALUES\*

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.610( .710) 3 7 AERIAL\_A 28.1 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT GRASS TEEC	TALL GRASS TEEC	BROAD LEAF TEEC	SEED FRUIT TEEC
------------------------	-----------------------	-----------------------	-----------------------

170.44(60.36) 78.12(25.57) 95.87(31.96) 10.65( 4.97)

RUN No. 3 FOR Malathion ON Alfalfa \*INPUT VALUES\*

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.610( .623) 2 14 AERIAL\_A 28.1 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT GRASS TEEC	TALL GRASS TEEC	BROAD LEAF TEEC	SEED FRUIT TEEC
------------------------	-----------------------	-----------------------	-----------------------

149.42(52.92) 68.48(22.41) 84.05(28.02) 9.34( 4.36)

RUN No. 4 FOR Malathion ON Blueberry \*INPUT VALUES\*

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.760( .811) 5 10 AERIAL\_A 28.1 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT GRASS TEEC	TALL GRASS TEEC	BROAD LEAF TEEC	SEED FRUIT TEEC
------------------------	-----------------------	-----------------------	-----------------------

194.56(68.91) 89.17(29.18) 109.44(36.48) 12.16( 5.67)

052



**RUN No. 5 FOR Mushroom ON Malathion \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.940( 1.605) 4 3 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

385.15(136.41) 176.53(57.77) 216.64(72.21) 24.07(11.23)

**RUN No. 6 FOR Malathion ON Strawberry \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.940( 1.160) 6 6 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

278.32(98.57) 127.56(41.75) 156.56(52.19) 17.40(8.12)

**RUN No. 7 FOR Malathion ON Peppermint \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.940( 1.094) 3 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

262.64(93.02) 120.38(39.40) 147.74(49.25) 16.42(7.66)

**RUN No. 8 FOR Malathion ON Macadamia \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.940( 1.098) 7 7 GROUND\_A .0 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

263.42(93.30) 120.74(39.51) 148.18(49.39) 16.46(7.68)

**RUN No. 9 FOR Malathion ON Macadamia \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

.940( 1.098) 7 7 AERIAL\_B 16.4 NO(100%)

153

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

---

263.42(93.30) 120.74(39.51) 148.18(49.39) 16.46( 7.68)

**RUN No. 10 FOR Malathion ON Melons/Squ \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)

---

1.000( 1.168) 6 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

---

280.24(99.25) 128.44(42.04) 157.63(52.54) 17.51( 8.17)

**RUN No. 11 FOR Malathion ON Corn \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)

---

1.250( 1.794) 2 3 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

---

430.58(152.50) 197.35(64.59) 242.20(80.73) 26.91(12.56)

**RUN No. 12 FOR Malathion ON Mustards \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)

---

1.250( 2.198) 6 3 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

---

527.62(186.87) 241.83(79.14) 296.79(98.93) 32.98(15.39)

**RUN No. 13 FOR Sorghum ON Malathion \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)

---

1.250( 1.455) 3 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

---

349.26(123.70) 160.08(52.39) 196.46(65.49) 21.83(10.19)

**RUN No. 14 FOR Malathion ON Guava \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.250( 1.460) 12 7 GROUND\_B 6.7 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

350.30(124.06) 160.55( 52.54) 197.04( 65.68) 21.89( 10.22)

**RUN No. 15 FOR malathion ON carrot \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.250( 1.460) 7 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

350.30(124.06) 160.55( 52.54) 197.04( 65.68) 21.89( 10.22)

**RUN No. 16 FOR malathion ON clover \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.250( 1.276) 2 14 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

306.19(108.44) 140.33( 45.93) 172.23( 57.41) 19.14( 8.93)

**RUN No. 17 FOR malathion ON celery \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.500( 1.746) 3 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

419.11(148.44) 192.09( 62.87) 235.75( 78.58) 26.19( 12.22)

**RUN No. 18 FOR malathion ON onion \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?

153

ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.560( 1.821) 5 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

437.15(154.82) 200.36( 65.57) 245.89( 81.96) 27.32( 12.75)

**RUN No. 3 FOR malathion ON blackberry \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.880( 2.194) 4 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

526.62(186.51) 241.37( 78.99) 296.23( 98.74) 32.91( 15.36)

**RUN No. 2 FOR malathion ON strawberry \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.880( 2.195) 6 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

526.84(186.59) 241.47( 79.03) 296.35( 98.78) 32.93( 15.37)

**RUN No. 3 FOR malathion ON grapes \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

1.880( 1.919) 2 14 GROUND\_B 6.7 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

460.50(163.09) 211.06( 69.08) 259.03( 86.34) 28.78( 13.43)

**RUN No. 4 FOR malathion ON raspberry \*INPUT VALUES\***

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

2.030( 2.369) 4 7 GROUND\_A .9 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
-------	------	-------	------

GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC
568.64(201.39)	260.63(85.30)	319.86(106.62)	35.54(16.59)

**RUN No. 5 FOR malathion ON figs \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)
2.500( 3.281)	3 5	GROUND_B	6.7	NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC
<hr/>			
787.50(278.91)	360.94(118.13)	442.97(147.66)	49.22(22.97)

**RUN No. 6 FOR malathion ON peas \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)
2.500( 2.919)	5 7	GROUND_A	.9	NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC
<hr/>			
700.55(248.11)	321.09(105.08)	394.06(131.35)	43.78(20.43)

**RUN No. 7 FOR malathion ON tomato \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)
3.430( 4.569)	5 5	GROUND_A	.9	NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC
<hr/>			
***** (388.35)	502.58(164.48)	616.80(205.60)	68.53(31.98)

**RUN No. 8 FOR malathion ON cotton \*INPUT VALUES\***

RATE (#/AC)	APPLICATIONS	SPRAY	AVG PERCENT	INCRP?
ONE(MULT)	NO.-INTERVAL	TYPE	SPRAY DRIFT	(%SURF)
1.250( 2.213)	25 3	AERIAL_B	16.4	NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC
<hr/>			
531.23(188.14)	243.48(79.68)	298.82(99.61)	33.20(15.49)

**RUN No. 9 FOR malathion ON cotton \*INPUT VALUES\***

057

RATE (#/AC) APPLICATIONS SPRAY AVG PERCENT INCORP?  
ONE(MULT) NO.-INTERVAL TYPE SPRAY DRIFT (%SURF)

-----  
2.500( 4.427) 25 3 AERIAL\_B 16.4 NO(100%)

ON-FIELD FLETCHER RESIDUE LEVELS: MAXIMUM(TYPICAL) IN PPM

SHORT	TALL	BROAD	SEED
GRASS	GRASS	LEAF	FRUIT
TEEC	TEEC	TEEC	TEEC

\*\*\*\*\*(376.29) 486.96(159.37) 597.64(199.21) 66.40( 30.99)